

1 **Antioxidant effect of seasonings derived from wine pomace in**
2 **refrigerated and frozen beef patties**

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11 **ABSTRACT**

12 The food industry is constantly looking for natural solutions to retard lipid
13 oxidation and to extend the shelf-life of foodstuffs. Beef patties made with three natural
14 seasonings derived from red wine pomace showed higher stability against lipid oxidation
15 than control patties stored under refrigeration conditions in a high-oxygen atmosphere.
16 The seasoning produced from seedless wine pomace presented the highest activity in
17 comparison with the other seasonings. It showed higher efficacy than sulfites in delaying
18 the formation of thiobarbituric acid reactive substances (TBARS) and other volatile
19 organic compounds related to lipid oxidation. The seasoning also retarded the
20 development of lipid oxidation in cooked and raw beef patties stored under freezing
21 conditions. In the frozen samples, sulfites presented a contradictory effect, since it
22 inhibited hexanal formation, but promoted the formation of TBARS. The results of the
23 present study reveal that the seasoning is able to limit the development of lipid
24 oxidation, extending the shelf-life of meat products without using synthetic antioxidants.

25 **KEYWORDS: vegetal seasoning, lipid oxidation, wine pomace, beef patties, sulfites**

26 **1. INTRODUCTION**

27 Lipid oxidation is one of the major factors that limits the shelf-life of meat
28 products. It involves the formation of hydroperoxides that are easily broken down to
29 form low molecular weight compounds (Kanner, 1994; Zhao, Wells, & McMillin,
30 1994). Different volatile organic compounds (VOCs) such as alkanes, alkenes,
31 aldehydes, ketones, alcohols, esters and acids, are extensively formed, causing rancid
32 and unpleasant flavors and reducing the sensorial quality of meat products (Frankel,
33 1983; Kanner, 1994). Furthermore, lipid oxidation decreases the nutritional value of
34 foodstuffs due to the loss of essential compounds such as polyunsaturated fatty acids
35 (Kanner, 1994). Meat products are affected by lipid oxidation, as they usually contain
36 high levels of fat and pro-oxidants such as salt. Furthermore, processes such as grinding
37 or cooking also decrease the stability against lipid oxidation, due to structural
38 degradation and the release of pro-oxidants (Alfawaz, Smith, & Jeon, 1994; Kanner,
39 1994).

40 Nowadays, meat products for storage under refrigerated conditions are usually
41 packaged under high-oxygen atmospheres (70-80% O₂, 20-30% CO₂). This type of
42 atmosphere maintains the attractive bright red color associated with fresh meat and
43 present antimicrobial effect. However, storage with high levels of oxygen also
44 accelerates the process of lipid oxidation and the appearance of off-flavors (Zhao et al.,
45 1994).

46 The development of a global meat market and the increase of distance between
47 producers and consumers have increased the use of freezing as a preservation technique
48 (Leygonie, Britz, & Hoffman, 2012). Low temperatures and low levels of available
49 water drastically reduce microbial and chemical reactions, thereby extending the shelf-
50 life of these products. However, freezing also modifies the homeostasis of the meat

51 system, due to the cryoconcentration of solutes in the unfrozen phase. These changes
52 may damage cell membrane leading to leakage of intracellular pro-oxidants such as
53 lipases and metals, affecting the chemical stability of the product (Leygonie et al.,
54 2012). Moreover, the catering and retail sector is increasingly interested in ready-to-eat
55 foods that can be stored over long periods. Some consumers also prefer cooked products
56 due to their convenience and their shorter preparation times. However, storage of
57 cooked products is usually linked to extensive lipid oxidation and the development of
58 the unpleasant “warmed-over flavor” (Carpenter, O’Grady, O’Callaghan, O’Brien, &
59 Kerry, 2007; Igene, Pearson, Merkel, & Coleman, 1979).

60 Several chemical additives can be used to inhibit lipid oxidation and
61 consequently extend the shelf-life of meat products (Cornelius & Lilian, 2005). For
62 instance, sulfites are mainly applied as antimicrobial agents but also exert an important
63 function as antioxidant. However, their consumption has been linked to allergic
64 reactions and safety concerns over their long-term consumption have yet to be clarified
65 (Cornelius & Lilian, 2005). Moreover, consumer rejection of chemical additives is
66 growing and the food industry is constantly looking for natural additives. Natural
67 products, such as spices and plant extracts, have been proposed as natural antioxidants
68 to replace the use of chemical additives (Brewer, 2011; Lindberg Madsen & Bertelsen,
69 1995; Yanishlieva, Marinova, & Pokorný, 2006). Due to the complex reactions involved
70 in lipid oxidation of foodstuffs, the efficacy of natural antioxidants may be affected
71 under different storage conditions (Lindberg Madsen & Bertelsen, 1995). For instance,
72 different herbs and spices such as curry and cinnamon showed high antioxidant
73 protection in refrigerated storage of minced chicken meat, but not at the frozen storage
74 (El-Alim, Lugasi, Hóvári, & Dworschák, 1999). Then, if their full potential is to be
75 evaluated, natural antioxidants should be tested under different conditions.

76 Different seasonings recently developed from wine pomace presented excellent
77 properties to be used as food ingredients. The seasonings are rich in fiber, minerals,
78 especially potassium, and in phenolic compounds, including extractable and non-
79 extractable fractions such as catechins, anthocyanins and proanthocyanidins, among
80 other phenols (García-Lomillo, González-SanJosé, Del Pino-García, Rivero-Pérez, &
81 Muñoz-Rodríguez, 2014). They have successfully been applied in different food
82 matrices and have shown a preservative effect in low salt patties with high levels of
83 consumer acceptance (González-SanJosé et al., 2014). Furthermore, the seasonings
84 were found to successfully inhibit lipid oxidation under accelerated oxidative conditions
85 in the Rancimat test, which points out to their potential use as natural antioxidants in the
86 food industry (García-Lomillo et al., 2014).

87 The main aim of this work is to study the potential of red wine pomace
88 seasonings (RWPSs) to retard or inhibit lipid oxidation in beef patties under different
89 storage conditions (raw refrigerated under high-oxygen atmosphere as well as raw and
90 cooked frozen vacuum-packaged patties), comparing the antioxidant effect with the
91 antioxidant capacity of sulfites, a common additive in meat patties.

