

1 **Influence of red wine pomace seasoning and high-oxygen atmosphere storage on**
2 **carcinogens formation in barbecued beef patties**

3

4 Javier García-Lomillo ^a, Olga Viegas* ^{b,c}, Maria L. Gonzalez-SanJose ^a, Isabel M. P. L. V. O. Ferreira ^b

5 ^a Department of Biotechnology and Food Science, Faculty of Science, University of Burgos, Plaza
6 Misael Bañuelos, 09001, Burgos, Spain.

7 ^b LAQV/REQUIMTE, Laboratório de Bromatologia e Hidrologia, Departamento de Ciências Químicas,
8 Faculdade de Farmácia da Universidade do Porto, 4099-313 Porto, Portugal

9 ^c Faculdade de Ciências da Nutrição e Alimentação da Universidade do Porto, 4200-465 Porto, Portugal

10

11

12

13

14

15 * **Corresponding author:** Laboratório de Bromatologia e Hidrologia, Faculdade de Farmácia da
16 Universidade do Porto, Rua de Jorge Viterbo Ferreira n.º 228, 4050-313 Porto, Portugal

17 E-mail address: olgaviegas@fcna.up.pt (Olga Viegas)

18

19

20 **ABSTRACT**

21 Polycyclic aromatic hydrocarbons (PAHs) and heterocyclic aromatic amines (HAs) are carcinogenic
22 compounds formed in barbecued meat. Conditions that reduce their formation are of major interest.
23 This study aims to evaluate the influence of red wine pomace seasoning (RWPS) and high-oxygen
24 atmosphere storage on the formation of PAHs and HAs in barbecued beef patties. In general, the
25 levels of PAHs and HAs quantified were low. The storage (9 days) promoted higher formation of
26 PAHs in control patties without increase of HAs. RWPS patties cooked at preparation day presented
27 higher levels of PAHs and HAs than control. Nevertheless, RWPS patties cooked after storage
28 presented lower levels of PAHs and HAs than control. ABTS assay pointed out that higher radical
29 scavenging activity may be related to with lower PAHs or HAs formation. In conclusion, RWPS can
30 be an interesting ingredient to inhibit the formation of cooking carcinogens in barbecued patties
31 stored at high-oxygen atmosphere.

32 **KEYWORDS:** polycyclic aromatic hydrocarbons, heterocyclic aromatic amines, red wine pomace,
33 beef patties

34

35 **1. INTRODUCTION**

36 Meat and meat products contain valuable nutrients including proteins, vitamins, iron and zinc. Meat
37 cooking improves the digestibility and prevents microbiological hazards, but also produces
38 carcinogenic chemicals that have drawn the attention of the scientific community. In October 2015,
39 the International Agency for Research on Cancer (IARC) classified “processed meat” and “red meat”
40 as “carcinogenic” and “probably carcinogenic” to humans (groups 1 and 2A), respectively, based on
41 more than 800 epidemiological studies that reported a link between meat consumption and cancer.
42 The potential carcinogenicity of red and processed meat consumption is likely due to a combination
43 of carcinogens present in the meat at the moment of consumption and carcinogens formed in the
44 gastrointestinal tract. Concerning barbecued meat, polycyclic aromatic hydrocarbons (PAHs) and
45 heterocyclic aromatic amines (HAs) were pointed out as two meat components with high
46 carcinogenic potential (Bouvard et al., 2015).

47 PAHs are mainly formed when meat is cooked over an open flame. The mechanisms are not
48 completely understood, but organic matter seems to be fragmented at high temperatures through
49 pyrolysis. Free radicals are produced and can recombine producing polynuclear aromatic compounds
50 by pyrosynthesis. High temperatures (at least 200°C) are required to form relevant amounts of PAHs
51 during cooking process (Chen & Chen, 2001; Sharma, Chan, & Hajaligol, 2006). The incomplete
52 combustion of the heat source (charcoal) may generate low molecular weight PAHs, or “light PAHs”
53 (with 2-3 aromatic rings) and the melted fat that drips from patties to the heat source generates PAHs
54 with more than 3 aromatic rings (“heavy PAHs”). These PAHs are carried up by the smoke to the
55 meat surface (Viegas, Novo, Pinto, Pinho, & Ferreira, 2012).

56 The health risk assessment for PAHs exposure has been addressed by several ways (Yebrá-Pimentel,
57 Fernández-González, Martínez-Carballo, & Simal-Gándara, 2015). However, the EU Scientific
58 Committee on Food established the sum of the following PAHs (PAH8): benzo(a)anthracene (BaA),

59 chrysene (CHR), benzo(b)fluoranthene (BbFA), benzo(k)fluoranthene (BkFA), BaP,
60 dibenzo(a,h)anthracene (DBahA), benzo(g,h,i)perylene (BghiP), and indeno(1,2,3-c,d)pyrene (IP) as
61 the most suitable indicator for the carcinogenic potency of PAHs in food (EFSA, 2008).

62 HAs are another category of carcinogenic compounds formed in cooked meat at high temperature.
63 HAs contain 3 fused aromatic rings, one or more nitrogen atoms in the ring and one exocyclic amino
64 group. HAs are classified as thermic (formed at temperatures between 150 and 250°C) and pyrolytic
65 (formed at temperatures above 250°C). The formation of HAs is associated with reaction between
66 creatinine and Strecker degradation products from Maillard reaction (Skog, Johansson, & Jägerstad,
67 1998).

