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Desulfurization of thiosemicarbazones: the role of metal ions and biological implications

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Abstract

Thiosemicarbazones are biologically active substances whose structural formula is formed by an azomethine, an hydrazine and a thioamide fragments, to generate a $R_2C=N-NR-C(=S)-NR_2$ backbone. These compounds often act as ligands to generate highly stable metal-organic complexes. In certain experimental conditions, however, thiosemicarbazones undergo reactions leading to the cleavage of the chain. Sometimes, the breakage involves desulfurization processes. The present work summarizes the different chemical factors that influence the desulfurization reactions of thiosemicarbazones, as pH, the presence of oxidant reactants or the establishment of redox processes as those electrochemically induced, the effects of the solvent, the temperature and the electromagnetic radiation. Many of these reactions require coordination of thiosemicarbazones to metal ions, even those present in the intracellular environment. The nature of the products generated in these reactions, their detection *in vivo* and *in vitro*, together with the relevance for the biological activity of these compounds, mainly as antineoplastic agents, is discussed.

Keywords

 $Antitumor \cdot Biological \ activity \cdot Cyclization \cdot Desulfurization \cdot Metal \ complexes \cdot Thiosemicarbazone$

1. Introduction

Thiosemicarbazones (TSCs) constitute a broad family of compounds that have been under consideration since middle of last century because of their rich coordination chemistry with a wide range of transition and non-transition metal ions and their wide biological and pharmacologic activities, as exemplified by pioneering Bayley work in the 60s, who reported different copper complexes derived from thiosemicarbazone that showed antifungal activities [1,2].

Thiosemicarbazone ligands can be obtained under mild conditions by reaction between a suitable carbonyl compound (aldehyde or ketone) and a thiosemicarbazide [3]. In this process an imine bond is formed as part of the thiosemicarbazone $\mathbb{R}^1\mathbb{R}^2\mathbb{C}=\mathbb{N}-\mathbb{N}\mathbb{R}^3-\mathbb{C}(=S)-\mathbb{N}\mathbb{R}^4\mathbb{R}^5$ skeleton, with the release of a water molecule. The structure of a basic thiosemicarbazone skeleton is depicted in Fig. 1, where \mathbb{R}^1 and \mathbb{R}^2 may be nucleophilic groups and atoms, while \mathbb{R}^4 and \mathbb{R}^5 are the terminal $\mathbb{N}_{\text{thioamide}}^{\text{thioamide}}$ substituents. The wide structural diversity found in thiosemicarbazone chemistry is the result of modifying the type of carbonyl compound (aldehydes or ketones), substituents attached to the carbonyl moiety, the metal ion and its oxidation state, geometries, counterion, presence of solvent, added molecules, substituents on the S or $\mathbb{N}_{\text{thioamide}}^{\text{thioamide}}$ -atoms, or synthetic methodology [4,5,6]. They are highly delocalized systems, particularly when attached to the azomethine carbon.

$$\begin{array}{c} R^{1} \\ R^{2} \\ R^{2} \\ R^{3} \\ R^{4} \end{array} \xrightarrow{\mathsf{S}}_{\mathsf{R}^{4}} R^{5} \\ R^{5$$

Fig. 1 A general thiosemicarbazone skeleton

In solution, TSCs where $\mathbb{R}^3 = \mathbb{H}$ may exist as an equilibrium mixture of thione and thiol tautomeric forms [7] depending on the pH of the medium, with the thione form being the most stable in the solid state [8,9,10]. Upon coordination, the thione tautomer ligand usually acts in a neutral and bidentate mode [11], whereas the removal of the hydrazide NH proton gives rise to monoanionic thiolate ligands. These two coordination modes (Fig. 2) are strongly dependant on the reaction conditions, specifically the pH, or on the experimental methodology employed (chemical or electrochemical). Conformational and configurational changes occur, as those related with the N_{hydrazine}-C_{thioamide} bond being the *cis* configuration/*syn* conformation the usually formed after metal ion coordination [12]. In the presence of an additional coordinating group introduced *via* carbonyl compound, the TSC ligand increases its denticity. The alkylation of the thiocarbonyl sulfur in the derivatives causes complexation from the terminal amino group also leading to acidic character [13]. The properties of the thiosemicarbazones can be altered with the modifications in their chelating power and the binding patterns to the metal atom [14]. Under certain experimental conditions, carbonyl thiosemicarbazones also undergo cyclization. In these cases, the ligands can coordinate through the sulfur, the azomethine nitrogen and other heteroatoms present in the structure [15].



Fig. 2 Usual bidentate coordination modes of a thiosemicarbazone ligand

The versatility exhibited by TSC ligands is illustrated by the introduction of small modifications in the TSCs skeleton that can improve their therapeutic properties [16-18], highlighting that these properties can also be enhanced by the formation of complexes with different metal ions [18-20]. It seems that the coordination of metal ions modifies the lipophilicity that regulates the cell entry. It has been also found that some of the thiosemicarbazone complexes are more active than the uncoordinated precursor ligands. The biological activity of thiosemicarbazone complexes can be modulated by tuning the aromatic moiety as exemplified by the α (N)-heterocyclic tridentate thiosemicarbazone series. In the case of these ligands the hydrophobic moiety is more exposed to the solvent making it feasible to cross the cell membrane and to interact with essential enzymes in cells [21].

Taking into account all the features mentioned above, the coordination chemistry of thiosemicarbazone compounds and its biological applications have been of great interest and for that reason both aspects have been deeply explored in parallel during the last decades, giving rise to a long list of reviews on TSCs derived metal complexes and their biological properties. These reviews emphasised the relation between structural features in TSCs and the biological activities of their complexes. Some of the most relevant are commented below. Regarding to this, it is important to mention that in the present review, the abbreviations of the most biologically relevant TSCs are maintained as they appear in the literature.

