

Subcritical water extraction and identification of phenolic compounds from **Brewer's Spent Grain (BSG)**

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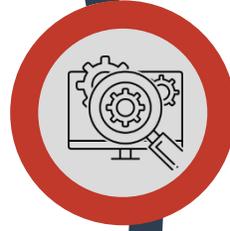
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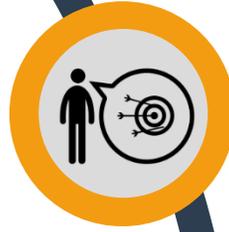


05 - CONCLUSIONS

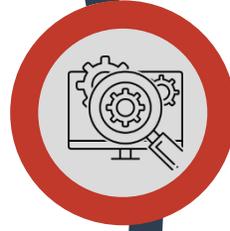
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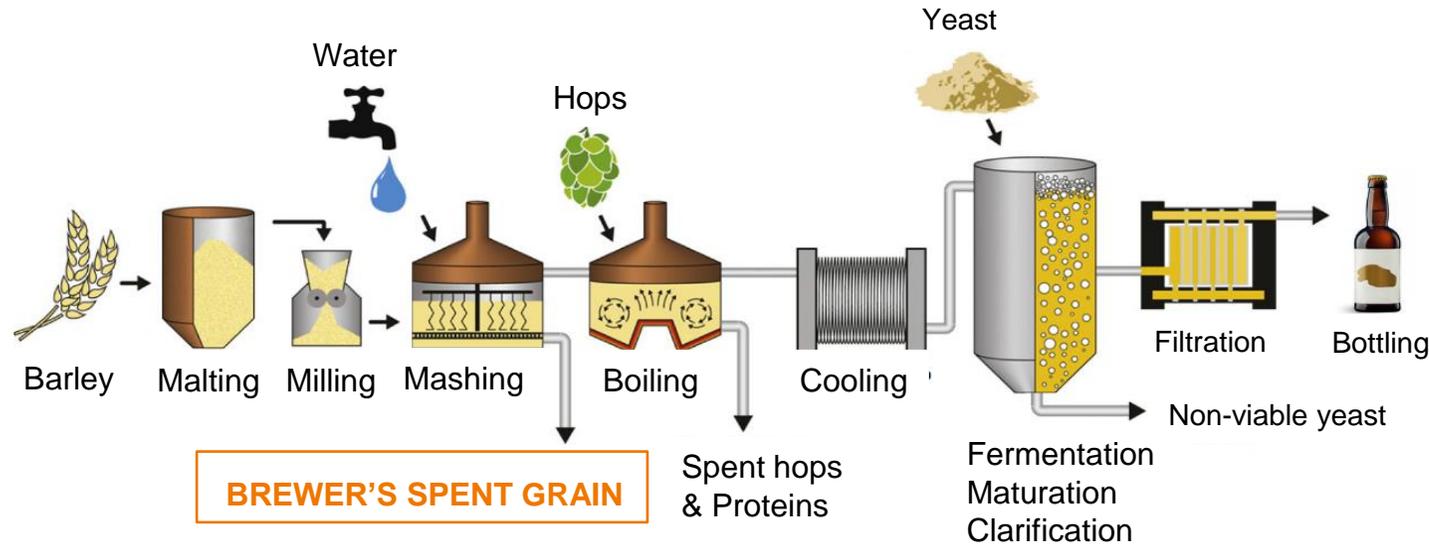


Circular Economy





Brewer's Spent Grain (BSG)

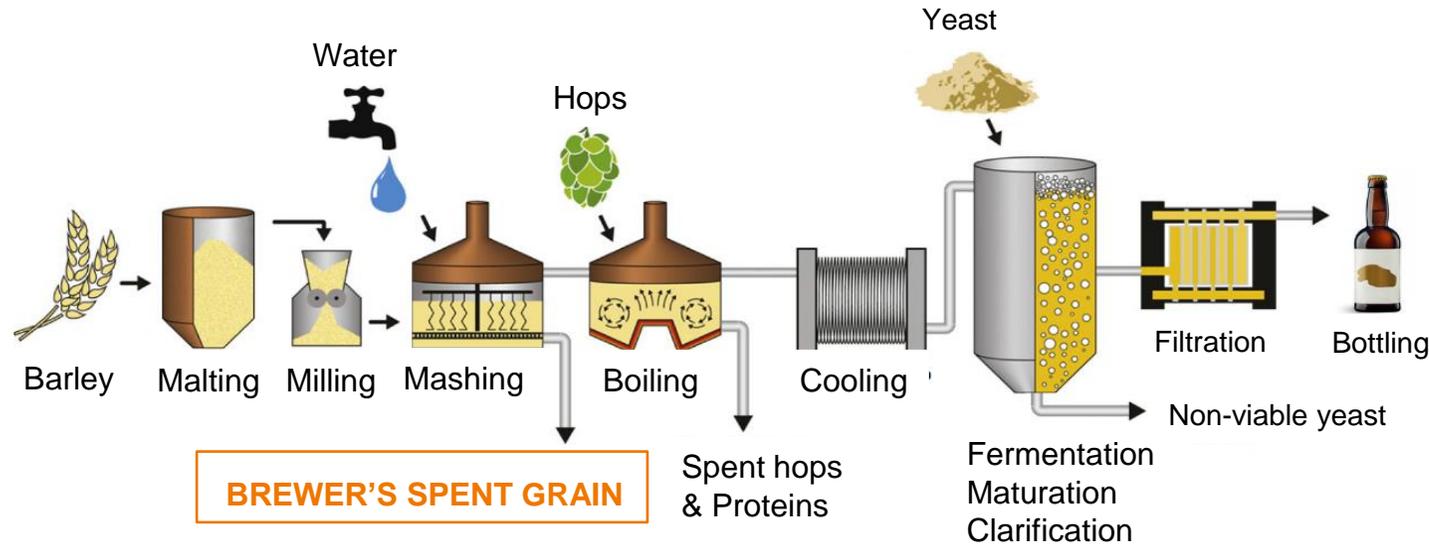


85 % of the total **by-products** generated → 20 kg of BSG/ 100 L of beer produced.

- **BSG** generated by a **small local** company → small breweries within the biorefinery concept
- ✓ Europe craft beer market → US\$ 42.52 million in 2020 → Expected to grow up to US\$ 91.26 million by the end of 2025



Brewer's Spent Grain (BSG)



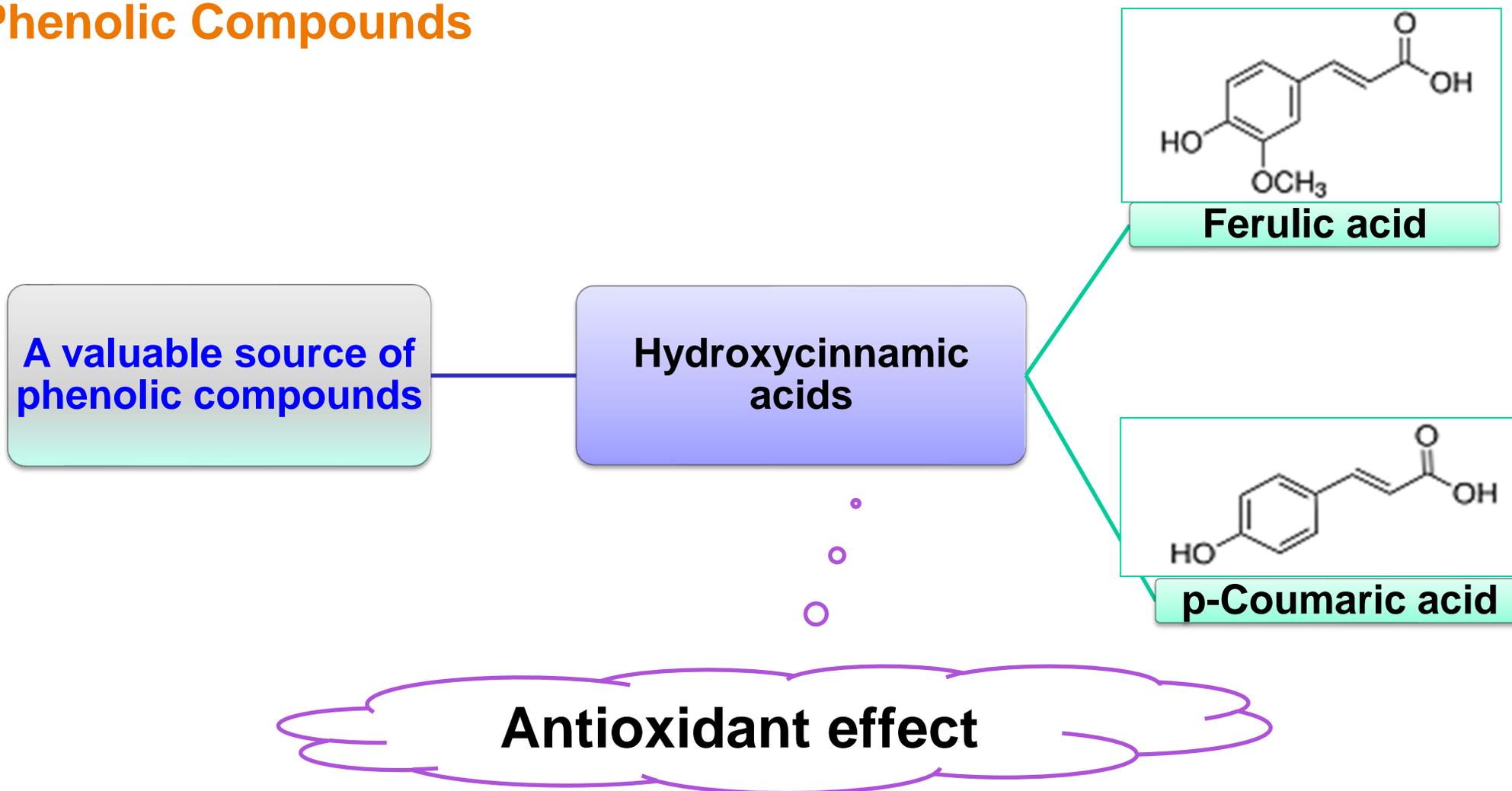
85 % of the total **by-products** generated → 20 kg of BSG/ 100 L of beer produced.

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Integration of **BSG** within a **biorefinery** concept is of great interest to obtain different high value **biocompounds**

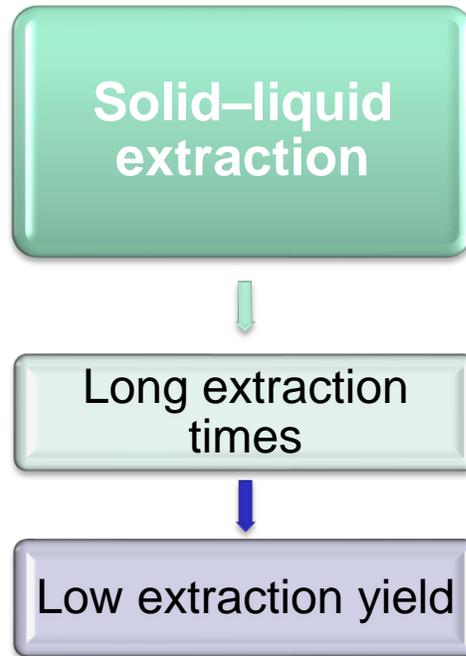


✓ Phenolic Compounds

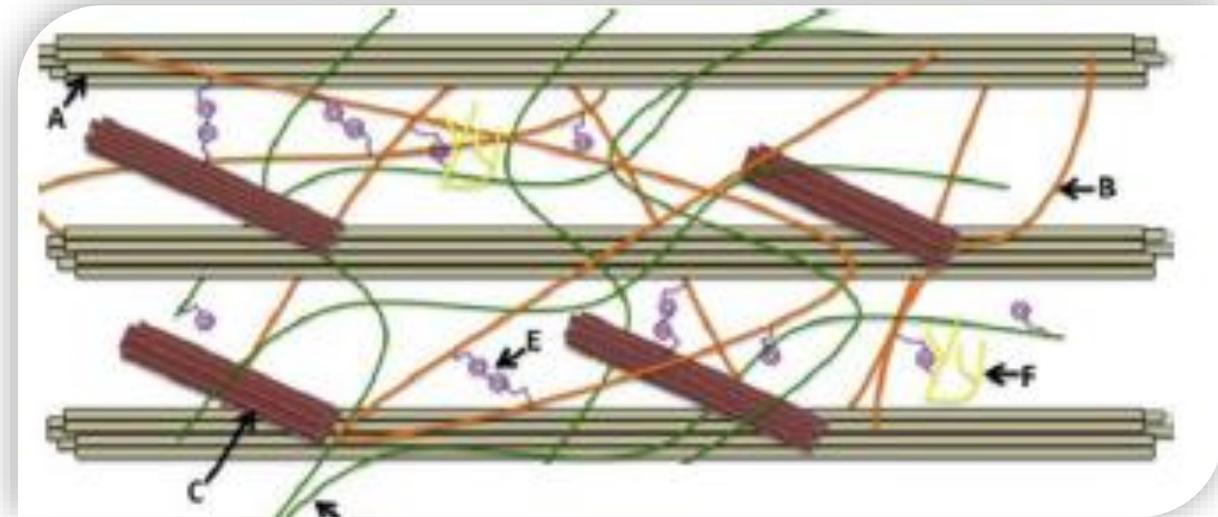




✓ Extract Phenolic Compounds



BSG, a lignocellulosic material



Lignin is connected to the cell wall polysaccharides by phenolic acids, being necessary a hydrolytic method to release them



✓ Extract Phenolic Compounds

Different hydrolytic techniques

Ultrasound assisted
extraction (UAE)

Chemical hydrolysis

Enzymatic hydrolysis



✓ Extraction of Phenolic Compounds



Subcritical Water (subW)

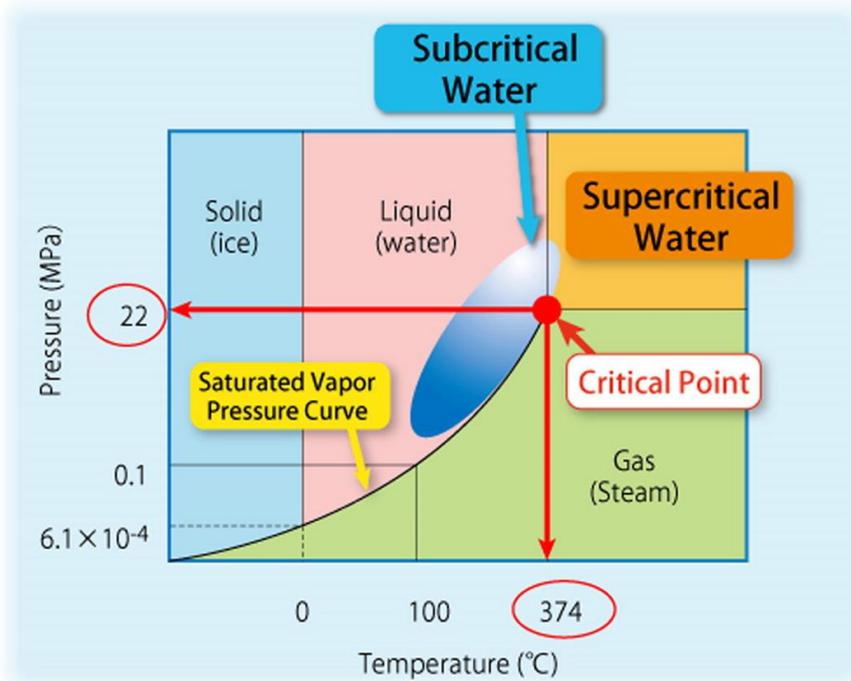
An **alternative technology** to **chemical and enzymatic hydrolysis** to release **Phenolic Compounds** from lignocellulosic materials



Subcritical Water (subW)

Water is the **greenest** solvent of all considering the principles of green chemistry

subW is pressurized **water** in its liquid state in the T range from **100 °C** to **374 °C**.