68 Packaged raw meat is usually stored under refrigerated conditions in high-oxygen atmosphere to
69 keep the bright red color appreciated by consumers. However, those conditions induce oxidative
70 processes, namely depletion of endogenous antioxidants, lipid and protein oxidation and radical
71 accumulation. Recently, it was described that meat cooked after storage presented higher formation
72 of HAs, which was ascribed to the increase of precursors during storage, especially free amino acids
73 (Polak, Andrenšek, Žlender, & Gašperlin, 2009; Szterk et al., 2012; Szterk & Waszkiewicz-Robak,
74 2014). Meanwhile, PAHs formation also involves radical reactions, thus the oxidative status of meat
75 may also affect its formation, although the effect of meat storage on PAHs formations during grilling
76 has not been described.

77 Mitigation strategies to reduce the formation of PAHs and HAs in barbecued meat have been
78 proposed, cooking at lower temperatures, reduction of smoke release and avoid fat dripping (Lee et
79 al., 2016; Skog et al., 1998). Moreover, natural products such as spices and plant extracts that can act
80 as radical scavengers, have been proposed to limit the formation of PAHs and HAs. Recent studies,
81 confirmed that marinating with beer (Viegas, Yebra-Pimentel, Martínez-Carballo, Simal-Gandara, &
82 Ferreira, 2014) or cooking with onion and garlic (Janoszka, 2011) reduced the formation of PAHs,

83 whereas grape seed extract, and wine marinades inhibited HAs formation (Ahn & Grün, 2005;
84 Busquets, Puignou, Galceran, & Skog, 2006; Melo, Viegas, Petisca, Pinho, & Ferreira, 2008; Viegas,
85 Amaro, Ferreira, & Pinho, 2012; Viegas, Moreira, & Ferreira, 2015).

86 Recently, a new seasoning derived from red wine pomace (RWPS) rich in phenolic compounds,
87 mainly flavonoids (Del Pino-García et al., 2016), presented preservative activity by inhibiting
88 microbial growth, lipid and protein oxidation (García-Lomillo, González-SanJosé, Skibsted, &
89 Jongberg, 2016; García-Lomillo, González-SanJosé, Del Pino-García, Rivero-Pérez, & Muñiz-
90 Rodríguez, 2014). The aim of the present work was to evaluate the effect of RWPS on the formation
91 of PAHs and HAs in barbecued beef patties before and after 9 days of storage in high-oxygen
92 atmosphere.

93 **2. MATERIALS AND METHODS**

94 **2.1. Materials**

95 Potassium persulfate ($K_2O_8S_2$) was from Panreac (Barcelona, Spain). 2,2'-Azinobis 3-
96 ethylbenzothiazoline-6-sulfonic acid (ABTS reagent), 6-hydroxy-2,5,7,8-tetramethyl-2-carboxylic acid
97 (Trolox) were obtained from Sigma (St Louis, MO, USA). All the solvents used for PAHs and HAs
98 analysis were of HPLC grade (Merck Darmstadt, Germany) and water was purified using a Milli-Q
99 System (Millipore, Bedford, MA, USA). Hydrochloric acid, ammonium acetate and ammonia
100 solution 25% (v/v) and triethylamine were obtained from Fisher Scientific (Pittsburgh, PA, USA).
101 Extrelut reservoirs and Extrelut HM-N diatomaceous earth refill material were obtained from Merck.
102 The cartridges: Bond Elut PRS (500 mg), Bond Elut C18 (100 and 500 mg) and Mega BE-Si (5 g
103 silica) were obtained from Agilent Technologies (USA). Supelco Visiprep and a Visidry SPE
104 vacuum manifold (Supelco) were used for extraction of PAHs and HAs.

105 The standard mixture containing naphthalene, acenaphthene, acenaphthylene, fluorene,
106 phenanthrene, anthracene (A), fluoranthene (FA), pyrene (PYR), BaA, CHR, BbFA, BkFA, BaP,
107 DBahA, BghiP, and IP was provided by Supelco (Bellefonte, PA, USA). 2-amino-3-
108 methylimidazo[4,5-*f*]quinoline (IQ), 2-amino-3,8-dimethylimidazo[4,5-*f*]quinoxaline (MeIQx), 2-
109 amino-3,4,8-trimethylimidazo[4,5-*f*]quinoxaline (4,8-DiMeIQx), 2-amino-1-methyl-6-
110 phenylimidazo[4,5-*b*]pyridine (PhIP), 3-amino-1,4-dimethyl-5*H*-pyrido[4,3-*b*]indole (Trp-P-1), 3-
111 amino-1-methyl-5*H*-pyrido[4,3-*b*]indole (Trp-P-2), 2-amino-9*H*-pyrido[2,3-*b*]indole (AαC), 2-
112 amino-3-methyl-9*H*-pyrido[2,3-*b*]indole (MeAαC) and 2-amino-6-methyldipyrido[1,2-*a*:3',2'-
113 *d*]imidazole (Glu-P-1) were obtained from Toronto Research Chemicals (North York Ontario,
114 Canada).

115 **2.2. RWPS preparation**

116 RWPS was obtained from dehydrated seedless red wine pomace (González San José, García
117 Lomillo, Del Pino García, Dolores Rivero, & Muñiz Rodríguez, 2015), whose chemical composition
118 as well antimicrobial and antioxidant activities have been previously reported (García-Lomillo et al.,
119 2014). The seasoning was milled (particle size less than 250 µm mesh) and kept in dark until use.

120 **2.3. Patty preparation and cooking**

121 A mixture of different beef cuts, especially sold for the elaboration of patties was obtained from a
122 local supplier (GrosMercat, Burgos, Spain). Doscadesa (Murcia, Spain) provided ingredients and
123 additives used in the formulation (common salt, food grade starch and a commercially available
124 mixture of phosphates).