92 **2. MATERIALS AND METHODS**

93 **2.1. Material**

94 Beef meat was obtained from a local supplier (Gros Mercat, Burgos, Spain).
95 Common salt, food grade starch and a commercially available mixture of phosphates
96 provided by Doscadesa (Murcia, Spain) and sodium metabisulfite ($\text{Na}_2\text{S}_2\text{O}_5$) (Panreac,
97 Barcelona, Spain) were used in the formulation of beef patties. The seasonings under
98 study were prepared at the pilot plant of the Food Technology Department of the
99 University of Burgos (Spain), following the process described by González San José,
100 García Lomillo, Del Pino García, Dolores Rivero, & Muñoz Rodríguez (2015). The

101 seasoning obtained directly from whole red wine pomace was labeled as WRWPS
102 (whole red wine pomace seasoning). A second seasoning was obtained from seedless
103 red wine pomace and was labeled as SkRWPS (skin red wine pomace seasoning). The
104 third seasoning, obtained from the seeds that had been separated was labeled SdRWPS
105 (seed red wine pomace seasoning). The chemical and phenolic composition as well as
106 their antimicrobial capacity of the three seasonings were previously described (García-
107 Lomillo et al., 2014).

108 For lipid oxidation assessment, perchloric acid was purchased from VWR
109 International (Barcelona, Spain), and 2-thiobarbituric acid, cyclopentanone,
110 dichloromethane, hexanal, 1,1,3,3-tetraethoxypropane were purchased from Sigma (St.
111 Louis, USA).

112 **2.2.Patty preparation**

113 Control beef patties were prepared by mixing 920 g of previously chopped and
114 minced meat, 12 g of potato starch and 50 mL of water in which 3 g of a commercially
115 available mix of food grade phosphates had previously been dissolved, and the
116 corresponding quantity of salt to obtain a final concentration of 1.5 g of salt per kg of
117 patty. Beef patties with seasonings were formulated in the same way, but suspending the
118 corresponding amount of seasoning in water to obtain a final concentration of 2 %
119 (w/w). Sulfite samples were similarly prepared obtaining a final concentration of 300
120 mg of SO₂ per kg of patty (300 ppm). Ingredients were mixed in a food processor for 5
121 minutes and patties of 100 grams were manually formed and packaged or cooked and
122 then packaged. Cooked samples were processed according to the procedure described
123 by Carpenter, et al. (2007) in a fan-assisted oven (Berto's, Padova, Italy) previously
124 heated at 180°C, until an internal temperature of 72°C was reached (8 minutes) and
125 subsequently maintained for 8 minutes.

126 Raw patties were stored under two different conditions, high-oxygen atmosphere
127 (70% O₂/30% CO₂) and refrigeration, and vacuum packing and freezing. In the high-
128 oxygen atmosphere, samples were packaged in trays of polyethylene/ethylene vinyl
129 alcohol/polystyrene (Sanviplast, Barcelona, Spain) with permeability to oxygen of 0.99
130 cm³/(m² day atm). They were then sealed using a polyethylene terephthalate
131 polyvinylidene chloride/polyethylene film with an oxygen permeability of 7 cm³/(m²
132 day atm) (Ampcor, Burgos, Spain). The vacuum-packaged samples (raw and cooked),
133 were sealed in polyamide/polyethylene bags (20/70 μm) (Vacioplast, Salamanca, Spain)
134 with an oxygen permeability lower than 40 cm³/(m² day atm). The samples were frozen
135 using a conventional freezer working at -30 °C stored over 6 months at -18°C.

136 Sampling of the refrigerated patties (conserved at 4°C) was conducted at days 0,
137 4, 8, 12 and 15 days of storage, and at months 0, 2, 4 and 6 of the frozen samples.
138 Frozen samples were thawed at 4°C for 12 hours before analysis. This study was
139 performed in duplicate and, in each experiment, three independent replicates (trays or
140 bags) were considered on each sampling date.

141 The chemical composition of the beef patties was analyzed using a FoodScan™
142 near-infrared spectrophotometer (Foss Electric A/S, Hillerød, Denmark) and data
143 processing done with ISIScan™ Software.

144 **2.3.Thiobarbituric acid reactive substances (TBARS) analysis**

145 TBARS method was conducted according to the method proposed by Tarladgis,
146 Watts, Younathan, & Dugan (1960) with minor modifications. Briefly, samples were
147 homogenized in 3.86% perchloric acid, filtered and distilled. Three mL of the resulting
148 distillate were mixed with 1 mL of 0.04 M of 2-thiobarbituric acid in perchloric acid
149 (10%) and incubated at 100°C for 45 minutes. After incubation, samples were cooled

150 and their absorbance was measured at 532 nm using a U-2000 Hitachi
151 spectrophotometer (Tokyo, Japan) against a blank where sample was replaced with 3
152 mL of perchloric acid. Quantification was conducted by preparing a standard curve with
153 tetraethoxypropane and the results were expressed in ppm of malondialdehyde (MDA).
154 The quantification limit (0.08 ppm of MDA) of the method was calculated using the
155 DETARCHI software (Sarabia & Ortiz, 1994).