Asia Biomass Office. (2014)



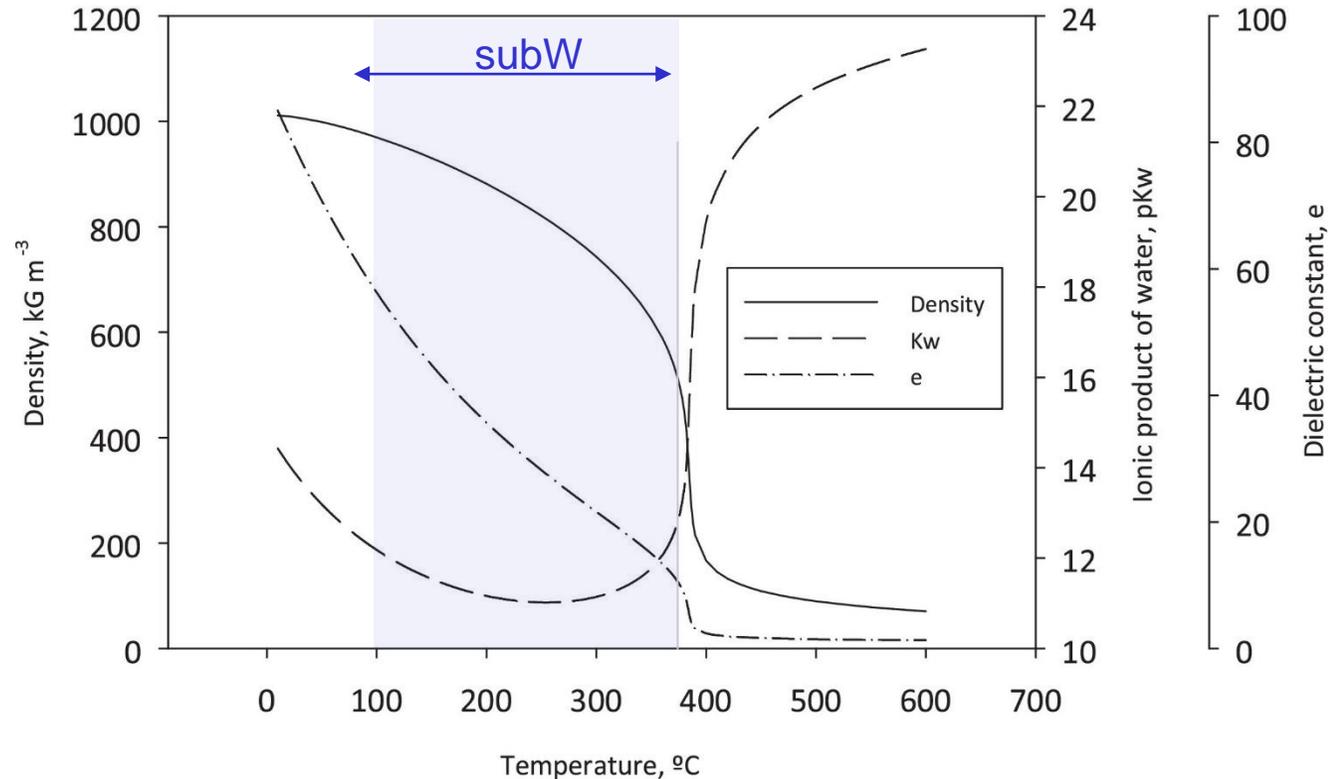
Subcritical Water (subW)

subW presents unique properties

↑ Ionic product

↓ Dielectric constant

↓ Density



↑ Affinity towards non-polar compounds

Cocero et al. (2018)



Subcritical Water (subW)

Tunable properties with **T**

Different **selectivity** in the release of the bioactive compounds from the biomass

T in **subW** is a **critical** parameter

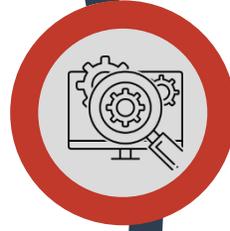
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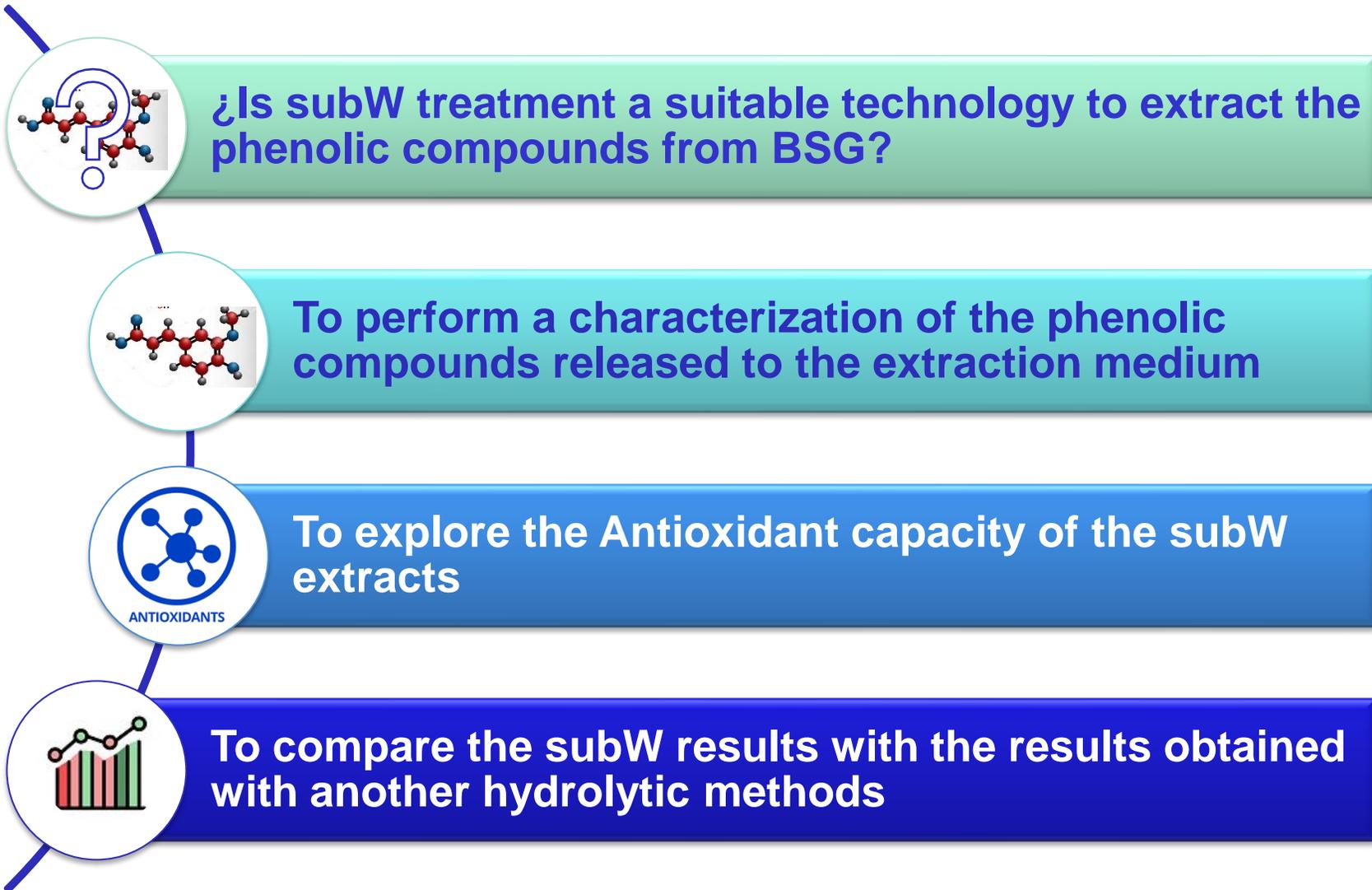


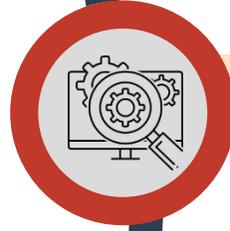
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Preconditioning



Washing with water



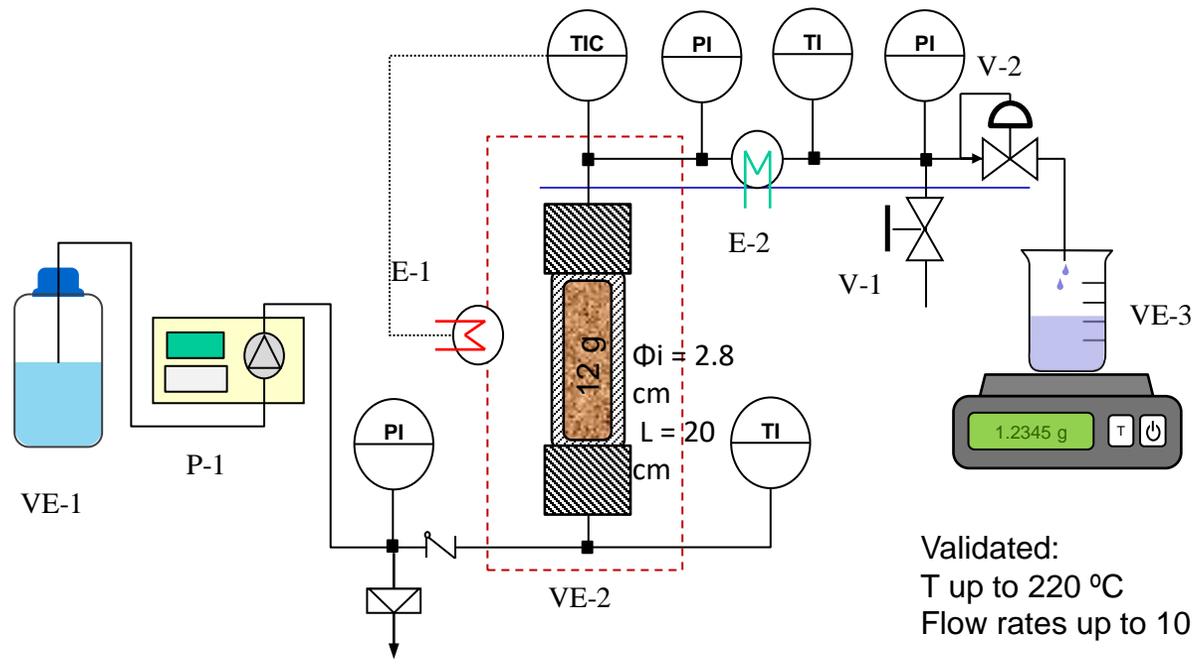
Drying in an air convection oven (45 °C, 3 h)

Humidity < 8 % (w/w)

BSG generated in a craft beer industry.

Subcritical water (subW) experiments

Semi-continuous fixed-bed reactor



Temperature (125-185 °C)

P= 50 bar

F= 4 mL/min

Validated:
 T up to 220 °C
 Flow rates up to 10 mL/min.