125 Patties were made by grounding beef and mixing with additives in a food mixer, according to the
126 following formulations: Control patties (920 g of meat, 12 g of starch, 15 g of salt, 3 g of phosphates,
127 and 50 mL of water) and RWPS patties (same formulation than control patties with the addition of

128 RWPS to the final concentration of 2% (w/w, seasoning/patty). Finally, patties were manually
129 formed with a thickness between 12 and 15 mm and a weight between 100 and 105 g. Stored samples
130 were placed in polyethylene/ethylene vinyl alcohol/polystyrene trays (Sanviplast, Barcelona, Spain),
131 filled with gas (70% O₂/30% CO₂), and sealed using a polyethylene terephthalate polyvinylidene
132 chloride/polyethylene (PETPVdC/PE) film and kept in dark at 4°C. Four different groups of patties
133 were formed, Control and RWPS at day 0, which were immediately cooked; and Control and RWPS
134 at day 9, which were stored under refrigeration during 9 days before cooking.

135 Four different patties were made for each four groups, in order to take into account the intrinsic
136 variability of the patties and of the cooking procedures.

137 Patties were cooked in a barbecue using wood charcoal, and the temperature was assessed using a
138 thermometer Crison 638 Pt (Barcelona, Spain). When the temperature was 210°C, samples were
139 placed at 8 cm of distance from the heat source. During patties barbecuing (8 min) the inner
140 temperature of patties was monitored and samples were turned once at 4 min. Charcoal was replaced
141 after cooking each sample.

142 Raw and cooked patties were weighted and the cooking loss was calculated. The four patties of each
143 group were mixed, homogenized and frozen at -80°C. Half of the sample was kept frozen for HAs
144 analysis, whereas the other half was freeze-dried for PAHs analysis.

145 The chemical composition of the raw and cooked beef patties were analyzed using a FoodScan™
146 near-infrared spectrophotometer (Foss Electric A/S, Hillerød, Denmark) and the data processed by
147 the ISIScan™ Software.

148 The study was carried out in duplicate on two different days and from two different batches of beef.
149 Thus, two batches per group, each one composed by four patties were barbecued and analyzed three
150 times.

151 **2.4. ABTS⁺ assay**

152 The radical scavenger activity of raw patties was assessed according to the method described by
153 Rivero-Pérez et al. (2007) adapted to meat samples. The ABTS reagent was prepared by mixing
154 ABTS solution and K₂O₈S₂ in Milli Q water (1:1). 75 ± 2 mg of raw patty were mixed with 15 mL of
155 the solution of ABTS reagent and vortexed. After 30 min of reaction with agitation, the radical
156 scavenger activity was evaluated through the absorbance decrease at 734 nm during 30 min. Standard
157 calibration was conducted using Trolox and results were expressed as µmol of Trolox/ g of patty.

158 **2.5. PAHs extraction and quantification**

159 PAHs extraction was carried out according to Viegas, Novo, Pinho, & Ferreira (2012). A HPLC unit
160 (Jasco, Japan) equipped with one PU-1580 HPLC pump, an AS-950 auto sampler with a 20 µL loop
161 and a FP-920 fluorescence detector were used. The system was controlled by Borwin PDA
162 Controller Software (JMBS Developments, Le Fontanil, France). The column was a C18 Supelcosil
163 LC-PAH (25 cm length; 4.6 mm internal diameter, 5 µm particle size) (Supelco, Bellefonte, PA,
164 USA) thermostated at 32°C. Gradient elution and fluorescence detector excitation/emission program
165 were set up according to Viegas, Novo, Pinho, & Ferreira (2012). Quantification was performed by
166 standard addition method at two fortification levels (10–20 ng/g).

167 **2.6. HAs extraction and quantification**

168 HAs were extracted and purified as described by(Santos et al., 2004). The same HPLC unit (Jasco,
169 Japan) described for PAHs was used, but a diode array detector was also coupled (MD 910
170 Multiwavelength detector) (HPLC-DAD/FLD). The column was a TSK gel ODS80 (Toyo Soda)
171 (5 µm; 250 mm length; 4.6 mm internal diameter). The mobile phase gradient was set as described
172 by Melo, Viegas, Petisca, Pinho, & Ferreira (2008). DAD was set at 263 nm, whereas FLD was set at
173 excitation 307 nm and emission at 370 nm for quantification of PhIP, MeAαC, and AαC.

174 Quantification was performed by standard addition method at two fortification levels (5 and 10 ng/g,
175 for thermic HAs and 10 and 20 ng/g for pyrolytic HAs).

176 **2.7. Statistical analysis**

177 The effect of seasoning treatment on the concentration of PAHs and HAs and the possible
178 interrelation with the storage treatment was evaluated by a linear model with the fixed effects of
179 seasoning, storage, and the seasoning x storage interaction term. Furthermore, least significant
180 difference (LSD) at a *P*-value < 0.05 was applied to determine statistical differences among the
181 levels of PAHs and HAs of the four groups of patties. Statistical analysis was conducted using IBM
182 SPSS Statistics 19.0 (IBM Corporation, Somers, NY, USA).

183 **3. RESULTS AND DISCUSSION**

184 **3.1. Effect of seasoning and storage on total antioxidant capacity of raw patties**

185 Control patties at day 0 presented a radical scavenger activity of 2.98 ± 0.15 $\mu\text{mol/g}$, whereas after
186 the addition of RWPS it decreased to 1.99 ± 0.11 $\mu\text{mol/g}$. Del Pino-García et al. (2015) observed that
187 RWPS presented high radical scavenger activity (120 $\mu\text{mol/g}$ of RWPS). However, RWPS blocks
188 thiol groups of proteins that are known to have a predominant role in the antioxidant activity of meat
189 products (Garcia-Lomillo et al., 2016; Serpen, Gökmen, & Fogliano, 2012), which could explain the
190 decrease in the antioxidant activity of RWPS patties. Furthermore, RWPS may induce protein cross-
191 linking of protein tissue and limit the release of endogenous antioxidants from muscle tissue during
192 the assay (Nirmal & Benjakul, 2011).