156 **2.4. Analysis of volatile organic compounds (VOCs)**

157 Volatile organic compounds were evaluated by a headspace solid-phase dynamic
158 extraction (HS-SPDE) coupled with gas chromatography–mass spectrometry (GC-MS)
159 (Agilent Technologies 6890N) fully controlled by a CTC-CombiPAL autosampler
160 (Bender and Hobein, Zurich, Switzerland), following the method optimized by
161 Corcuera-Tecedor (2013) Samples were introduced in a glass vial with 10 mL of
162 headspace (Chromacol Ltd. Herts, United Kingdom), mixed with 5 µl of a
163 cyclopentanone:dichloromethane (1:9) solution and sealed using a metallic cap with a
164 chlorobutyl/polytetrafluoroethylene seal (Chromacol Ltd. Herts). Extraction was
165 conducted in an incubation station at 55°C, using a previously conditioned SPDE-
166 syringe with a non-polar PDMS/AC (90% polydimethylsiloxane and 10% activated
167 carbon) needle (60 extraction strokes). After extraction, VOCs were then desorbed from
168 the fiber in the injection port at 250°C, using helium as the carrier agent. Separation was
169 conducted in 007-WAX capillary column (Quadrex Corporation, New Haven, USA) (60
170 m length, 0.32 mm inside diameter and 1µm film thickness). Compounds were
171 identified by comparing their mass spectra with those found in the Wiley 7th and NIST
172 98 libraries and confirmed with their retention times. The retention index was calculated
173 in relation to a series of standard alkanes (C6-C18) that were used for calculating
174 Kovats indexes. The results were expressed as relative areas of each chromatographic

175 peak (compounds) in relation to the peak area of the internal standard (arbitrary units,
176 AU) that was used.

177 **2.5.Statistical analysis**

178 Statistical analysis was performed using StatGraphics ® Centurion XVI. Fisher´s
179 least significant difference test was performed in order to identify significant differences
180 between different formulations and at different days of storage (p-value < 0.05).

181

182 **3. RESULTS AND DISCUSSION**

183 **3.1.Chemical analysis of beef patties**

184 The main compositional parameters of the raw and the cooked samples were
185 measured and significant differences were not observed between the different batches.
186 Raw samples averaged $63.0 \pm 1.3\%$ of moisture, $16.2 \pm 0.3\%$ of total protein and $13.4 \pm$
187 0.9% of total lipid. Cooked samples presented values of $64.3 \pm 1.3\%$ moisture, $23.8 \pm$
188 1.2% of protein and $9.9 \pm 0.9\%$ of total lipid content.

189 **3.2.Lipid oxidation under high-oxygen atmosphere and refrigeration** 190 **conditions**

191 TBARS analysis is a widely used indicator to evaluate secondary lipid oxidation
192 during the storage of meat products. It is usually considered a measure of the MDA
193 content, although other aldehydes may also participate in the reaction (Kanner, 1994).

194 Previous studies reported that SkRWPS showed a higher antioxidant effect in the
195 Rancimat test in comparison to SdRWPS and WRWPS (García-Lomillo et al., 2014).
196 This study also showed different inhibition capacities for each seasoning (Figure 1).

197 TBARS values of the control patties increased during refrigerated storage, due
198 to the lipid instability of meat products stored under a high-oxygen atmosphere

199 (Frankel, 1983). Patties are minced meat products with high fat and relatively high salt
200 contents, factors that enhance lipid oxidation (Alfawaz et al., 1994; Kanner, 1994).
201 These facts could explain the extensive formation of TBARS not only during storage
202 but also during patty preparation. The three seasonings under study were able to prevent
203 lipid oxidation during patty processing and subsequent storage, but with different levels
204 of efficacy. SkRWPS was the most effective inhibiting lipid oxidation completely until
205 the end of storage (two weeks). WRWPS and SdRWPS were also able to delay the
206 onset of lipid oxidation for around one week and to reduce the oxidation rate
207 significantly. Then, patties made with these seasonings showed TBARS values that
208 were 55% lower than the control patties, and no significant differences were observed
209 between them.

210 In view of the high effectiveness of SkRWPS against lipid oxidation of patties,
211 further experiments were conducted to compare the effect of this seasoning with sulfites
212 (Figures 2a and 2b). The results obtained in the two assays showed that sulfite was less
213 efficient than SkRWPS, which totally inhibited TBARS formation. Sulfites were not
214 able to retard the onset of the oxidation, but they were able to reduce the intensity of
215 lipid oxidation, and lower TBARS values than in control samples were obtained on all
216 sampling dates. At the end of storage, formulation with sulfites presented 55% lower
217 TBARS values than the control samples.

218 Besides the information provided by TBARS value, which quantifies those
219 aldehydes (mainly MDA) that react to thiobarbituric acid, other low molecular weight
220 compounds, formed by the breakdown of the fatty acids, are also considered good
221 indices of this process. Among these Volatile Organic Compounds (VOCs), saturated
222 aldehydes and especially hexanal are extensively recognized as indicators of the lipid
223 oxidation. They play a major role in the development of rancid odors, due to their

224 extensive formation and their relatively low threshold (Calkins & Hodgen, 2007). Other
225 VOCs of interest are alcohols, 2,3-octanedione and 2-pentylfuran which also contribute
226 to the development of rancid off-odors (Calkins & Hodgen, 2007; Gravador et al.,
227 2015).

228 Different VOCs were detected in all the patties, although they were mainly
229 present in control patties, and, there was a general increase throughout the storage
230 period (Figures 3 and 4). Hexanal content of control samples increased notably
231 throughout the storage and similar results were obtained in the two assays carried out
232 (Figure 3A and 3B). This result indicated an advanced stage of lipid oxidation, in
233 agreement with trends of TBARS values for these samples. These results coincide with
234 previous studies reporting high correlations between TBARS values and hexanal
235 formation (Frankel, 1983; Kanner, 1994). In agreement with hexanal levels, significant
236 increases of levels of 1-pentanol, 1-hexanol, 1-octen-3-ol, 2,3-octanedione and 2-
237 pentylfuran were also detected in control samples (Figure 4), corroborating the
238 development of secondary lipid oxidation.

239 Patties made with sulfites showed lower levels of hexanal and the other analyzed
240 VOCs in comparison to control patties (Figures 3 and 4), which is in agreement with the
241 antioxidant activity of this additive (Günther, König, Habicher, & Schwetlick, 1997).
242 The antioxidant activity of sulfites is usually linked to their capacity to reduce lipid
243 hydroperoxides without producing radicals (Günther et al., 1997). Moreover, reactive
244 oxygen species (ROS) can also be scavenged by sulfites leading to radical termination
245 processes (Kaneda, Takashio, Osawa, Kawakishi, & Tamaki, 1996). It might also
246 involve the reduction of other pro-oxidants such as H₂O₂ and the ability of sulfites to
247 avoid the accumulation of protein radicals that could act as lipid pro-oxidants (Garcia-
248 Lomillo, González-SanJosé, Skibsted, & Jongberg, 2016).