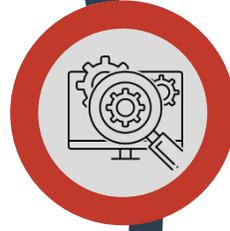
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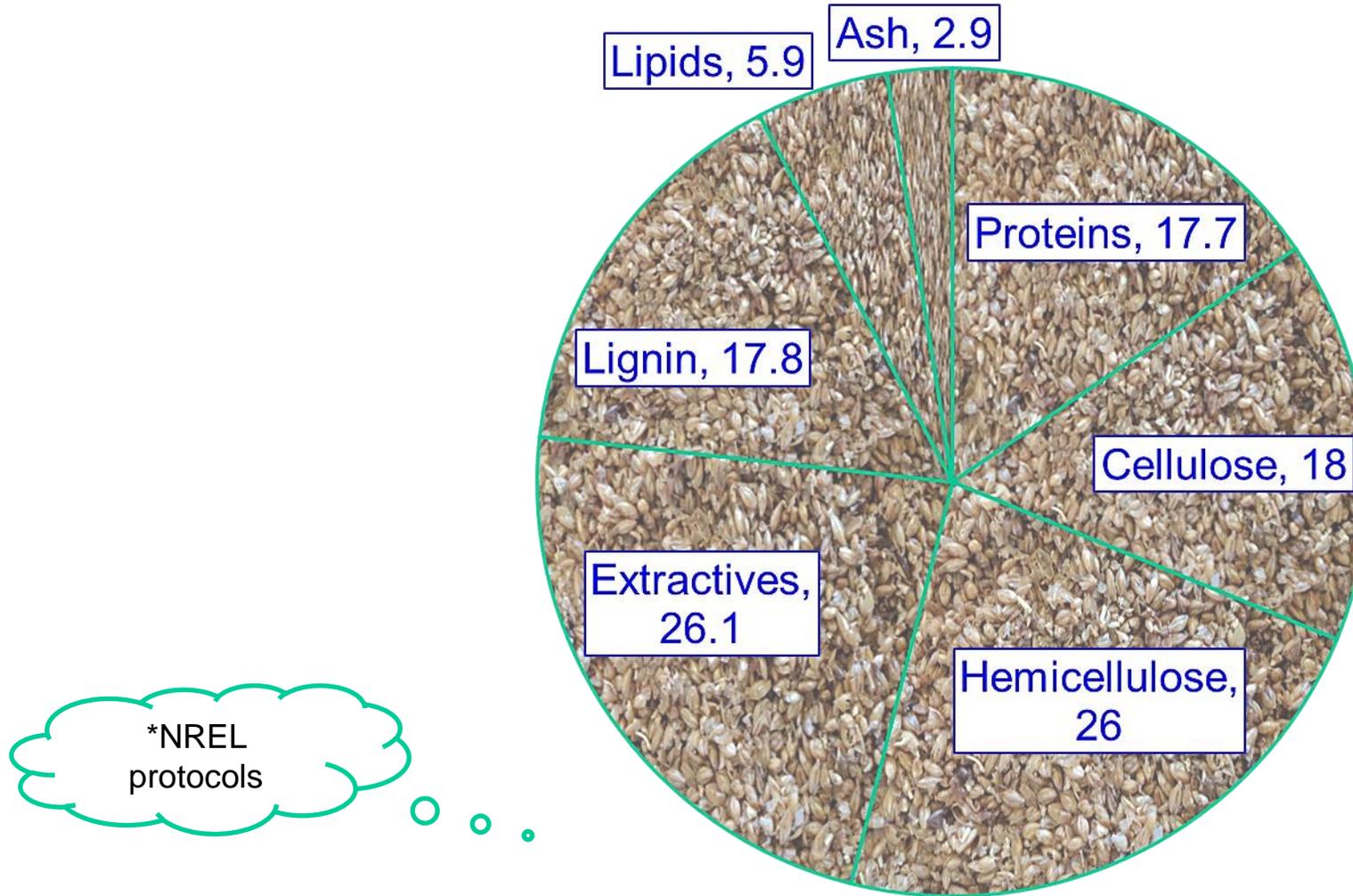
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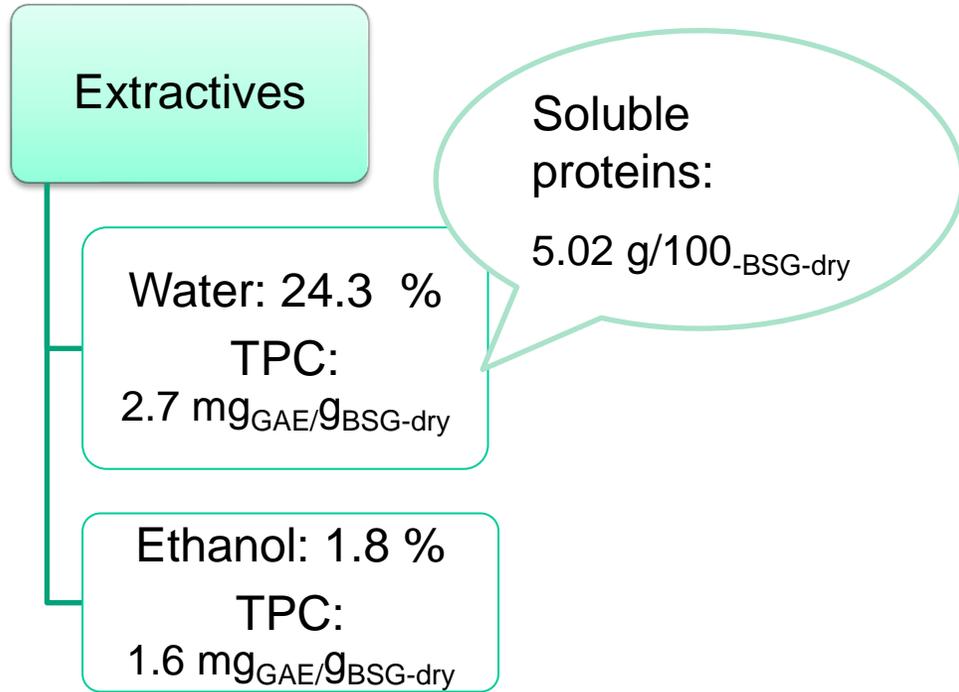
BSG characterization



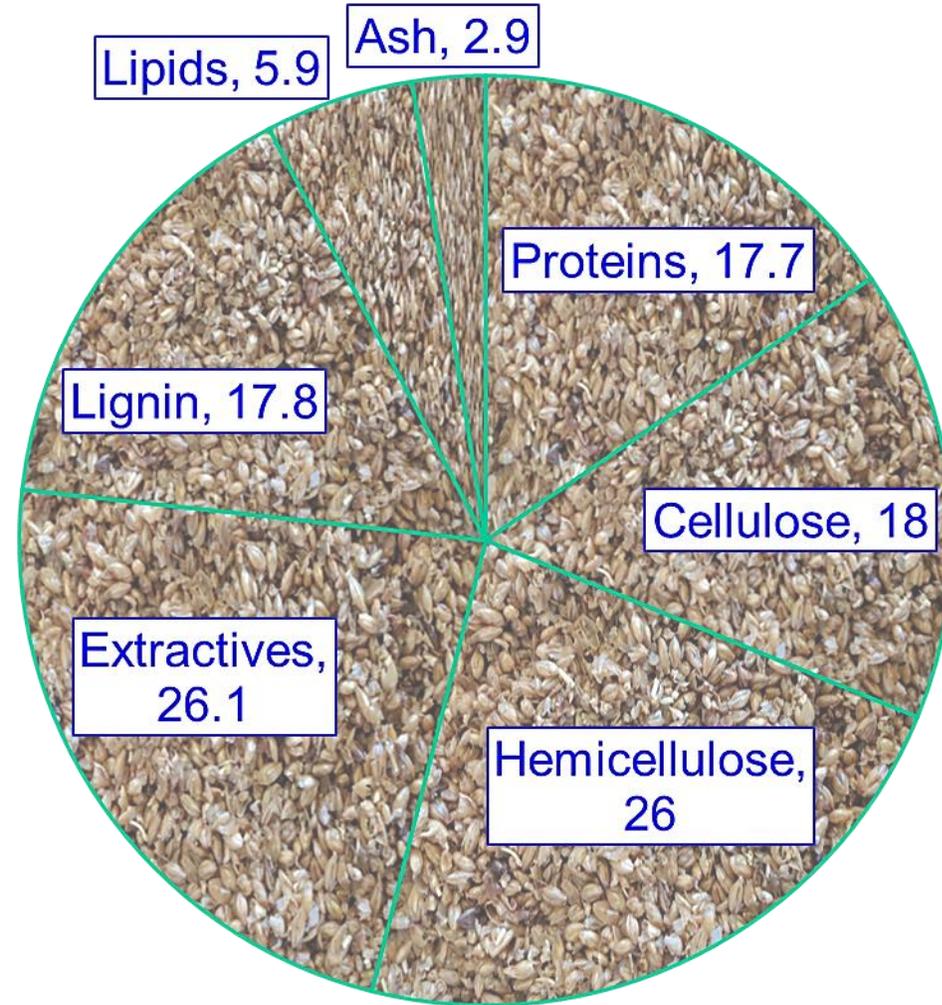
BSG composition, g/100g_{BSG-dry}



BSG characterization



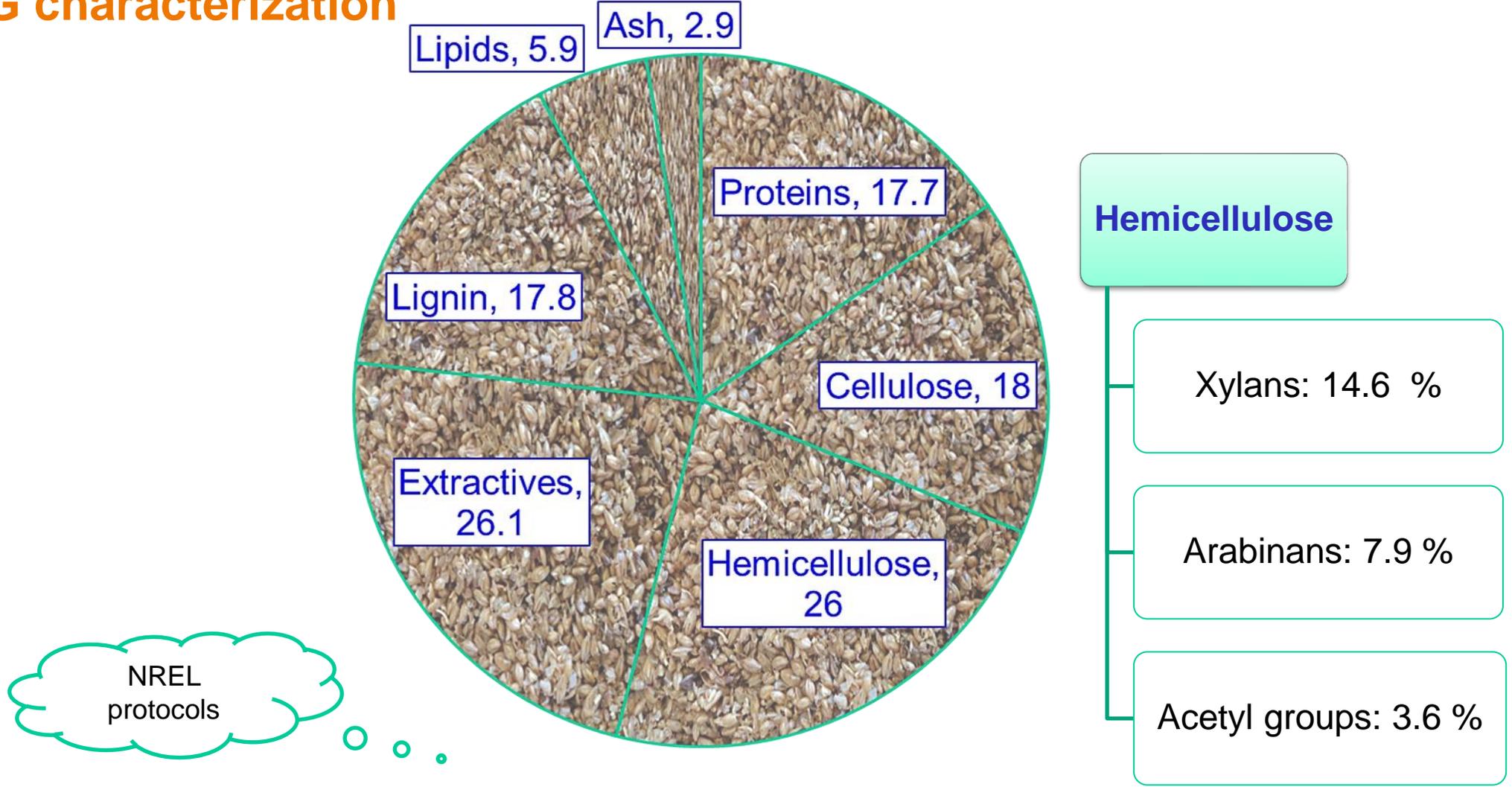
NREL protocols



BSG composition, g/100g_{BSG-dry}



BSG characterization

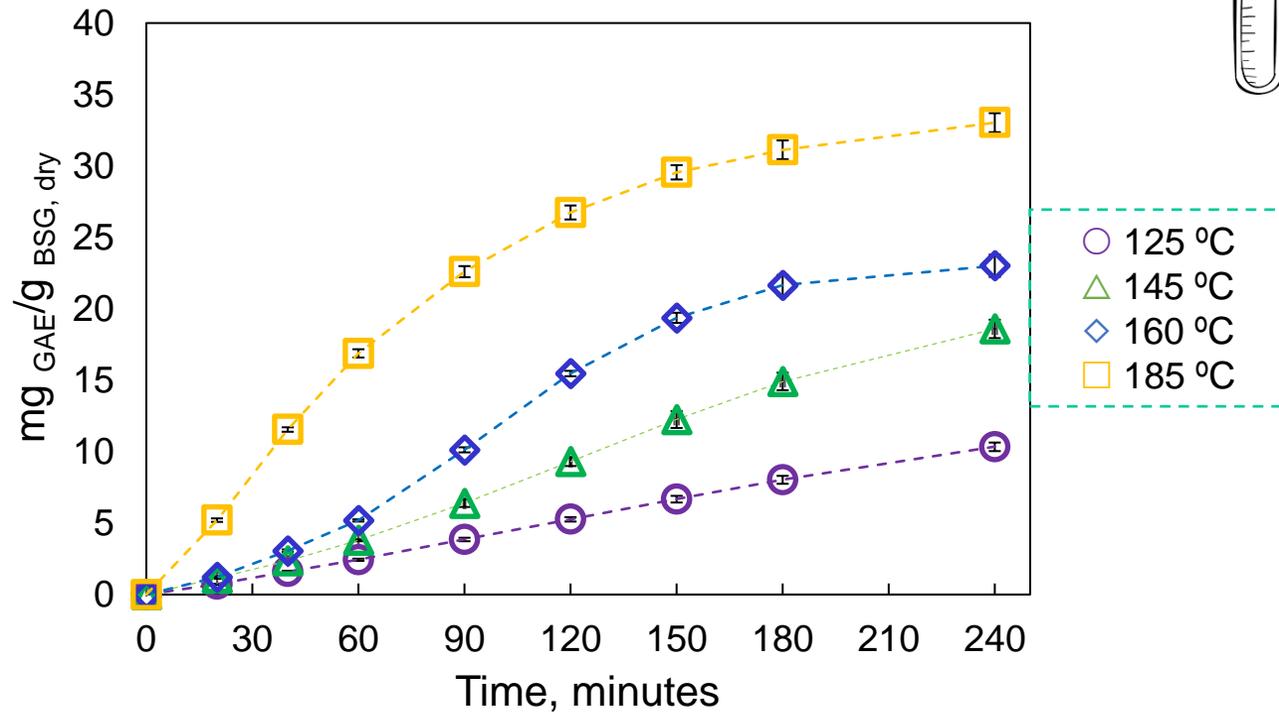


BSG composition, g/100g_{BSG-dry}

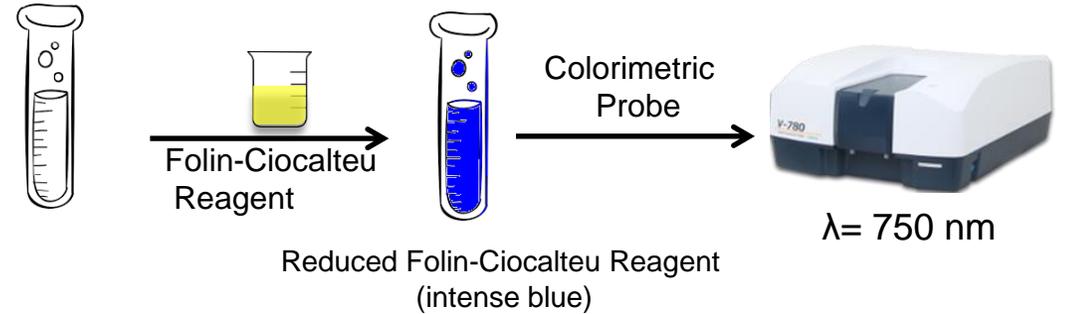


Total Phenolic Content (TPC)