193 After 9 days of storage, opposite results were observed. Control samples decreased their radical
194 scavenger activity (1.89 ± 0.17 $\mu\text{mol/g}$), which may be due to the depletion of endogenous
195 antioxidants such as thiols groups or vitamin E during storage. In contrast, patties with RWPS

196 exhibited higher radical scavenger activity ($2.89 \pm 0.16 \mu\text{mol/g}$) than control samples, and very
197 similar to control at 0 day. This could be explained by the release of phenolic compounds from
198 RWPS matrix during the storage and the protective effect of grape polyphenols on the endogenous
199 chain-breaking antioxidants (vitamin E, vitamin C, thiol groups, amine groups or enzymes) (Pazos,
200 González, Gallardo, Torres, & Medina, 2005). Marchiani et al (2016) also observed a gradual release
201 of quercetin during storage of yogurt enriched with grape pomace.

202 **3.2. Patty composition, cooking temperature and cooking loss**

203 Raw patties presented mean values of 64 % of moisture, 18% of protein, 12% of lipid. The
204 incorporation of RWPS did not affect moisture, protein and lipid contents of either raw or cooked
205 patties. Cooked patties presented 57% of moisture, 22% protein and 11% lipid, no significant
206 differences were observed between samples cooked at 0 and 9 days (Table 1). The inner temperature
207 at the end of the cooking was around 83°C and no significant differences were observed in the
208 temperature profile between control and RWPS samples at 0 and 9 days. The cooking loss averaged
209 29.8%, without significant differences between the different groups. The results are within the range
210 reported for beef patties (U.S. Department of Agriculture, 2012), but lower than other products such
211 as beef steaks or pork loin that ranged between 40% and 48% (Viegas, Novo, Pinto, et al., 2012).
212 The use of salt, starch, and phosphates increased the water binding and reduced cooking loss
213 (Sebranek, 2009), and this could explain the low levels of cooking loss observed in the present study.

214 **3.3. Effect of RWPS on PAHs formation in barbecued beef patties before and after storage**

215 The PAH8 (EFSA, 2008), as well as FA, PYR and A were quantified in all samples (Table 2). The
216 levels were low, but within the range reported previously for beef steaks (Viegas, Novo, Pinho, et al.,
217 2012; Viegas, Novo, Pinto, et al., 2012). PAHs formation is related to dripping juices from meat to
218 the charcoal and exposition time, the low cooking losses and short cooking time of patties may
219 explain the low level of PAHs found. Additionally, the fat content is an important parameter for

220 PAHs formation in charcoal grilled muscle foods. Lean beef is known to form lower amount of
221 PAHs compared with pork, chicken and salmon (Viegas, Novo, Pinto, et al., 2012; Viegas et al.,
222 2014). Chung et al. (2011) compared the formation of PAHs in pork and beef, and the PAHs
223 formation in charcoal grilled beef was similar to that obtained in our study, while pork meat
224 produced higher content.

225 Statistical analysis revealed significant effects and interaction between “seasoning” and “storage”
226 factors. At day 0, RWPS patties presented significantly higher values of A, FA, PYR, CHR, BbFA,
227 BaP and IP than control patties (Table 2). The total amount in RWPS patties (\sum PAHs 16.63 ± 0.44
228 ng/g) was also significantly higher than control samples (\sum PAHs 9.67 ± 0.95 ng/g). These results
229 were surprising since antioxidants and spices were reported to mitigate the formation of PAHs
230 (Janoszka, 2011; Viegas et al., 2014). However, RWPS patties at day 0 presented lower scavenging
231 activity against ABTS radical than the corresponding control. Since PAHs formation seems to be
232 mediated by free-radical chain, the lower scavenging activity observed in RWPS patties may explain
233 the higher value of PAHs compared with control. Despite the observed correlation between PAHs
234 formation and ABTS test, it is worth remarking that the mechanisms involved in ABTS^{•+} radical
235 scavenging may be different from those mechanisms taking place during meat cooking. Additionally,
236 the pyrolysis of phenolic compounds may contribute to the higher PAHs measured in RWPS patties.
237 Lignin, cellulose and pectin may render PAHs under pyrolytic conditions (McGrath, Sharma, &
238 Hajaligol, 2001; Sharma & Hajaligol, 2003). RWPS is a complex product with high levels of plant
239 fiber and polyphenols (García-Lomillo et al., 2014) that could explain the higher levels of PAHs
240 measured in RWPS patties at day 0.