249 According to TBARS results, SkRWPS patties showed no hexanal except for
250 very low levels at the end of storage. The seasoning was also able to inhibit the
251 formation of 1-octen-3ol, and to reduce the formation of the other VOCs (Figure 4).
252 These results showed higher effectiveness of SkRWPS than sulfites, pointing out the
253 high potential of the new seasoning as an antioxidant in beef patties stored under a high-
254 oxygen atmosphere.

255 The results obtained agree with those reported by other authors. Sáyago-Ayerdi,
256 Brenes, & Goñi, (2009) reported that grape antioxidant dietary fiber was capable of
257 inhibiting lipid oxidation in chicken hamburger and Garrido, Auqui, Martí, & Linares
258 (2011) observed the protective effect of different grape pomace extracts in pork burgers
259 stored under aerobic conditions. Herbs and spices have also reported antioxidant
260 activity in refrigerated meat products (Lindberg Madsen & Bertelsen, 1995; Yanishlieva
261 et al., 2006). El-Alim, et al.(1999) tested different spices observing that the highest
262 inhibition was produced by clove, nutmeg, curry and cinnamon. However, the inhibition
263 was lower than the protection reported for SkRWPS. The protection exerted by
264 SkRWPS was also higher than the antioxidant activity of other spices such rosemary,
265 sage or ground mustard (Lindberg Madsen & Bertelsen, 1995).

266 The strong inhibition produced by the seasonings can be ascribed to their high
267 content of phenolic compounds (García-Lomillo et al., 2014). According to Pazos et al.
268 (2005) grape polyphenols present an optimal degree of polymerization and galloylation
269 to delay lipid oxidation in muscle systems. Some authors pointed out the importance of
270 simple extractable compounds such as flavanol monomers, phenolic acids or
271 anthocyanins (Yu & Ahmedna, 2013). In contrast, Ursini et al. (2001) reported the
272 relevance of polymerized compounds in comparison to their correspondent monomers.
273 Most probably, synergistic reactions between the high content in extractable

274 polyphenols and polymerized proanthocyanidins of the seasoning could explain the
275 observed antioxidant activity (García-Lomillo et al., 2014). The antioxidant
276 mechanisms of polyphenols are usually ascribed to their capacity to donate electrons to
277 free radicals formed during lipid oxidation and to their capacity to stabilize their
278 structure by the resonance delocalization of an electron within their aromatic ring
279 (Brewer, 2011). Moreover, grape phenolics are able to reduce highly-oxidizing ferryl
280 species (Kanner, Frankel, Granit, German, & Kinsella, 1994) as well as to scavenge
281 superoxide anions (Chen, Zheng, Jia, & Ju, 1990) and hydroxyl radicals (Rafat Husain,
282 Cillard, & Cillard, 1987). Other proposed mechanisms include the chelation of metal
283 initiators and the inhibition of the enzymatic activity of lipoxygenase (Duque, Pinto, &
284 Macias, 2011).

285 **3.3.Lipid oxidation under frozen storage of raw and cooked patties**

286 Meat products stored under different conditions are affected by different
287 oxidative mechanisms and natural antioxidants may show different efficacy in products
288 stored under different storage conditions (Lindberg Madsen & Bertelsen, 1995). Then,
289 the potential efficacy of the SkRWPS against lipid oxidation was also tested during
290 frozen storage of raw and cooked patties.

291 TBARS content increased during storage in both the cooked and the raw control
292 samples (Figure 5). At the end of storage, TBARS levels were 0.18 ppm in the raw and
293 0.08 ppm in the cooked control patties. The observed values were relatively low (ten
294 times lower than in patties stored under refrigeration) indicating only slight lipid
295 oxidation. This fact agrees with the stability of beef, in comparison to other meats,
296 during frozen storage (Igene et al., 1979). The slight formation of TBARS correlated
297 well with the low levels of hexanal that were detected (Figure 6).

298 The cooked patties showed higher stability against lipid oxidation than the
299 control patties (Figures 5 and 6). Although cooking usually produces pro-oxidant effects
300 due to the disruption of cellular organization and the release of iron with catalyst
301 activity (Kanner, 1994). Utrera, Morcuende, Ganhão, & Estévez (2015) also showed
302 lower oxidative stages in cooked samples, ascribing this fact to potential reactions
303 between formed aldehydes and meat proteins. Other authors also hypothesized that
304 Maillard reaction products formed during cooking may have antioxidant effect (Alfawaz
305 et al., 1994). Moreover, the inactivation of pro-oxidant enzymes such as lipases could
306 also contribute to the higher stability of cooked rather than raw samples (Kanner, 1994).

307 SkRWPS was able to reduce lipid oxidation in both the raw and the cooked
308 frozen patties significantly, which showed lower TBARS and hexanal values than the
309 control samples through the storage (Figures 5 and 6). No TBARS were formed in the
310 cooked patties and the raw patties showed a 60% reduction in comparison to the control
311 patties. The seasoning also inhibited hexanal formation by around 80% in both types of
312 patties. The formation of other VOCs such as alcohols and ketones was also inhibited
313 by the seasoning (data not shown) in a similar way to hexanal.

314 Similar products derived from wine pomace or herbs and spices also showed
315 capacity to inhibit lipid oxidation during frozen storage. For instance, Sánchez-Alonso
316 et al. (2006) found that red grape antioxidant dietary fibre was able to minimize the
317 development of primary and secondary oxidation in minced fish muscle stored at -20°
318 C. These authors highlighted the importance of the so-called “non-extractable
319 polyphenols” in the observed antioxidant capacity. Cagdas & Kumcuoglu (2015) also
320 found that grape seed powder added into the batter inhibited oxidation in precooked
321 chicken nuggets. Wine pomace extracts lowered TBARS values in vacuum packaged
322 chicken (Selani et al., 2011). El-Alim et al. (1999) tested different spices at 1% and

323 found that the most effective spices were marjoram, caraway and rosemary oleoresin
324 preventing TBARS formation in frozen meat products (Lindberg Madsen & Bertelsen,
325 1995), but the inhibition was lower than the protection exerted by SkRWPS.