TPC. Folin-Ciocalteu method

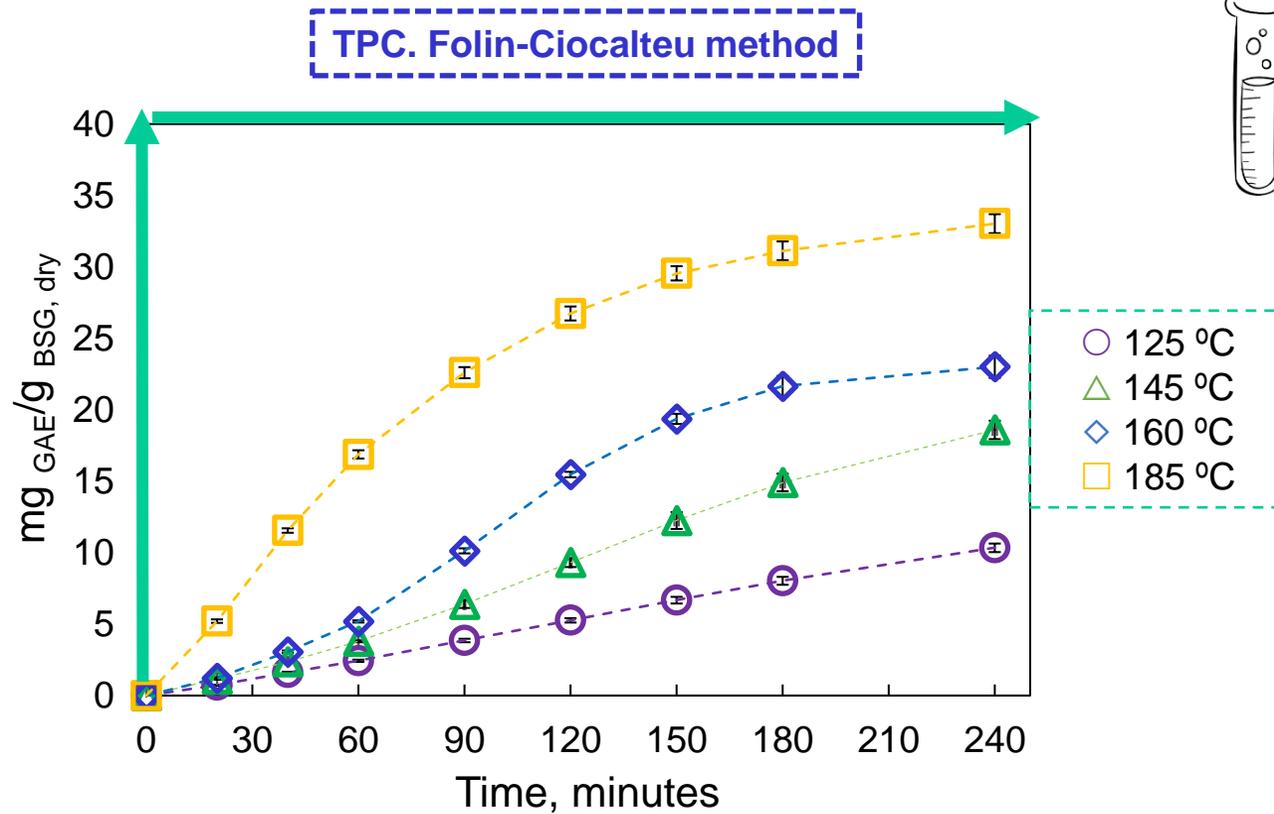


Effect of temperature on TPC release in subW extracts

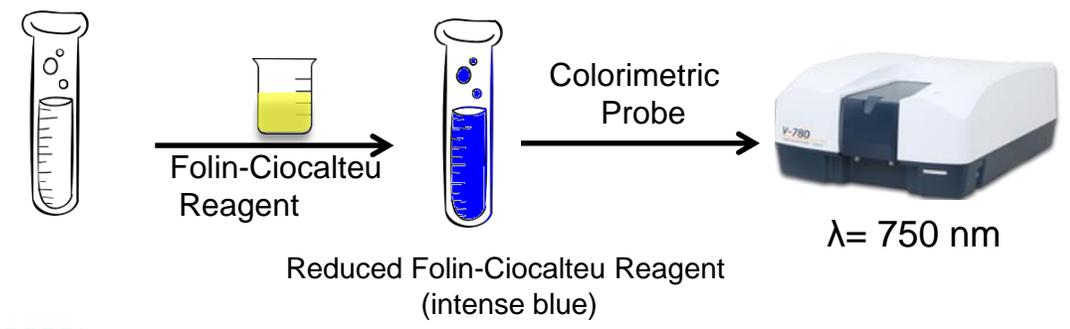




Total Phenolic Content (TPC)



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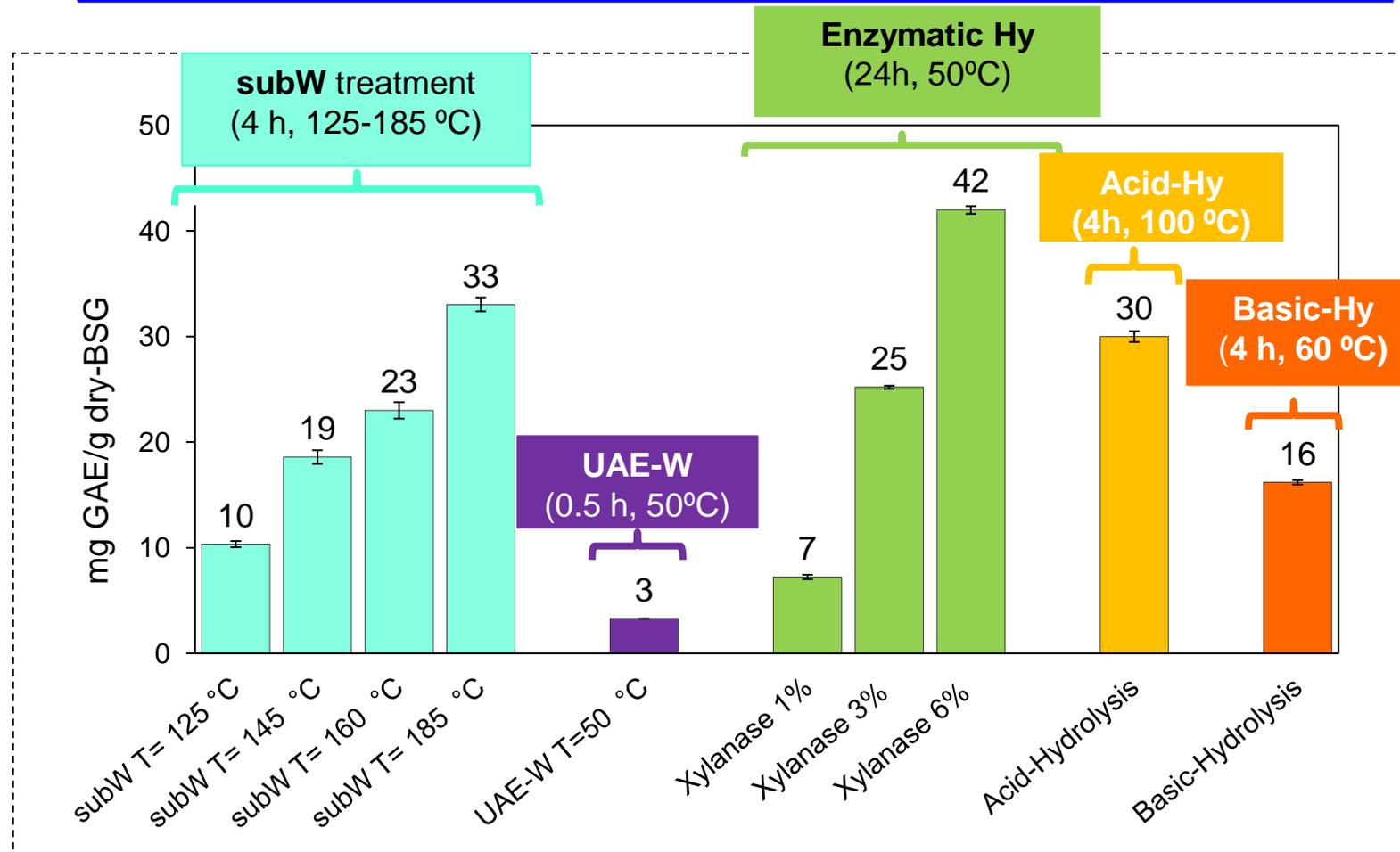


An increase in the operating **temperature** led to an **increase in TPC** in the extraction medium



Total Phenolic Content (TPC)

Comparison between the different hydrolytic methods



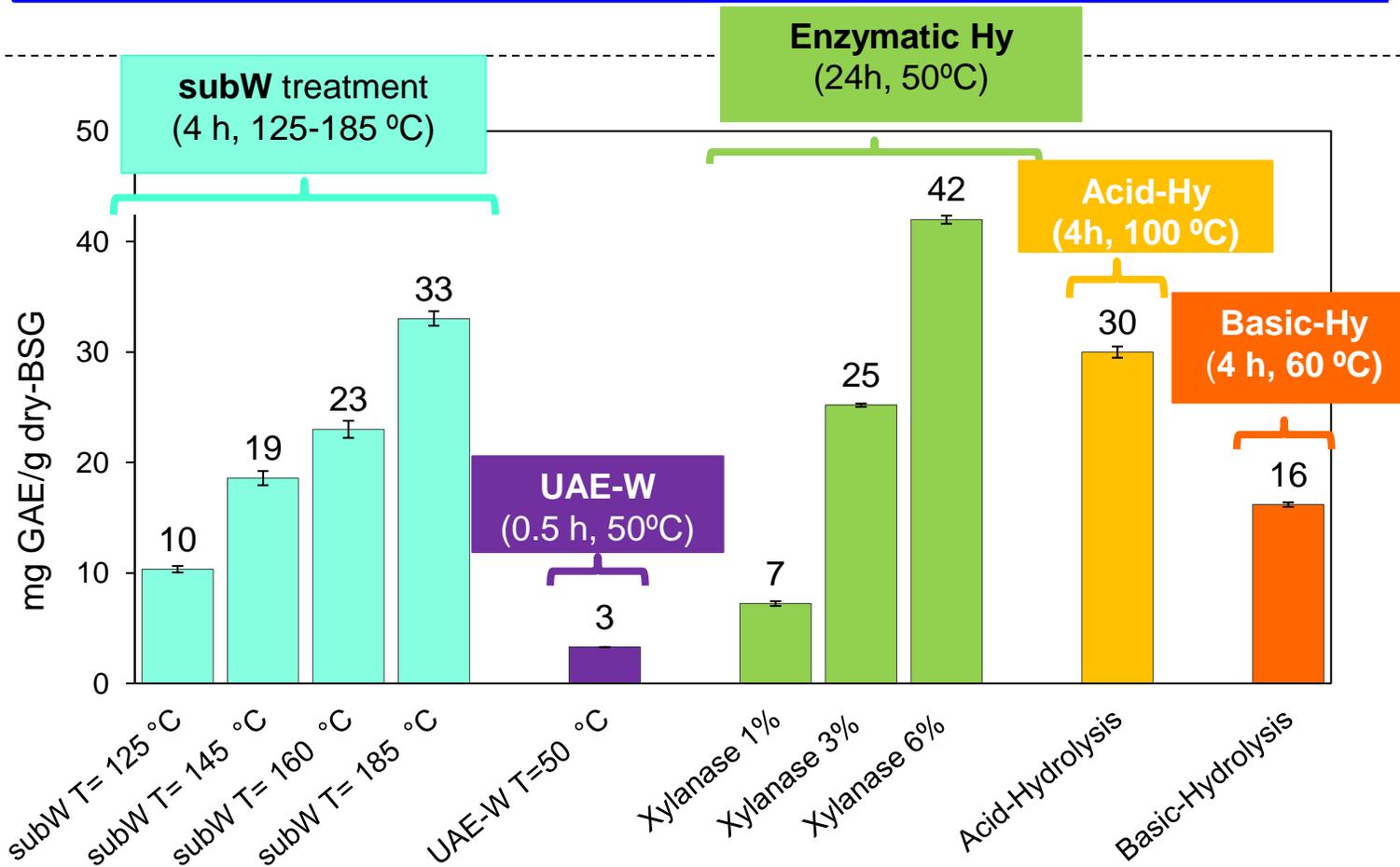
TPC determined by Folin-Ciocalteu in different extracts. UAE-W= Ultrasonic Assisted Extraction with water



Total Phenolic Content (TPC)

subW treatment at 185 °C led to higher amount of TPC than acid and basic hydrolysis and UAE-W

Comparison between the different hydrolytic methods



A maximum TPC release was found with 6% of xylanase for 24 h

TPC determined by Folin-Ciocalteu in different extracts. UAE-W= Ultrasonic Assisted Extraction with water





Total Phenolic Content (TPC)

Comparison between the different hydrolytic methods

TPC Productivity (P)*

$$P = \frac{mg_{GAE}}{g_{BSG, dry} \cdot \Delta t}$$

Method	P mg _{GAE} /(g _{BSG dry} ·min)
subW T= 160 °C	0.100 ± 0.001
subW T= 185 °C	0.28 ± 0.01
UAE T= 50 °C	0.109 ± 0.002
Basic Hydrolysis	0.55 ± 0.04
Xylanase 6%	0.087 ± 0.001

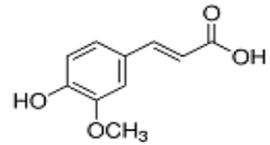
Productivity by enzymatic hydrolysis was lower than for subW at 160 °C and 185 °C.