241 Control samples cooked after 9 days of storage presented an increase of PAHs content compared
242 with control patties cooked at day 0 (Table 2). Significant differences were found for FA, PYR and
243 BbFA, and for the sum of quantified PAHs (\sum PAHs showed values of 15.78 ng/g and 9.67 ng/g in
244 barbecued patties at 9 and 0 days of storage, respectively). However, RWPS patties cooked after 9

245 days of storage presented similar PAHs levels (Σ PAHs 8.98 ng/g) initially found for control patties.
246 These results could be due to the depletion of endogenous meat antioxidants during storage, which
247 decreased the scavenge capacity against the free radicals of pyrolytic reaction and increased PAHs
248 formation. Furthermore, conjugated dienes, formed during lipid oxidation, may directly undergo
249 dimerization and polymerization leading to the formation of PAHs via Diels-Alder type reactions
250 (Nawar, 1984). No information was found in literature concerning the effect of storage on PAHs
251 formation in barbecued meat, however, Szterk & Waszkiewicz-Robak (2014) demonstrated that raw
252 meat stored for longer periods, formed more HAs during grilling, probably due to the higher content
253 of free amino acids. RWPS patties grilled after 9 days of storage presented lower levels of PAHs
254 than stored control patties, although significant reduction (between 28 and 57 %) was observed only
255 for FA, PYR, BaA, and CHR. RWPS patties cooked at 9 days presented lower levels of A, FA, PYR,
256 BaA, CHR and IP than RWPS patties cooked at 0 days. These results may indicate that the action of
257 polyphenols was not immediate and then some time is required to exert their protective effect. This is
258 in agreement with the ABTS values of RWPS patties that increased from 1.99 $\mu\text{mol/g}$ to 2.89 $\mu\text{mol/g}$
259 after 9 days of storage. Furthermore, the protection exerted by RWPS to oxidation may contribute to
260 the lower PAHs formation. Previous works have already pointed out the capacity of RWPS to retard
261 the formation of products derived from lipid oxidation, and the loss of endogenous thiols groups
262 (with antioxidant activity) in meat proteins during the storage at high-oxygen atmosphere (García-
263 Lomillo et al., 2016; García-Lomillo et al., 2014).

264 **3.4. Effect of RWPS on HAs formation in barbecued beef patties before and after storage**

265 Among the nine HAs evaluated, only two (PhIP and A α C) were detected above the limit of detection
266 in all samples. Furthermore, MeA α C was also detected in patties cooked with RWPS at day 0 and
267 control samples after 9 days of storage (Table 3). The levels of PhIP and A α C found were within the
268 range reported in beef samples (Szterk et al., 2012). The low formation of HAs in the present study is

269 in agreement with the fact that beef forms lower amounts of HAs than other muscles foods such as
270 chicken or salmon (Viegas, Novo, Pinto, et al., 2012). Additionally, the low cooking loss limited the
271 transfer of precursors to the patty surface where HAs are formed (Skog et al., 1998). The use of
272 starch, salt and phosphate reduced the transport of precursors towards patties surface during cooking
273 (Borgen & Skog, 2004; Persson, Graziani, Ferracane, Fogliano, & Skog, 2003). The short cooking
274 time (4 min each side) and the relatively high thickness of beef patties (10 mm), may also explain the
275 observed low HAs formation. Costa et al. (2009) evaluated the effect of cooking time in HAs
276 formation in charcoal grilled sardines, and observed that no HAs were detected in samples cooked 5
277 minutes each side even at 280/300 °C.

278 Charcoal grilling creates a very dry environment, especially when samples are grilled near to the heat
279 source. PhIP and A α C formation are favored by higher temperatures and dry environment, in
280 opposite these conditions are disadvantageous to the formation of MeIQ_x and other thermic HAs
281 (Skog, Solyakov, & Jägerstad, 2000). Persson et al (2003) also observed that the addition of
282 NaCl/sodium tripolyphosphate to the beef burgers reduced the cooking loss and decreased the
283 formation of PhIP, MeIQ_x, and 4,8-DiMeIQ_x. This decrease was significant for MeIQ_x and 4,8-
284 DiMeIQ_x, which may explain that these HAs were not detected in our samples. Starch added to the
285 beef patties inhibited mutagenic activity by up to 54% (Skog, Jägerstad, & Laser Reuterswärd,
286 1992). The ingredients used may explain the absence of MeIQ_x in our samples, which is usually
287 found in cooked beef.

288 At day 0, RWPS patties showed higher contents of PhIP, A α C and MeA α C than control, although the
289 difference was not significant. Control samples cooked after 9 days of storage presented similar
290 values when compared with those from control at day 0 (Table 3). As observed for PAHs, RWPS
291 patties cooked after 9 days of storage presented values of HAs similar to day 0 control patties and
292 lower than those from day 0 RWPS patties. These results agree with the results from ABTS test as
293 described previously. The increased antioxidant activity observed during storage of RWPS patties

294 may also contribute the lower formation of HAs on patties cooked after 9 days of storage, as
295 previously stated for PAHs.

296 **4. CONCLUSIONS**

297 Low levels of PAHs and HAs were found in barbecued beef patties, probably linked to the low
298 cooking loss observed in the samples. Nine days of storage increased PAHs formation in barbecued
299 patties compared with control samples at day 0. RWPS patties cooked after 9 days of storage
300 presented similar PAHs levels found initially for control patties. HAs, other compound with
301 carcinogenic potential, were also evaluated but lower levels of these compounds were quantified and
302 no significant differences were observed in their formation after 9 days of storage. The addition of
303 RWPS during patties storage at high-oxygen may contribute to reduce the formation of carcinogenic
304 compounds after barbecuing in stored samples.

305 The formation of compounds with potential carcinogenic was reduced in those samples with higher
306 values in the ABTS assay, which may suggest a potential link between the formation of PAHs or
307 HAs and their ability to scavenge different radicals.

308 **ACKNOWLEDGEMENTS**

309 Authors are grateful for financial supports from the Autonomous Government of Castilla y León
310 through the research project BU282U13 and from project NORTE-01-0145-FEDER-000011 –
311 *Qualidade e Segurança Alimentar — uma abordagem (nano) tecnológica*. The PhD grant of J.
312 García-Lomillo (FPU grant) is funded by the Spanish “Ministerio de Educación, Cultura y Deporte”.