326 The results observed in patties with sulfites were controversial. Sulfites
327 promoted the formation of TBARS in both cooked and raw hamburgers (Figure 5),
328 which suggests a pro-oxidant activity. However, sulfites successfully delayed the
329 formation of other VOCs derived from lipid oxidation and assessed by GC-MS
330 including hexanal (Figure 6). In this case, their efficacy was even higher than in samples
331 with SkRWPS. The discrepancy between TBARS and VOCs analysis as well as the
332 different behavior of sulfite samples stored under different conditions may be ascribed
333 to diverse factors. In frozen samples, the amount of water decreases in the liquid phase
334 and the solutes (as sulfites) are cryoconcentrated in the liquid phase of water, and may
335 reach pro-oxidant levels (Leygonie et al., 2012). In this regard, sulfites may induce
336 direct intramolecular cleavage of polyunsaturated fatty acids and induce the formation
337 of low molecular weight compounds (Southerland, Akogyeram, Toghrol, Sloan, &
338 Scherrer, 1982). From among the predominant polyunsaturated fatty acids in beef
339 muscle, sulfites are more likely to attack linolenic acid, (that is MDA precursor) than
340 linoleic acid (whose degradation forms hexanal) (Daley, Abbott, Doyle, Nader, &
341 Larson, 2010; Southerland et al., 1982). Then, cryoconcentrated sulfites in frozen
342 samples may directly attack double bonds from linolenic leading to MDA formation.
343 The chemistry of sulfites in food is rather complex and depends on several factors such
344 as concentration, temperature, matrix effect, etc. (Günther et al., 1997; Kaneda et al.,
345 1996). Further experiments should therefore be conducted for a complete evaluation of
346 the effect of sulfites on lipid oxidation in frozen samples.

347

348 4. CONCLUSIONS

349 Taken together, the results of the present study suggest that, from among the
350 three seasonings, the seasoning derived from seedless red wine pomace presented the
351 highest activity. It inhibited the formation of compounds derived from lipid oxidation in
352 patties under different storage conditions. This seasoning also showed higher efficacy
353 than sulfites that showed contradictory results in frozen samples, suggesting a possible
354 pro-oxidant activity.

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473

474 **FIGURE CAPTIONS**

475 **Figure 1.** Formation of TBARS in beef patties without antioxidants (Control), with the
476 addition red wine pomace seasonings (Seed RWPS, Whole RWPS or Skin RWPS at 2%
477 w/w) stored under high-oxygen atmosphere (70% O₂/30% CO₂) for 15 days at 4 °C.
478 Points show mean values of three replicates, and bars indicate standard deviations at
479 each sampling point.

480 **Figure 2.** Formation of TBARS in beef patties without addition of any antioxidant
481 (Control), with addition red skin wine pomace seasonings (Skin RWPS at 2% w/w) or
482 sulfites (300 ppm) stored under high-oxygen atmosphere (70% O₂/30% CO₂) for 15
483 days at 4 °C in two independent experiments (A and B). Points show mean values of
484 three replicates, and bars indicated standard deviation at each sampling point.

485 **Figure 3.** Formation of hexanal in beef patties without addition of antioxidants
486 (Control), with the addition of skin red wine pomace seasoning (SkRWPS at 2% w/w)
487 or with addition of sulfites (300 ppm) stored under high-oxygen atmosphere (70%
488 O₂/30% CO₂) for 15 days at 4 °C in two independent experiments (A and B). Results
489 are expressed in Arbitrary Units (AU). Points show mean values of three replicates, and
490 bars indicated standard deviations at each sampling point.

491 **Figure 4.** Formation of five volatile organic compounds: 1-pentanol, 1-hexanol, 1-
492 octen-3-ol, 2,3-octanedione and 2-pentylfuran in beef patties without addition of
493 antioxidants (Control), with the addition of skin red wine pomace seasoning (SkRWPS
494 at 2% w/w) or with the addition of sulfites (300 ppm) stored under high-oxygen
495 atmosphere (70% O₂/30% CO₂) for 15 days at 4 °C. Results are expressed in Arbitrary
496 Units (AU). Points show mean values of three replicates, and bars indicate standard
497 deviations at each sampling point.

498 **Figure 5.** Formation of TBARS in raw and cooked beef patties without addition of
499 antioxidants (Control), with the addition of sulfites (300 ppm SO₂) or skin red wine
500 pomace seasoning (SkRWPS, 2% w/w) stored in vacuum packaging for 6 months at -18
501 °C. Points show mean values of three replicates, and bars indicate standard deviations at
502 each sampling point.

503 **Figure 6.** Formation of hexanal in raw and cooked beef patties without addition of
504 antioxidants (Control), with the addition of sulfites (300 ppm SO₂) or skin red wine
505 pomace seasoning (SkRWPS, 2% w/w) stored in vacuum packaging for 6 months at -18
506 °C. Results are expressed in Arbitrary Units (AU). Points show mean values of three
507 replicates, and bars indicate standard deviations at each sampling point.