*Evaluated from the initial linear extraction curve.

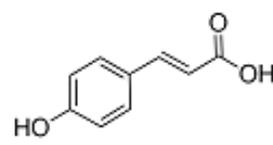


Individual Phenolic Compounds

Hydroxycinnamic acids

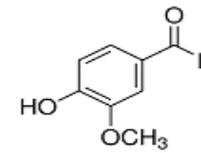


◆◆ Ferulic acid

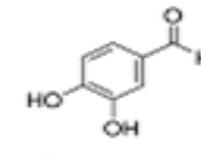


△▲ p-Coumaric acid

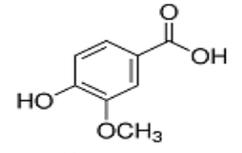
Phenolic aldehydes



□■ Vanillin



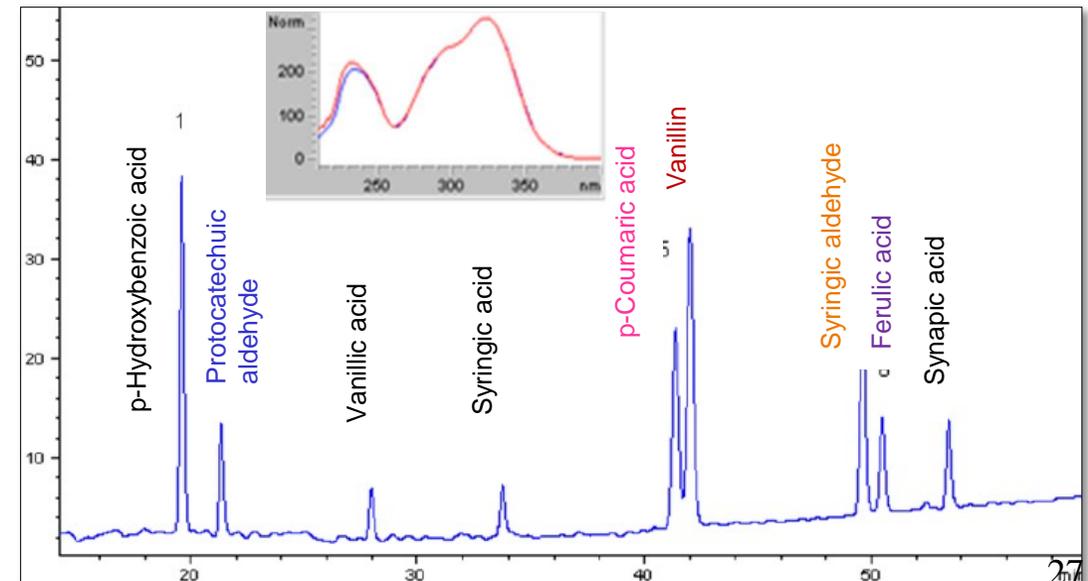
◆◆ Protocatechuic aldehyde



○● Syringic aldehyde



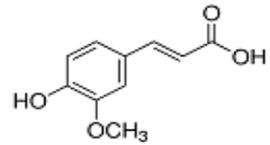
HPLC-DAD



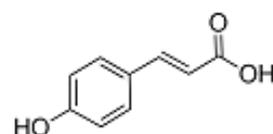


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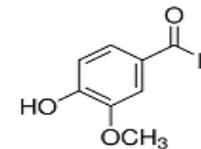


◆ Ferulic acid

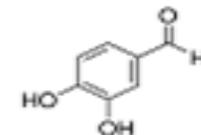


▲ p-Coumaric acid

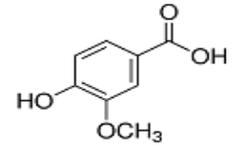
Aldehydes



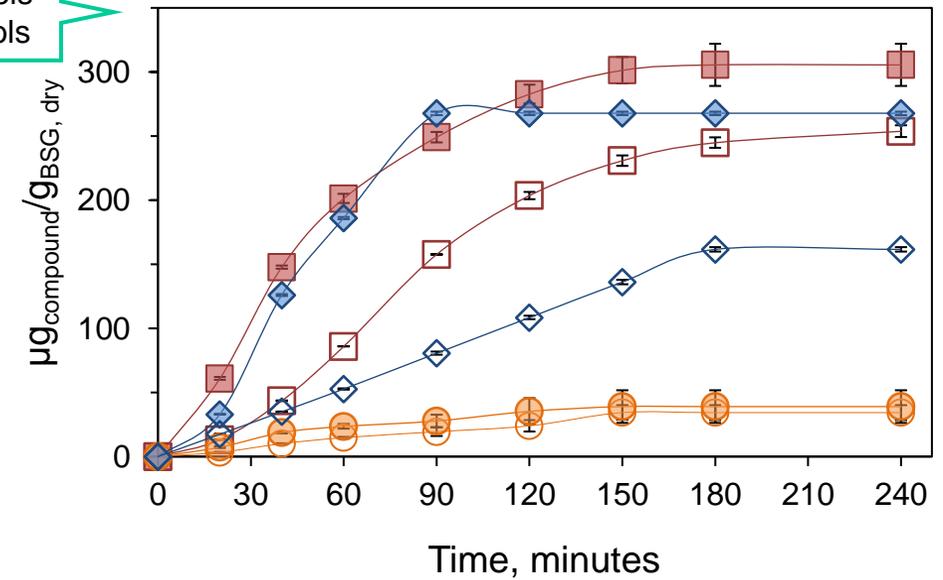
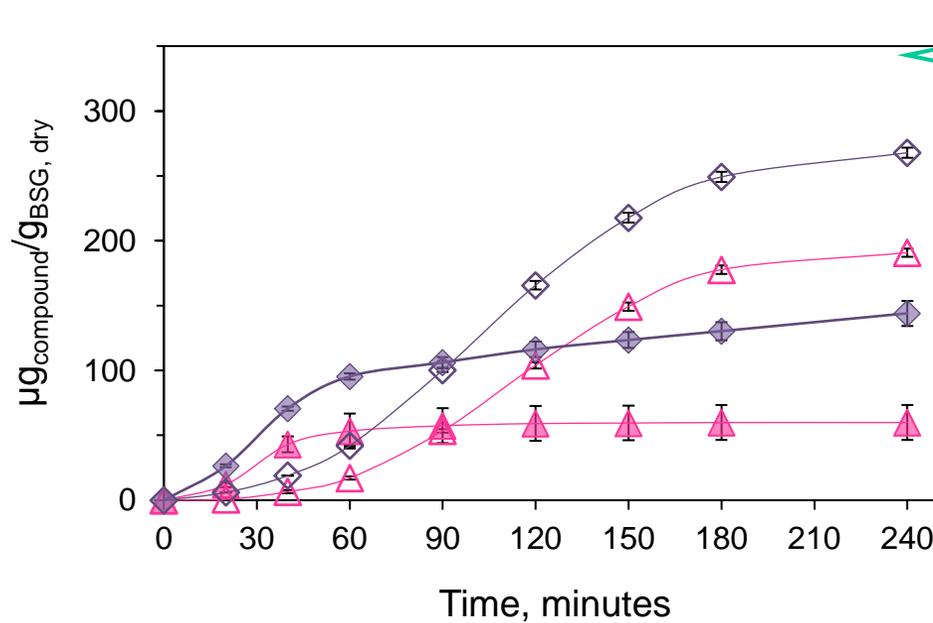
■ Vanillin



◆ Protocatechuic aldehyde



● Syringic aldehyde

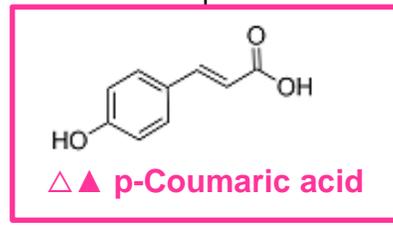
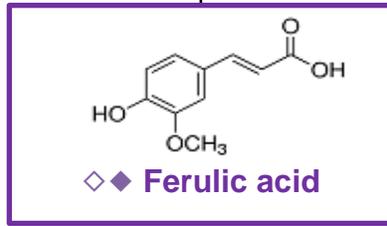


Accumulative release of the identified individual phenolic compounds in the subW extracts

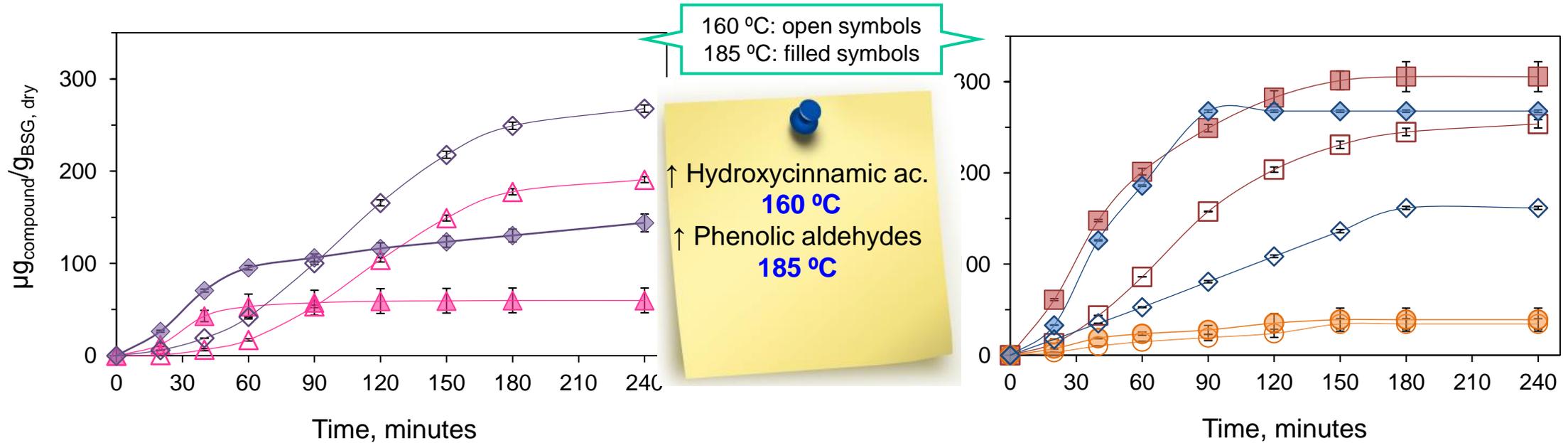
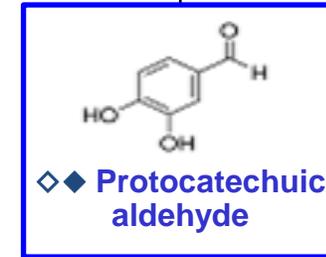
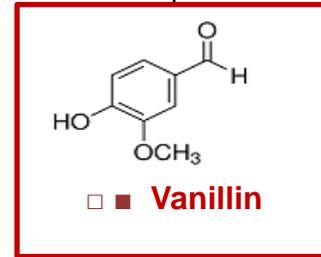


Individual Phenolic Compounds

Hydroxycinnamic acids



Aldehydes

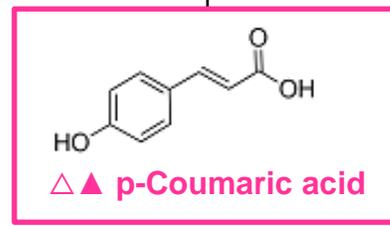
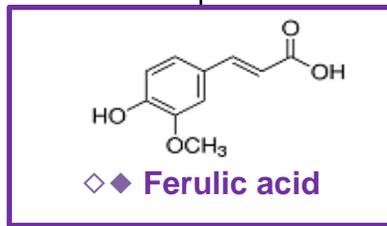