313

314 **REFERENCES**

- 315 Ahn, J., & Grün, I. U. (2005). Heterocyclic amines: 2. Inhibitory effects of natural extracts on the
316 formation of polar and nonpolar heterocyclic amines in cooked beef. *Journal of Food*
317 *Science*, 70(4), C263-C268.
- 318 Borgen, E., & Skog, K. (2004). Heterocyclic amines in some Swedish cooked foods industrially
319 prepared or from fast food outlets and restaurants. *Molecular Nutrition and Food Research*,
320 48(4), 292-298.
- 321 Bouvard, V., Loomis, D., Guyton, K. Z., Grosse, Y., Ghissassi, F. E., Benbrahim-Tallaa, L., Guha,
322 N., Mattock, H., & Straif, K. (2015). Carcinogenicity of consumption of red and processed
323 meat. *The Lancet Oncology*, 16(16), 1599-1600.
- 324 Busquets, R., Puignou, L., Galceran, M. T., & Skog, K. (2006). Effect of red wine marinades on the
325 formation of heterocyclic amines in fried chicken breast. *Journal of Agricultural and Food*
326 *Chemistry*, 54(21), 8376-8384.
- 327 Costa, M., Viegas, O., Melo, A., Petisca, C., Pinho, O., & Ferreira, I. M. P. L. V. O. (2009).
328 Heterocyclic Aromatic Amine Formation in Barbecued Sardines (*Sardina pilchardus*) and
329 Atlantic Salmon (*Salmo salar*). *Journal of Agricultural and Food Chemistry*, 57(8), 3173-
330 3179.
- 331 Chen, B. H., & Chen, Y. C. (2001). Formation of polycyclic aromatic hydrocarbons in the smoke
332 from heated model lipids and food lipids. *Journal of Agricultural and Food Chemistry*,
333 49(11), 5238-5243.
- 334 Chung, S. Y., Yettella, R. R., Kim, J. S., Kwon, K., Kim, M. C., & Min, D. B. (2011). Effects of
335 grilling and roasting on the levels of polycyclic aromatic hydrocarbons in beef and pork.
336 *Food Chemistry*, 129(4), 1420-1426.
- 337 Del Pino-García, R., García-Lomillo, J., Rivero-Pérez, M. D., González-Sanjosé, M. L., & Muñoz, P.
338 (2015). Adaptation and Validation of QUick, Easy, New, CHEap, and Reproducible

339 (QUENCHER) Antioxidant Capacity Assays in Model Products Obtained from Residual
340 Wine Pomace. *Journal of Agricultural and Food Chemistry*, 63(31), 6922-6931.

341 Del Pino-García, R., Gerardi, G., Rivero-Pérez, M. D., González-SanJosé, M. L., García-Lomillo, J.,
342 & Muñiz, P. (2016). Wine pomace seasoning attenuates hyperglycaemia-induced endothelial
343 dysfunction and oxidative damage in endothelial cells. *Journal of Functional Foods*, 22, 431-
344 445.

345 EFSA. (2008). Scientific opinion of the panel on contaminants in the food chain on a request from
346 the European Commission on polycyclic aromatic hydrocarbons in food. *EFSA Journal*, 724,
347 1-114.

348 Garcia-Lomillo, J., González-SanJosé, M., Skibsted, L., & Jongberg, S. (2016). Effect of skin wine
349 pomace and sulfite on protein oxidation in beef patties during high oxygen atmosphere
350 storage. *Food and Bioprocess Technology*, 9(3), 532-542.

351 García-Lomillo, J., González-SanJosé, M. L., Del Pino-García, R., Rivero-Pérez, M. D., & Muñiz-
352 Rodríguez, P. (2014). Antioxidant and antimicrobial properties of wine byproducts and their
353 potential uses in the food industry. *Journal of Agricultural and Food Chemistry*, 62(52),
354 12595-12602.

355 González San José, M., García Lomillo, J., Del Pino García, R., Dolores Rivero, M., & Muñiz
356 Rodríguez, P. (2015). Spain Patent No. ES 2 524 870 B2. Oficina Española de Patentes y
357 Marcas.

358 Janoszka, B. (2011). HPLC-fluorescence analysis of polycyclic aromatic hydrocarbons (PAHs) in
359 pork meat and its gravy fried without additives and in the presence of onion and garlic. *Food*
360 *Chemistry*, 126(3), 1344-1353.

361 Lee, J.-G., Kim, S.-Y., Moon, J.-S., Kim, S.-H., Kang, D.-H., & Yoon, H.-J. (2016). Effects of
362 grilling procedures on levels of polycyclic aromatic hydrocarbons in grilled meats. *Food*
363 *Chemistry*, 199, 632-638.

364 Marchiani, R., Bertolino, M., Belviso, S., Giordano, M., Ghirardello, D., Torri, L., Piochi, M., &
365 Zeppa, G. (2016). Yogurt enrichment with grape pomace: Effect of grape cultivar on
366 physicochemical, microbiological and sensory properties. *Journal of Food Quality*, 39, 77-89.

367 McGrath, T., Sharma, R., & Hajaligol, M. (2001). An experimental investigation into the formation
368 of polycyclic-aromatic hydrocarbons (PAH) from pyrolysis of biomass materials. *Fuel*,
369 80(12), 1787-1797.

370 Melo, A., Viegas, O., Petisca, C., Pinho, O., & Ferreira, I. M. P. L. V. (2008). Effect of beer/red wine
371 marinades on the formation of heterocyclic aromatic amines in pan-fried beef. *Journal of*
372 *Agricultural and Food Chemistry*, 56(22), 10625-10632.