Figure 1

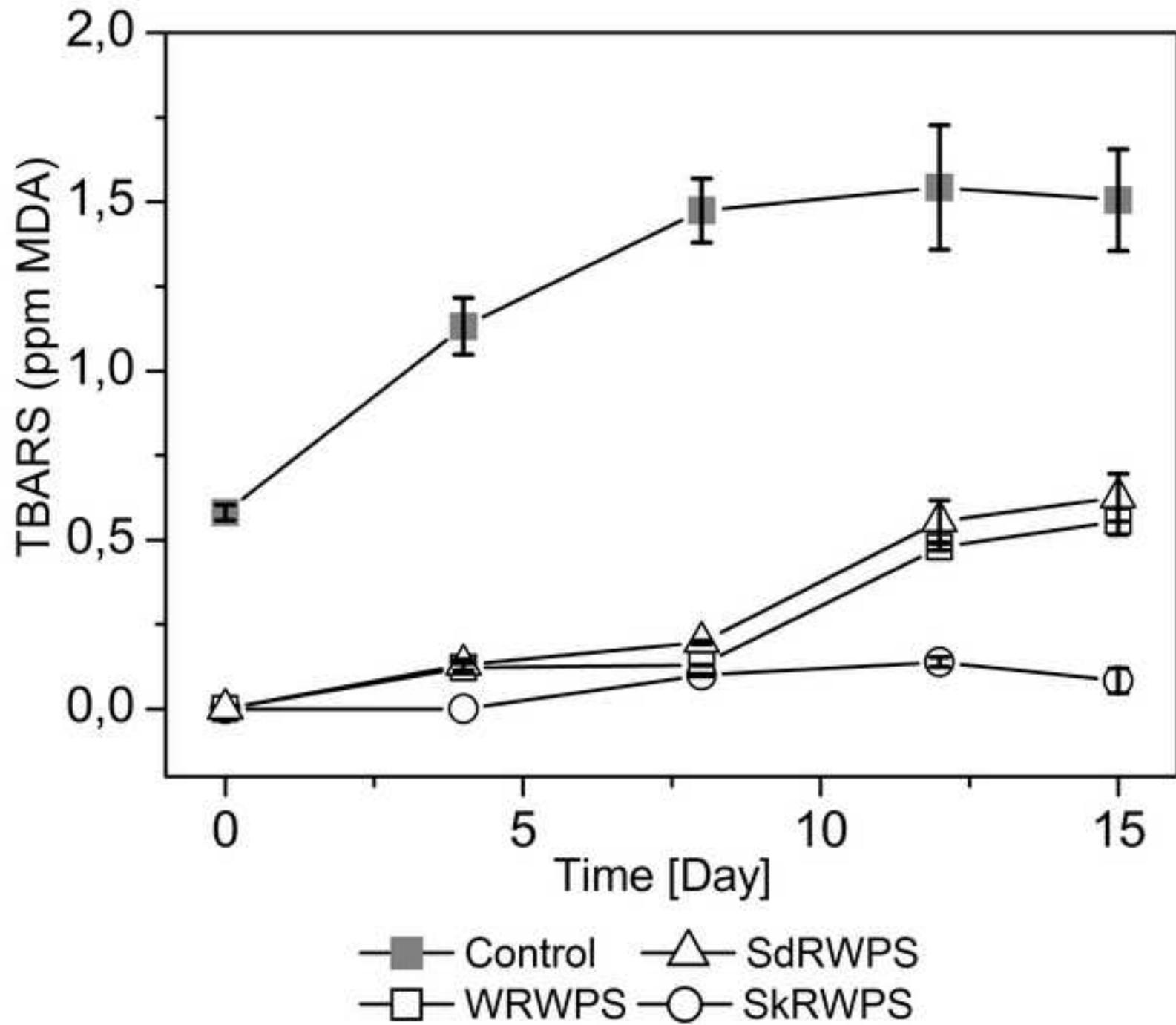


Figure 2

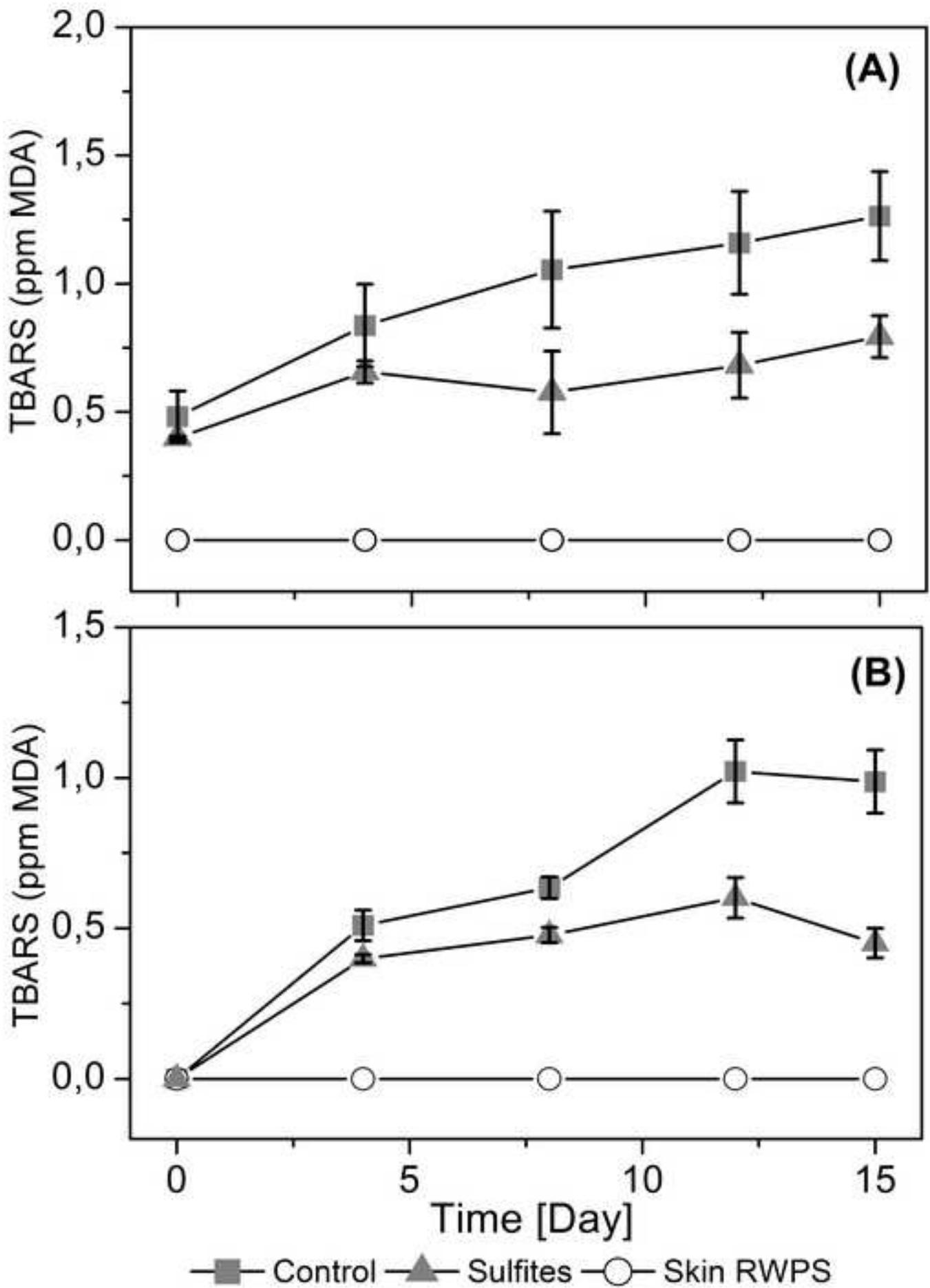