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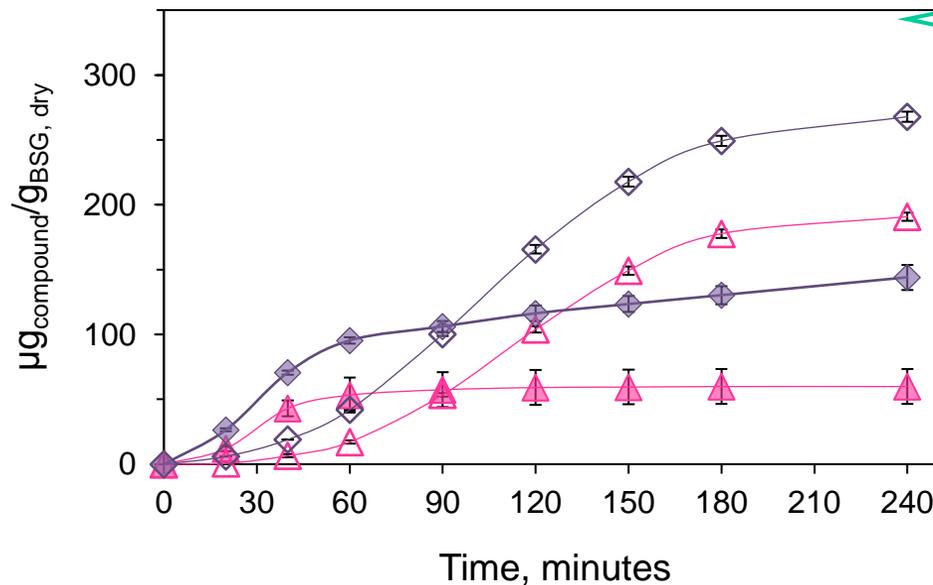
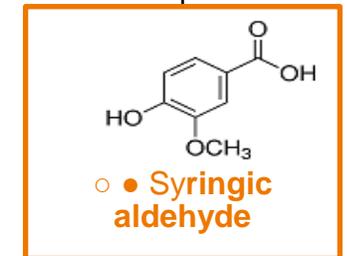
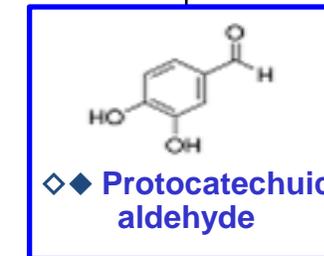
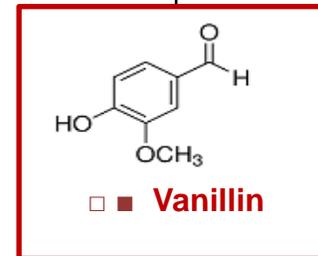


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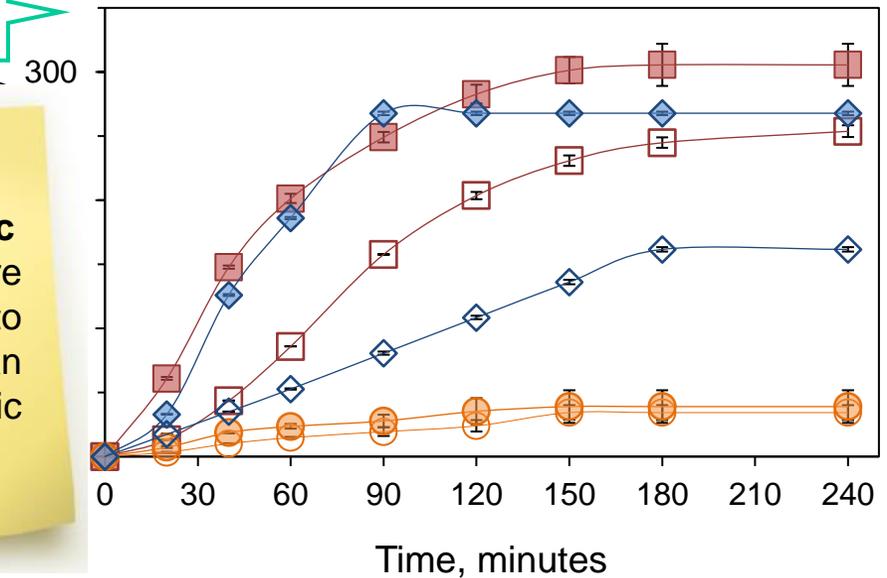


Aldehydes



160 °C: open symbols
185 °C: filled symbols

Hydroxycinnamic acids were more sensitive to temperature than aldehyde phenolic compounds.



Accumulative release of the identified individual phenolic compounds in the subW extracts



04 - RESULTS

Individual Phenolic Compounds

Comparison between the different hydrolytic methods

Phenolic compounds identified by HPLC-DAD ($\mu\text{g}_{\text{compound}}/\text{g}_{\text{BSG,dry}}$)

Compound	Formula	SubW T= 160 °C	SubW T= 185 °C	UAE-W T= 50 °C	Acid-Hy	Alkaline-Hy	Xylanase 6%
p-Hydroxibenzoic acid		n.d.	n.d.	10.0 ± 0.5	n.d.	59 ± 2	n.d.
Vanillic acid		17.9 ± 0.4	n.d.	n.d.	n.d.	49 ± 2	61 ± 3
Syringic acid		n.d.	n.d.	n.d.	n.d.	106.1 ± 5.7	n.d.
p-Coumaric acid		191 ± 3	60 ± 1	n.d.	n.d.	538 ± 4	5.3 ± 0.4
Vanillin		254 ± 5	306 ± 10	n.d.	n.d.	217 ± 1	203 ± 10
Ferulic acid		250 ± 3	144 ± 7	10.7 ± 0.3	54.4 ± 0.3	1305.7 ± 0.5	292 ± 3
Synapic acid		n.d.	n.d.	2.8 ± 0.2	31.1 ± 0.5	27 ± 1	15 ± 1
Protocatechuic aldehyde		162 ± 2	268 ± 1	n.d.	n.d.	n.d.	n.d.
Syringic aldehyde		34 ± 5	39 ± 2	n.d.	n.d.	n.d.	n.d.

n.d. not detected

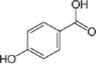
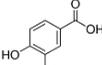
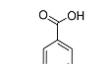
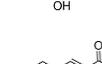
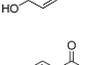
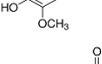
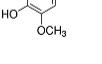
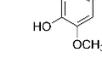
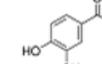


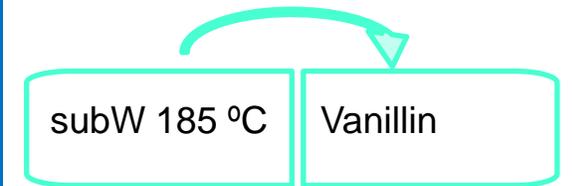
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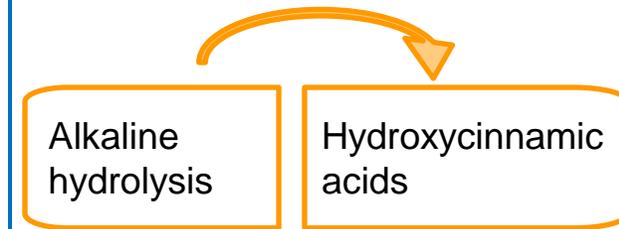
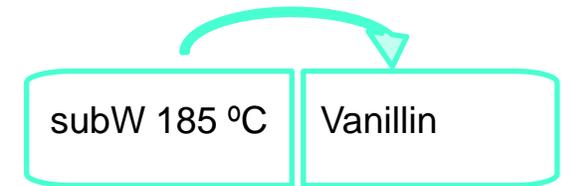
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Protocatechuic aldehyde		162 ± 2	268 ± 1	n.d.	n.d.	n.d.	n.d.
Syringic aldehyde		34 ± 5	39 ± 2	n.d.	n.d.	n.d.	n.d.



n.d. not detected





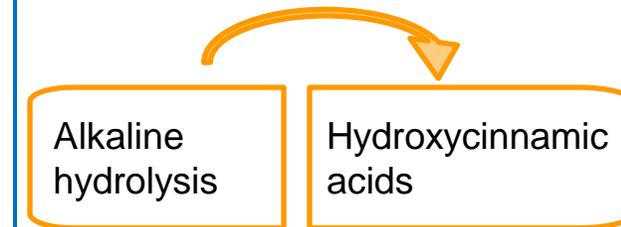
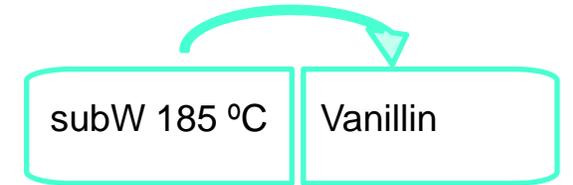
04 - RESULTS

Individual Phenolic Compounds

Comparison between the different hydrolytic methods

Phenolic compounds identified by HPLC-DAD ($\mu\text{g}_{\text{compound}}/\text{g}_{\text{BSG,dry}}$)

Compound	Formula	SubW T= 160 °C	SubW T= 185 °C	UAE-W T= 50 °C	Acid-Hy	Alkaline-Hy	Xylanase 6%
p-Hydroxibenzoic acid		n.d.	n.d.	10.0 ± 0.5	n.d.	59 ± 2	n.d.
Vanillic acid		17.9 ± 0.4	n.d.	n.d.	n.d.	49 ± 2	61 ± 3
Syringic acid		n.d.	n.d.	n.d.	n.d.	106.1 ± 5.7	n.d.
p-Coumaric acid		191 ± 3	60 ± 1	n.d.	n.d.	538 ± 4	5.3 ± 0.4
Vanillin		254 ± 5	306 ± 10	n.d.	n.d.	217 ± 1	203 ± 10
Ferulic acid		250 ± 3	144 ± 7	10.7 ± 0.3	54.4 ± 0.3	1305.7 ± 0.5	292 ± 3
Synapic acid		n.d.	n.d.	2.8 ± 0.2	31.1 ± 0.5	27 ± 1	15 ± 1
Protocatechuic aldehyde		162 ± 2	268 ± 1	n.d.	n.d.	n.d.	n.d.
Syringic aldehyde		34 ± 5	39 ± 2	n.d.	n.d.	n.d.	n.d.



Xylanase 6% vs subW:

- ↑ Ferulic acid
- ↓ p-Coumaric acid
- ↓ Aldehydes

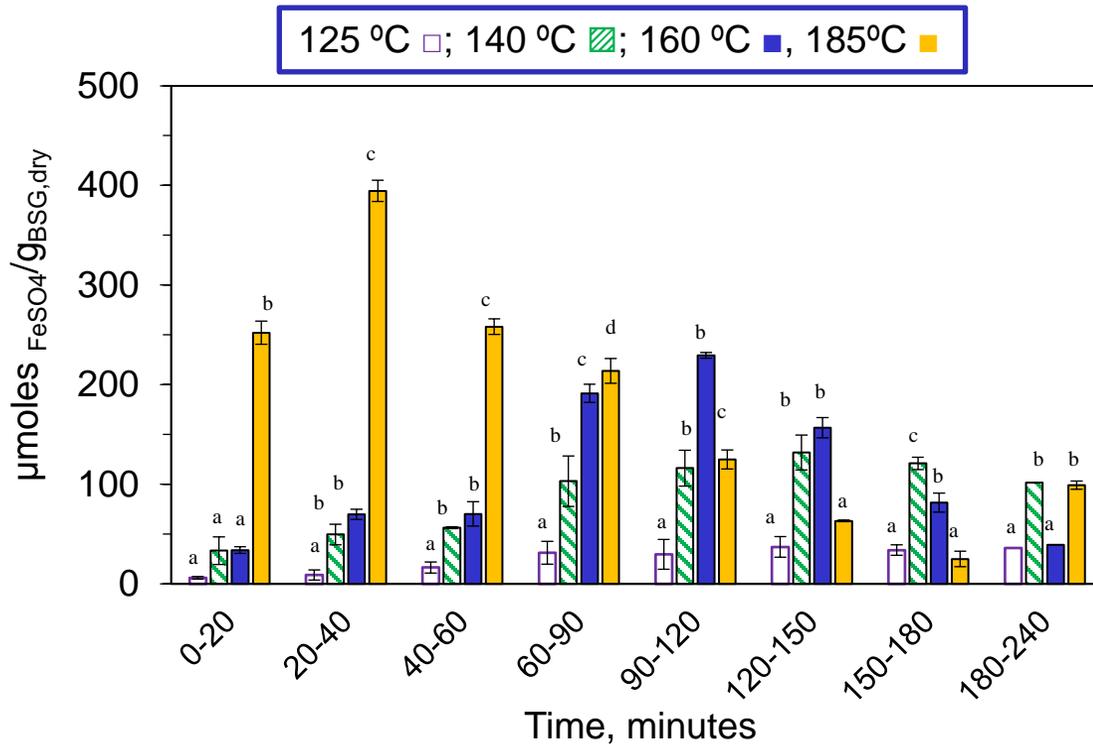
n.d. not detected



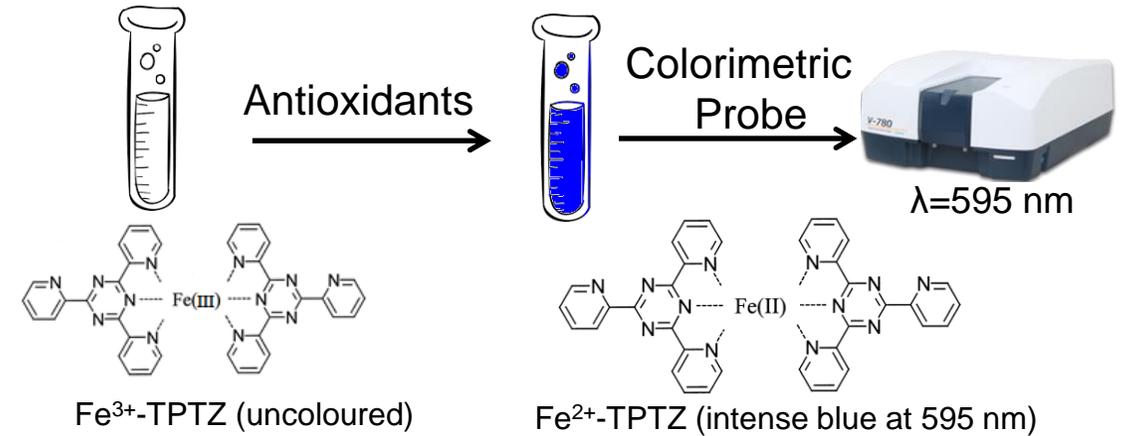


Antioxidant activity

FRAP (Ferric Reducing Antioxidant Power)