373 Nawar, W. W. (1984). Chemical changes in lipids produced by thermal processing. *Journal of*
374 *Chemical Education*, 61(4), 299.

375 Nirmal, N. P., & Benjakul, S. (2011). Use of tea extracts for inhibition of polyphenoloxidase and
376 retardation of quality loss of Pacific white shrimp during iced storage. *Lwt-Food Science and*
377 *Technology*, 44(4), 924-932.

378 Pazos, M., González, M. J., Gallardo, J. M., Torres, J. L., & Medina, I. (2005). Preservation of the
379 endogenous antioxidant system of fish muscle by grape polyphenols during frozen storage.
380 *European Food Research and Technology*, 220(5-6), 514-519.

381 Persson, E., Graziani, G., Ferracane, R., Fogliano, V., & Skog, K. (2003). Influence of antioxidants
382 in virgin olive oil on the formation of heterocyclic amines in fried beefburgers. *Food and*
383 *Chemical Toxicology*, 41(11), 1587-1597.

384 Polak, T., Andrenšek, S., Žlender, B., & Gašperlin, L. (2009). Effects of ageing and low internal
385 temperature of grilling on the formation of heterocyclic amines in beef Longissimus dorsi
386 muscle. *LWT - Food Science and Technology*, 42(1), 256-264.

387 Rivero-Pérez, M. D., Muñiz, P., & González-Sanjosé, M. L. (2007). Antioxidant profile of red wines
388 evaluated by total antioxidant capacity, scavenger activity, and biomarkers of oxidative stress
389 methodologies. *Journal of Agricultural and Food Chemistry*, 55(14), 5476-5483.

390 Santos, F. J., Barceló-Barrachina, E., Toribio, F., Puignou, L., Galceran, M. T., Persson, E., Skog,
391 K., Messner, C., Murkovic, M., Nabinger, U., & Ristic, A. (2004). Analysis of heterocyclic
392 amines in food products: interlaboratory studies. *Journal of Chromatography B*, 802(1), 69-
393 78.

394 Sebranek, J. G. (2009). Basic Curing Ingredients. In R. Tarté (Ed.), *Ingredients in Meat Products:
395 Properties, Functionality and Applications* (pp. 1-23). New York, NY: Springer New York.

396 Serpen, A., Gökmen, V., & Fogliano, V. (2012). Total antioxidant capacities of raw and cooked
397 meats. *Meat Science*, 90(1), 60-65.

398 Sharma, R. K., Chan, W. G., & Hajaligol, M. R. (2006). Product compositions from pyrolysis of
399 some aliphatic α -amino acids. *Journal of Analytical and Applied Pyrolysis*, 75(2), 69-81.

400 Sharma, R. K., & Hajaligol, M. R. (2003). Effect of pyrolysis conditions on the formation of
401 polycyclic aromatic hydrocarbons (PAHs) from polyphenolic compounds. *Journal of
402 Analytical and Applied Pyrolysis*, 66(1-2), 123-144.

403 Skog, K. I., Jägerstad, M., & Laser Reuterswärd, A. (1992). Inhibitory effect of carbohydrates on the
404 formation of mutagens in fried beef patties. *Food and Chemical Toxicology*, 30(8), 681-688.

405 Skog, K. I., Johansson, M. A. E., & Jägerstad, M. I. (1998). Carcinogenic Heterocyclic Amines in
406 Model Systems and Cooked Foods: A Review on Formation, Occurrence and Intake. *Food
407 and Chemical Toxicology*, 36(9-10), 879-896.

408 Skog, K. I., Solyakov, A., & Jägerstad, M. (2000). Effects of heating conditions and additives on the
409 formation of heterocyclic amines with reference to amino-carbolines in a meat juice model
410 system. *Food Chemistry*, 68(3), 299-308.

- 411 Szterk, A., Roszko, M., Małek, K., Kurek, M., Zbieć, M., & Waszkiewicz-Robak, B. (2012). Profiles
412 and concentrations of heterocyclic aromatic amines formed in beef during various heat
413 treatments depend on the time of ripening and muscle type. *Meat Science*, 92(4), 587-595.
- 414 Szterk, A., & Waszkiewicz-Robak, B. (2014). Influence of selected quality factors of beef on the
415 profile and the quantity of heterocyclic aromatic amines during processing at high
416 temperature. *Meat Science*, 96(3), 1177-1184.
- 417 U.S. Department of Agriculture, A. R. S. (2012). USDA Table of Cooking Yields for Meat and
418 Poultry.: Nutrient Data Laboratory Home Page: <http://www.ars.usda.gov/nutrientdata>.
- 419 Viegas, O., Amaro, L. F., Ferreira, I. M. P. L. V. O., & Pinho, O. (2012). Inhibitory effect of
420 antioxidant-rich marinades on the formation of heterocyclic aromatic amines in pan-fried
421 beef. *Journal of Agricultural and Food Chemistry*, 60(24), 6235-6240.
- 422 Viegas, O., Moreira, P. S., & Ferreira, I. M. P. L. V. O. (2015). Influence of beer marinades on the
423 reduction of carcinogenic heterocyclic aromatic amines in charcoal-grilled pork meat. *Food*
424 *Additives and Contaminants - Part A Chemistry, Analysis, Control, Exposure and Risk*
425 *Assessment*, 32(3), 315-323.
- 426 Viegas, O., Novo, P., Pinho, O., & Ferreira, I. M. P. L. V. O. (2012). A comparison of the extraction
427 procedures and quantification methods for the chromatographic determination of polycyclic
428 aromatic hydrocarbons in charcoal grilled meat and fish. *Talanta*, 88, 677-683.
- 429 Viegas, O., Novo, P., Pinto, E., Pinho, O., & Ferreira, I. M. P. L. V. O. (2012). Effect of charcoal
430 types and grilling conditions on formation of heterocyclic aromatic amines (HAs) and
431 polycyclic aromatic hydrocarbons (PAHs) in grilled muscle foods. *Food and Chemical*
432 *Toxicology*, 50(6), 2128-2134.
- 433 Viegas, O., Yebra-Pimentel, I., Martínez-Carballo, E., Simal-Gandara, J., & Ferreira, I. M. P. L. V.
434 O. (2014). Effect of beer marinades on formation of polycyclic aromatic hydrocarbons in
435 charcoal-grilled pork. *Journal of Agricultural and Food Chemistry*, 62(12), 2638-2643.