Figure 3

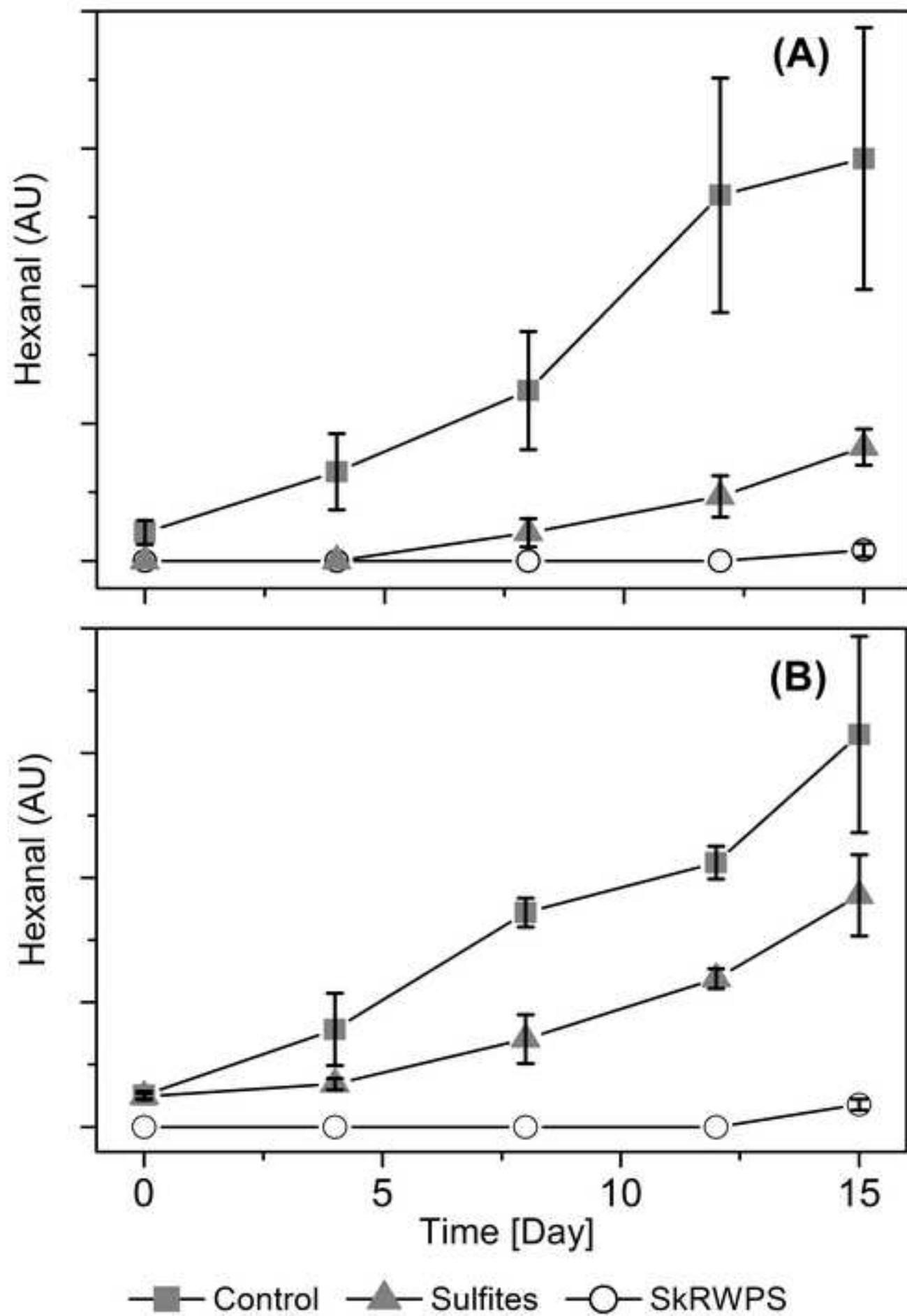


Figure 4