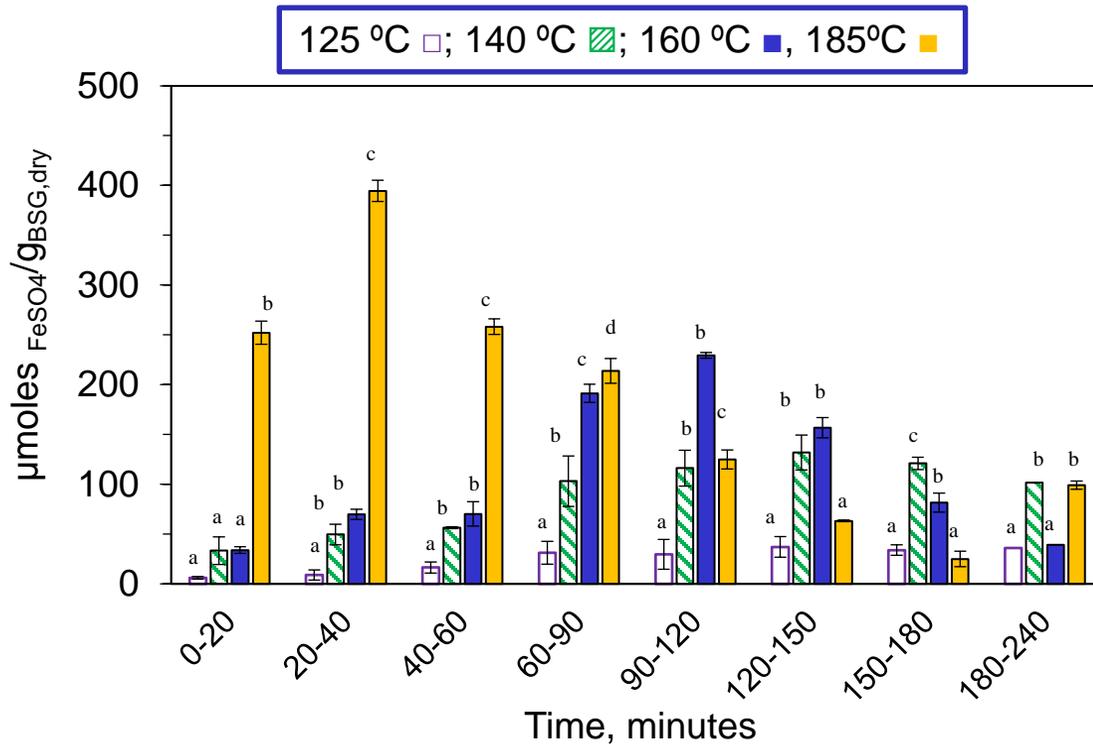
Antioxidant activity of subW extracts at different time intervals



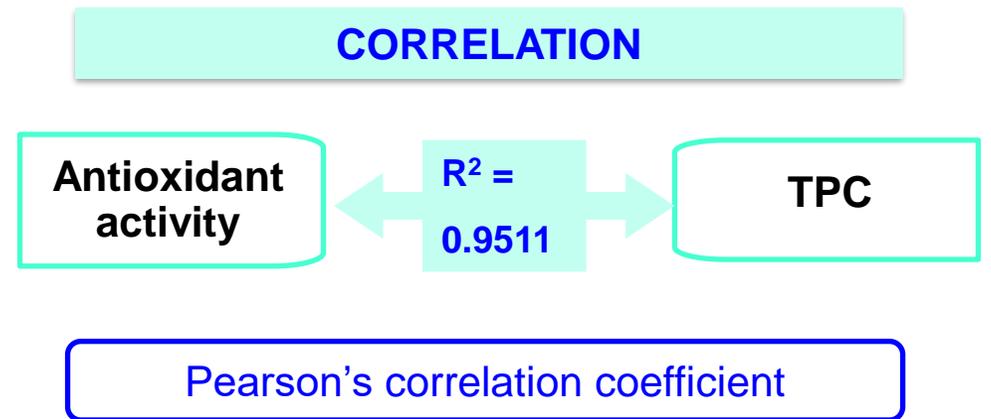
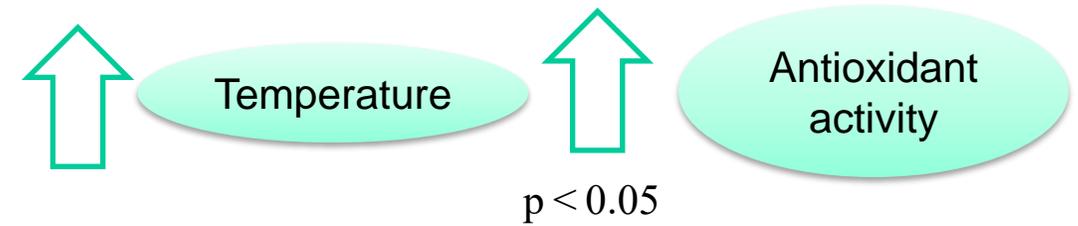


Antioxidant activity

FRAP (*Ferric Reducing Antioxidant Power*)



Antioxidant activity of subW extracts at different time intervals

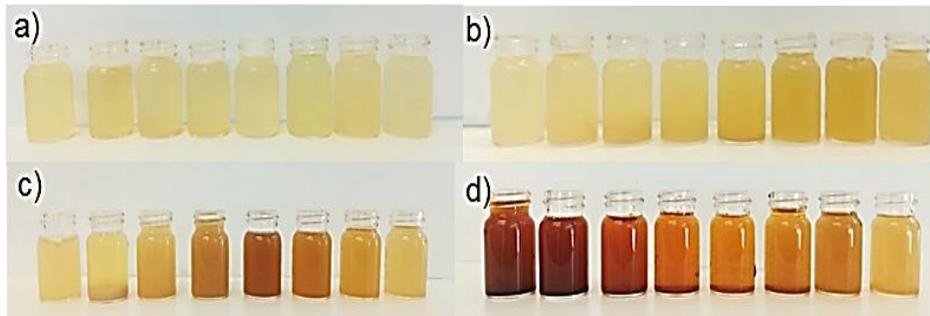




TPC-Antioxidant activity



Formation of compounds of **Maillard** reactions



- a) 125 °C
- b) 140 °C
- c) 160 °C
- d) 185°C

The extracts become darker by $\uparrow T$

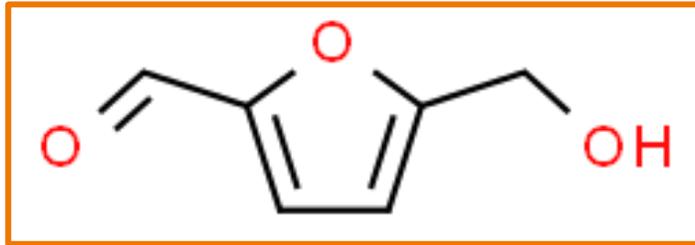
The colour change would be due to the formation of **hydrolysis** and **decomposition** products such as HMF and Furfural



04 - RESULTS

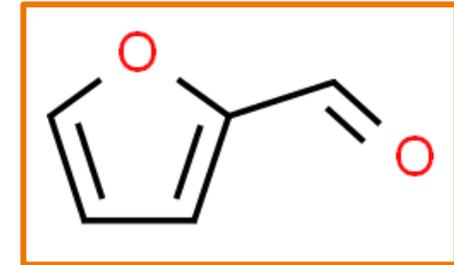
Hmf and Furfural

Concentration of HMF and furfural in subW extracts

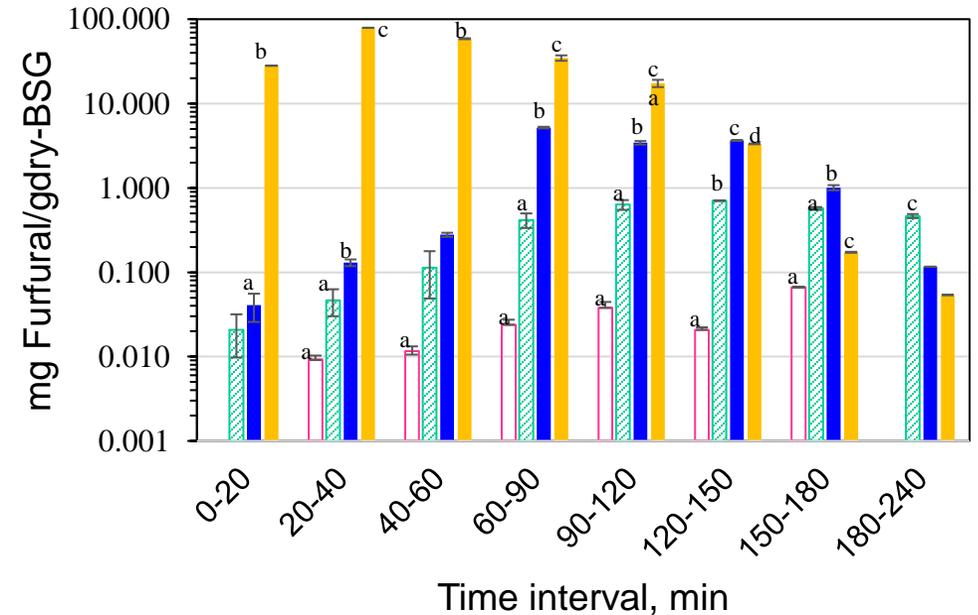
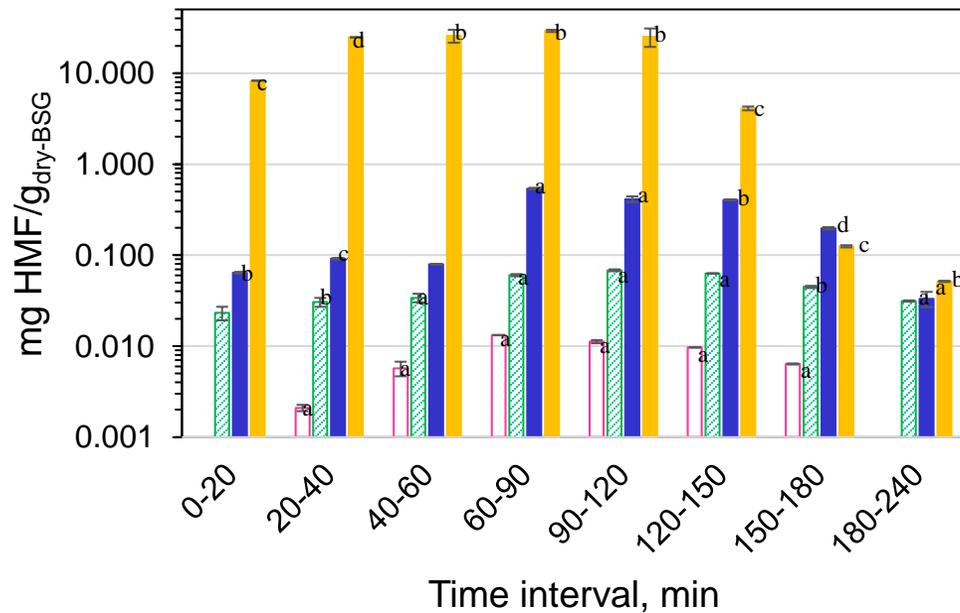
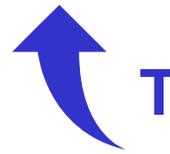


5-Hydroxymethyl furfural (HMF)

Take part in reactions leading to the formation of melanoidins and other polymers and aromatic substances



Furfural

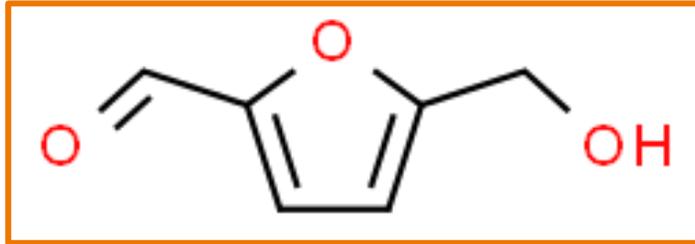


*Values with different letters at each interval time are significantly different when applying the Fisher's least significant differences (LSD) method at p-value ≤ 0.05.

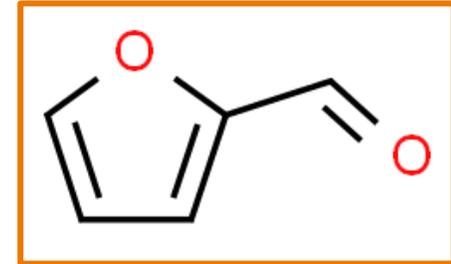


Hmf and Furfural

Concentration of HMF and furfural in subW extracts



5-Hydroxymethyl furfural (HMF)