436 Yebra-Pimentel, I., Fernández-González, R., Martínez-Carballo, E., & Simal-Gándara, J. (2015). A
437 Critical Review about the Health Risk Assessment of PAHs and Their Metabolites in Foods.
438 *Critical Reviews in Food Science and Nutrition*, 55(10), 1383-1405.
439

Table 1. Chemical composition and cooking loss of beef patties raw and cooked without red wine pomace seasoning (control) or with red wine pomace seasoning (RWPS).

	Raw patties	Cooked patties			
		Control		RWPS	
		Day 0	Day 9	Day 0	Day 9
Moisture (%)	64.2 ± 0.6	57.0 ± 0.6	57.0 ± 0.2	57.3 ± 0.1	57.9 ± 0.5
Protein (%)	18.3 ± 0.3	22.6 ± 0.6	21.3 ± 0.4	21.7 ± 0.3	20.6 ± 0.6
Lipid (%)	12.4 ± 0.8	9.7 ± 0.3	11.7 ± 0.6	11.0 ± 0.3	10.5 ± 0.4
% Cooking loss		28.6 ± 0.7	29.2 ± 0.7	30.7 ± 1.1	30.7 ± 0.7

^a Results are presented as the mean ± standard error (n = 2 batches x 3 analytical replicates). No significant differences were observed between cooked samples at a *P*-value < 0.05.

Table 2. PAHs content in barbecued beef patties without seasoning (control) and with seasoning (2% w/w) at day 0 and day 9 storage at high-oxygen atmosphere. ^a

	Control Patties		RWPS patties	
	Day 0	Day 9	Day 0	Day 9
<i>Light PAHs</i>				
A	0.86 ± 0.12 a	1.37 ± 0.35 a	2.28 ± 0.19b	0.71 ± 0.05 a
<i>Heavy PAHs</i>				
FA	3.35 ± 0.21 a	7.36 ± 0.85 c	5.42 ± 0.25 b	3.20 ± 0.27 a
PYR	3.04 ± 0.38 a	4.28 ± 0.21 b	5.10 ± 0.14 b	2.40 ± 0.20 a
BaA	0.42 ± 0.06 a,b	0.51 ± 0.04 b,c	0.66 ± 0.05 c	0.32 ± 0.10 a
CHR	0.63 ± 0.05 a,b	0.76 ± 0.09 b	1.30 ± 0.03 c	0.54 ± 0.09 a
BbFA	0.30 ± 0.06 a	0.55 ± 0.06 b	0.53 ± 0.05 b	0.37 ± 0.11 a,b
BkFA	0.19 ± 0.03	0.18 ± 0.01	0.18 ± 0.01	0.18 ± 0.03
BaP	0.22 ± 0.04 a	0.20 ± 0.03 a	0.34 ± 0.05 b	0.23 ± 0.04 a,b
DBahA	0.21 ± 0.03	0.20 ± 0.01	0.18 ± 0.02	0.17 ± 0.03
BghiP	0.30 ± 0.05	0.34 ± 0.08	0.32 ± 0.03	0.24 ± 0.07
IP	0.15 ± 0.02 a	0.24 ± 0.01 a,b	0.33 ± 0.03 b	0.43 ± 0.04 c
∑PAHs	9.67 ± 0.95 a	15.98 ± 0.94 b	16.63 ± 0.44 b	8.79 ± 0.81 a

^a Results are presented as the mean ± standard error (n = 2 batches x 3 analytical replicates). Means with different letters in the same row represent values significantly different (*P*-value < 0.05).

Table 3. HAs content (ng/g) in barbecued beef patties without seasoning (control) and with seasoning (2% w/w) at day 0 and day 9 storage at high-oxygen atmosphere.^a

	Control Patties		RWPS Patties	
	Day 0	Day 9	Day 0	Day 9
PhIP	0.86 ± 0.10 a,b	0.68 ± 0.17 a	1.16 ± 0.21 b	0.69 ± 0.08 a
AαC	0.33 ± 0.04 a,b	0.24 ± 0.01 a	0.49 ± 0.09 b	0.38 ± 0.09 a,b
MeAC	n.q.	n.d.	0.29 ± 0.06	n.d.

^aResults are presented as the mean ± standard error (n = 2 batches x 3 analytical replicates). Means with different letters in the same row are significantly different (*P*-value < 0.05).n.d.: not detected; n.q: not quantifiable. LOQ (0.02 ng/g) and LOD (0.25 ng/g) were previously determined (Melo et al., 2008)

Highlights

- Red wine pomace avoids the loss in ABTS scavenging activity during meat storage;
- Storage increases the PAHs formation in beef patties;
- Red wine pomace seasoning reduces PAHs formation in stored patties;
- A seasoning that inhibits cooking carcinogens in stored patties is presented.