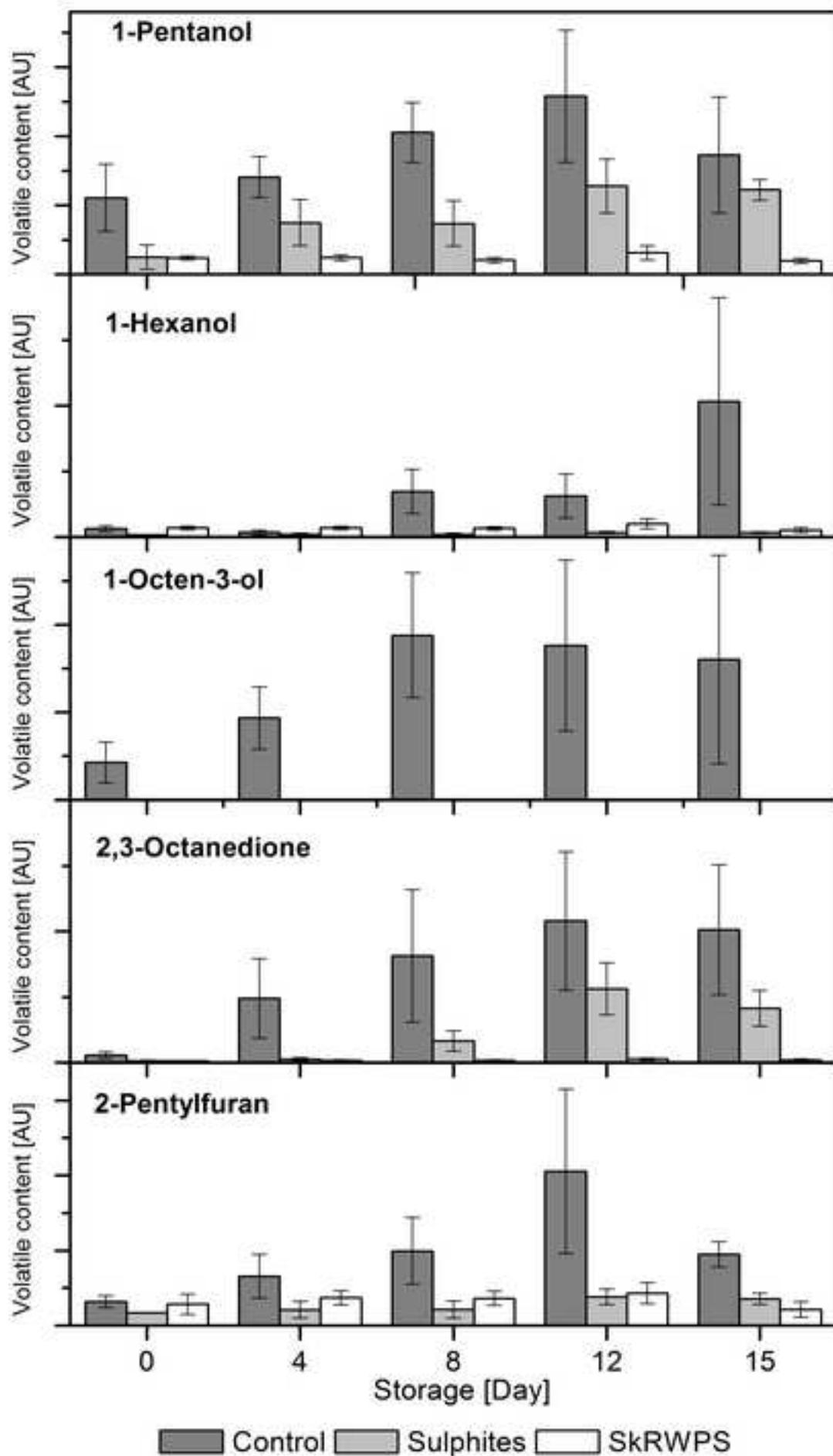


Figure 5

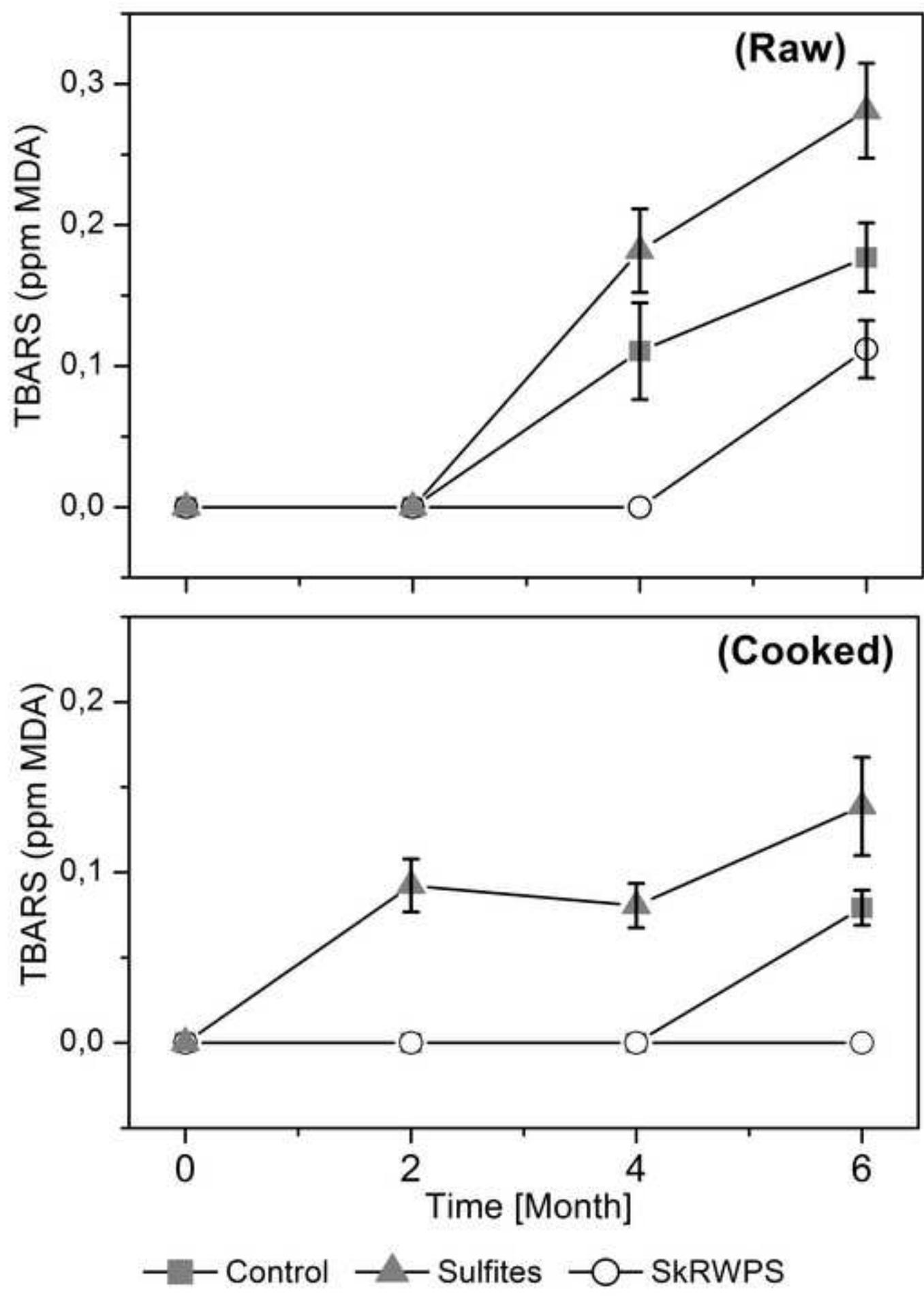


Figure 6