Furfural

CORRELATION

Pearsons correlation coefficients

Antioxidant activity

$R^2 = 0.6887$

HMF

Antioxidant activity

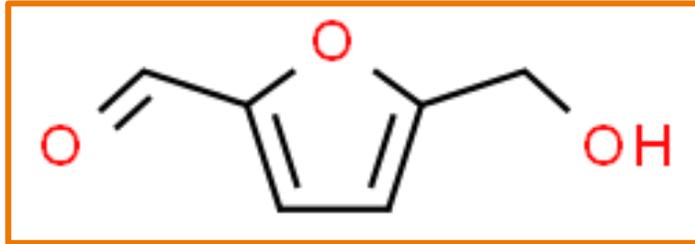
$R^2 = 0.8091$

Furfural

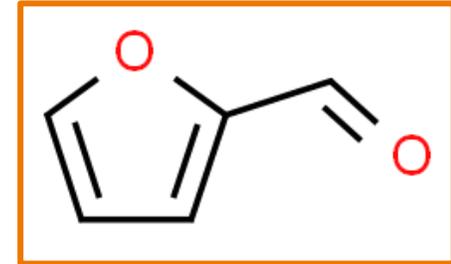


Hmf and Furfural

Concentration of HMF and furfural in subW extracts

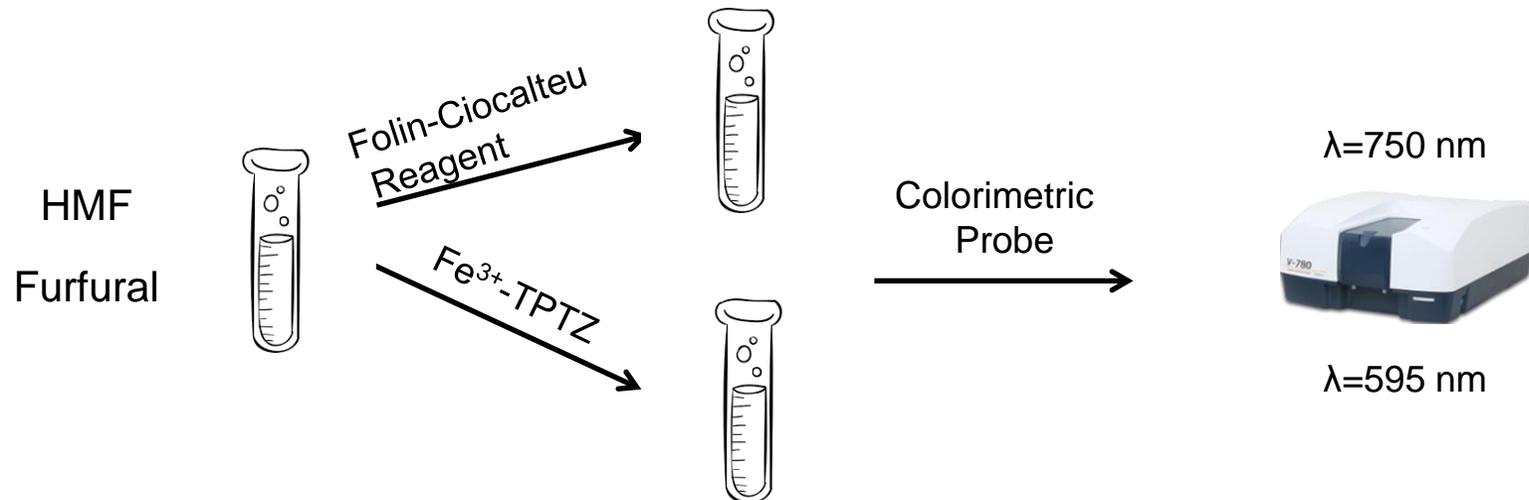


5-Hydroxymethyl furfural (HMF)



Furfural

TPC AND ANTIOXIDANT ACTIVITY ASSAYS





Antioxidant activity- Proteins



CORRELATION

Pearsons correlation coefficients

Antioxidant activity

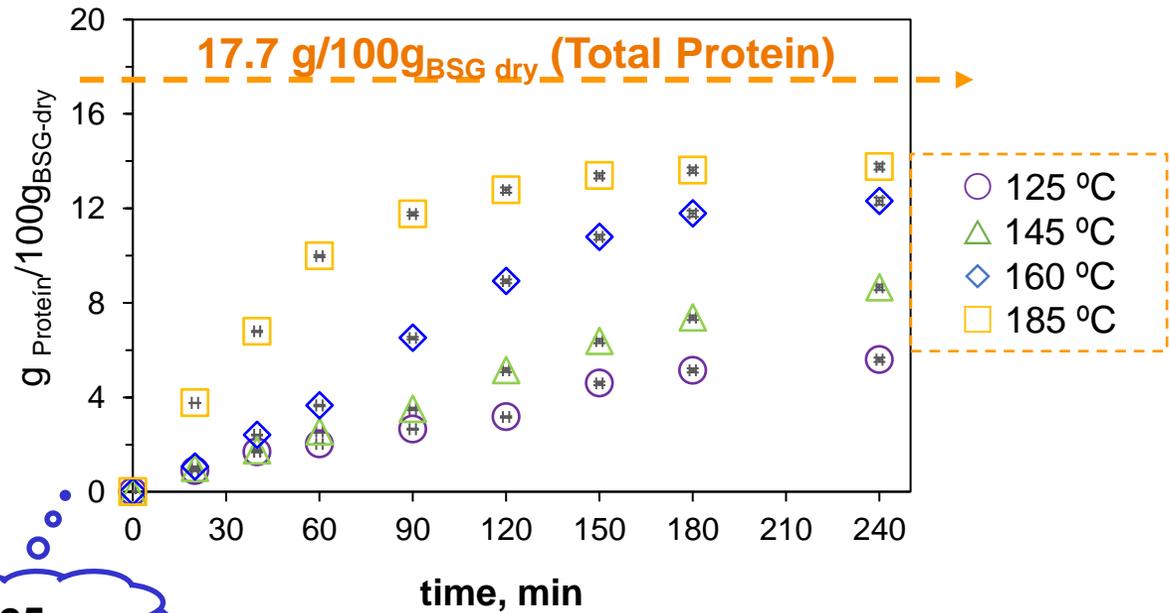
$R^2 = 0.7629$

Proteins

$$\% \text{ Yield} = \frac{\text{Protein in the subW extract}}{\text{Protein in dry BSG}} \cdot 100$$

T, °C	Yield, %
185	77.7
160	69.5
145	52.6
125	31.6

Accumulative total protein fraction in the subW extracts.



TN-6.25



04 - RESULTS

Antioxidant activity- Free amino acids

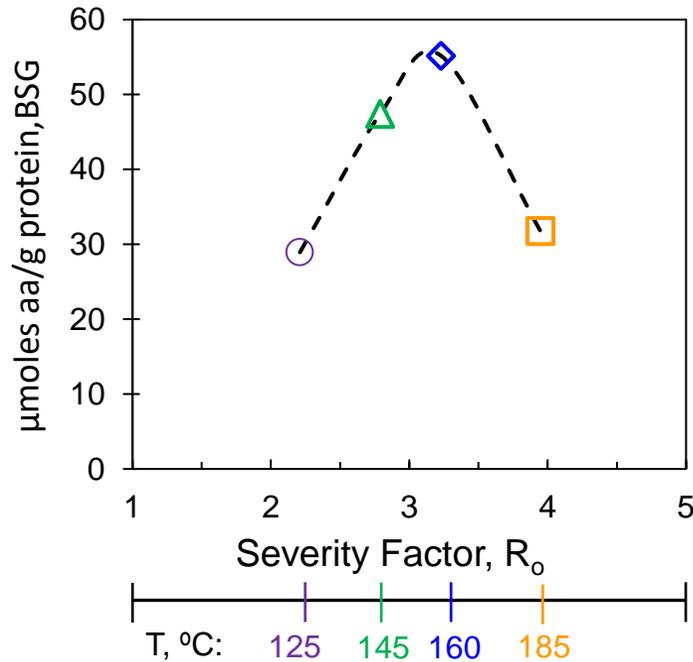
CORRELATION

Pearsons correlation coefficients

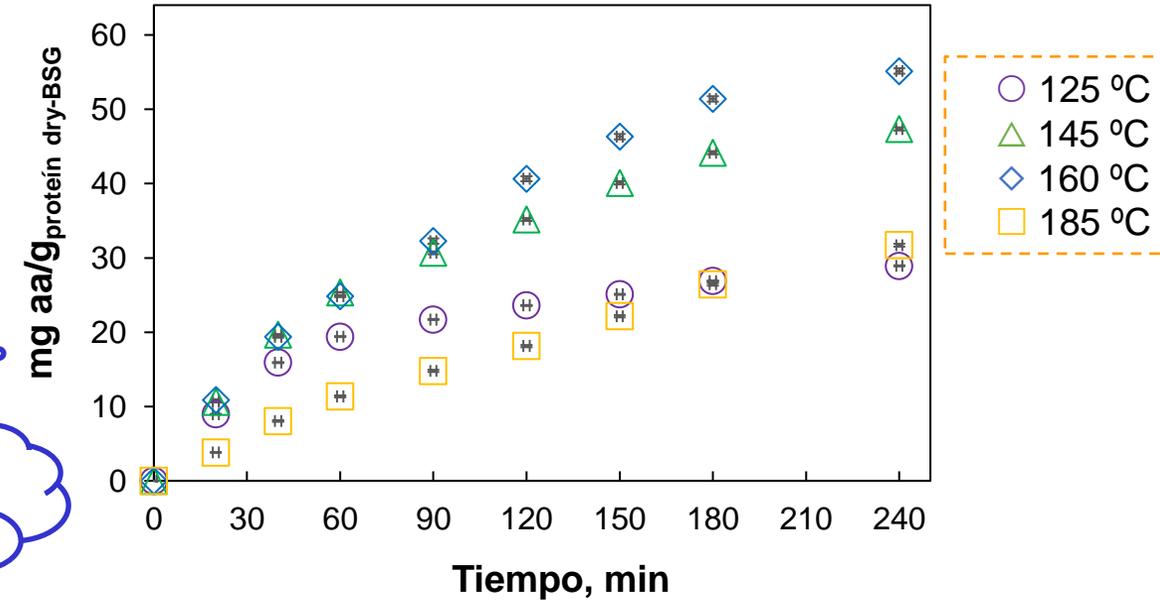
Antioxidant activity

$$R^2 = 0.1189$$

Free amino acids



Accumulative profile in the subW extracts of free amino acids



Sum of aa by GC

High residence time: $\tau \approx 29$ min

Highest level at 160 °C aa degradation for $R_o > 3.2$

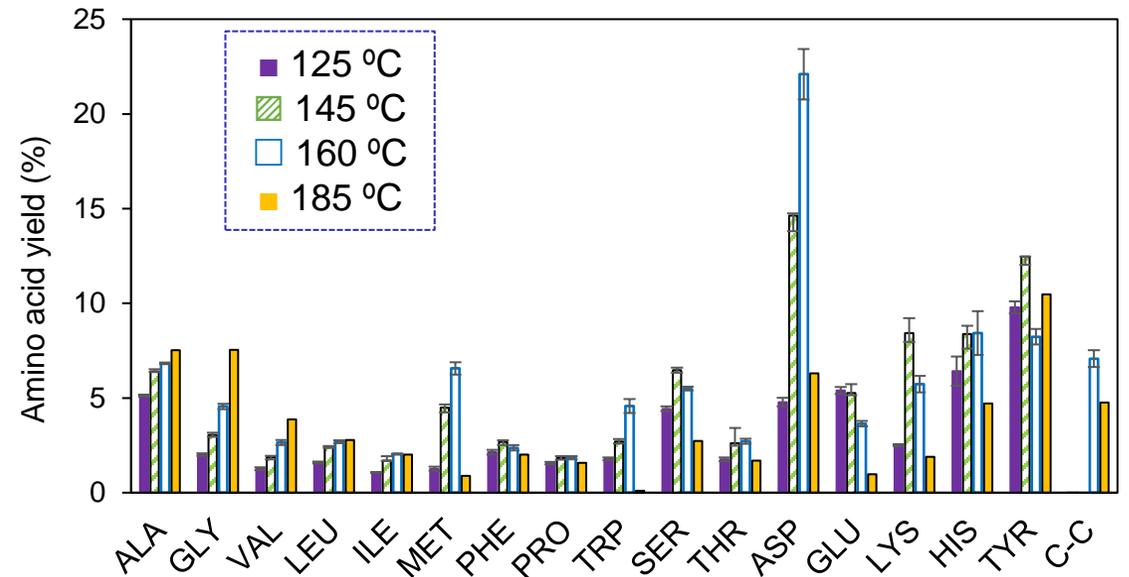


Free amino acids

Amino acid profile of BSG and subW extracts (mg_{aa}/g_{protein-dry-BSG})

Amino acid	BSG	125 °C	145 °C	160 °C	185 °C
ALA	48.1	2.46	3.08	3.29	3.62
GLY	41.9	0.84	1.28	1.91	3.16
VAL	122.6	1.56	2.30	3.25	4.76
LEU	87.1	1.39	2.13	2.35	2.43
ILE	69.3	0.74	1.18	1.42	1.40
THR	41.1	0.73	1.07	1.12	0.70
SER	44.1	1.96	2.86	2.43	1.21
PRO	123.2	1.93	2.25	2.28	1.94
ASP	69.5	3.32	10.17	15.35	4.38
MET	19.0	0.24	0.85	1.25	0.17
HYP	4.3	0.00	0.14	0.10	0.02
GLU	116.8	6.30	6.15	4.26	1.15
PHE	68.0	1.46	1.83	1.61	1.37
LYS	82.7	2.08	6.98	4.74	1.57
HIS	22.8	1.47	1.92	1.93	1.08
HYL	0.0	0.00	0.00	5.03	0.17
TYR	22.5	2.20	2.80	1.85	2.35
TRP	14.7	0.26	0.40	0.67	0.02
C-C	4.7	0.00	0.00	0.33	0.22
TAA	1002.4	28.9	47.4	55.2	31.7

$$\text{Yield (\%)} = \frac{\text{mg } aa_T}{\text{mg } aa_{BSG}} \cdot 100$$



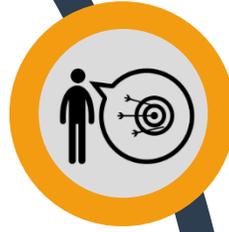
Final yield of individual amino acids as a function of temperature

↑ Aspartic Acid at 160 °C

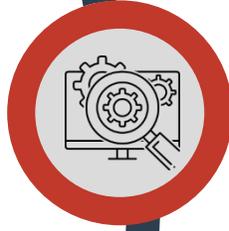
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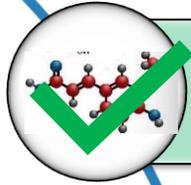


05 - CONCLUSIONS





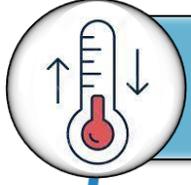
05 - CONCLUSIONS



SubW treatment was confirmed as an efficient extraction/hydrolysis method to extract phenolic compounds from BSG.



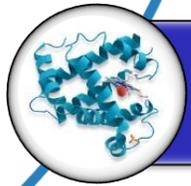
Extraction of phenolic compounds was maximized at the highest temperature studied in this work, 185 °C.



Hydroxycinnamic acids were more sensitive to temperature than aldehyde phenolic compounds.



SubW extracts showed a high antioxidant activity that increased with temperature.



A high protein yield extraction of 78 % was achieved at 185 °C.

THANKS FOR YOUR ATTENTION



Research group "Industrial and Environmental Biotechnology" (BIOIND)

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