The first review based on the coordination chemistry of thiosemicarbazones was published in 1975 by Campbell [22], with the focus on the study of the structure and bonding in different TSC transition metal complexes. In this report, the author highlights the influence of the sulfur atom nature on the behaviour of TSCs, which in all cases is coordinated to the metal ion. Since then, excellent reviews on TSCs have been published, completing Campbell's thiosemicarbazone study [23] or focusing on copper compounds [11], metal ions of groups 12, 13, 14 and 15 [24], palladium and platinum complexes [25], organotin(IV) [26] compounds. These reviews explore the potential as antitumour compounds [27] and, in particular, the ability of some thiosemicarbazone iron complexes to mediate in the generation of damaging reactive oxygen species (ROS) [28], deeping into their application as medicinal radiopharmaceuticals [19,29] and theranostic agents [30], or expanding the insight into antiviral, antimalarial and antifungal activities [31,32], highlighting those related to medicinal applications [33-42].

The first study on possible applications of TSCs in therapy is attributed to Domagk et al. [43]. In 1946, they suggested that these substances could be useful against tubercle bacilli given their chemical analogies with the antitubercular sulfamide and sulfone drugs. Since that, it has been explored their use against neurological pathologies [44,45], microbial diseases like small-pox [46], influenza, malaria, leishmaniosis, Chagas, leprosy and tuberculosis diseases [47-51], as antifungal compounds [31,52] and as potential theranostic agents [19,30,29,53,54]. The leap into the field of antitumour substances was made by Brockman et al. [55], who found antileukemic activity in pyridine-2-carbaldehyde thiosemicarbazone (HPTSC) and the corresponding thiocarbohydrazone. Many other studies have been performed in the field of cancer [20,31,26,56-80]. The seminal work performed during the 1960s established the basis for the TSCs with proved carcinostatic activity [81-83]. As a summary, TSCs have to be α -(N)-heterocyclic substituted, mainly with pyridine moieties, to give NNS tridentate ligand systems, which excludes semicarbazones that usually show a lower activity. In fact, these characteristics are present in all the TSCs screened in Phase I-III clinical trials until now (Fig. 3) [84-94]. Note that the common fragment to all of them is the structure of the pyridine-2-carbaldehyde thiosemicarbazone.



Fig. 3 TSCs tested in clinical trials: 5-hydroxypyridine-2-carbaldehyde thiosemicarbazone (5-HP, I), 3aminopyridine-2-carbaldehyde thiosemicarbazone (3-AP or Triapine[®], II), di-2-pyridylketone 4cyclohexyl-4-methyl-3-thiosemicarbazone (DpC, III), and (E)-N'-(6,7-dihydroquinolin)-8(5H)-ylidene 4-(pyridin-2-yl)piperazine-1-carbothiohydrazide (COTI-2, IV). In the middle of the scheme, the pyridine-2carbaldehyde thiosemicarbazone (HPTSC), as a common fragment to all of them

Different biological targets have been identified, like RNA [96], DNA [97-99], and several enzymes as thioredoxin reductase [100,101], xanthine oxidoreductase [102], RNA-dependent DNA polymerases [103], topoisomerase IIa [104,106] or succinate and NADH dehydrogenases [107]. Perhaps one of the most relevant findings about targets for the biological activity of TSCs was carried out by Sartorelli and Moore, who discovered that TSCs were able to inhibit the activity of the ribonucleotide reductases (RDRs), enzymes that catalyse the reduction of ribonucleoside diphosphates to deoxyribonucleoside diphosphates [108,109,110]. It is demonstrated that TSC-Fe complexes [111,112] generate ROS in the presence of O_2 that destroy the tyrosyl radical present in the M2 subunit of the enzyme, but binding to the protein surface and further chelation of Fe(III) ions from the active site has also been proposed [113]. The metal complexes,

or the presence of redox-active metal ions in the physiological medium, triggers the interaction with cell thiols and further reoxidation processes leading to the formation of ROS that destroy different cell structures [114-116].

The coordination to essential metal ions existing inside the cell seems to be necessary for TSCs to develop biological activity, to the point that the actual active forms are the TSC-Fe [117], and TSC-Cu complexes [118-122]. The inductive effects of the substituents in the TSC chain also have a notable influence on the redox properties and the biological activity of the TSC-metal complexes [123]. On the other hand, an increment of the lipophilicity of the metal complexes by modulating the substituents in the TSC backbone have been proved to increase the intracellular redox activity and to improve the antiproliferative efficacy [124-126], if the value of the Partition coefficient (P_{calcd}) is kept inside an optimal range log P_{calcd} 3.1–4.5 [127].

It is well established that transition metals induce activation and transformations in the carbon-sulfur bonds [128-133]. In the case of TSCs, the confluence of metal ions coordination and different physicochemical factors can lead to changes in the ligand skeleton involving the loss of the sulfur atom. Generally, a desulfurization process can be described as an oxidation reaction in which the sulfur is lost from the thioamide group. Nowadays, the literature shows a variety of examples of desulfurization reactions in thiosemicarbazone ligands but, as far as we are aware, there are still no reviews available. In this review we will give an insight of different desulfurization processes in thiosemicarbazone complexes that have been reported to date. It also explores the conditions that make possible desulfurization processes in TSC-metal complexes, the products arisen from those and the biological relevance of these reactions. We have performed a tentative classification of desulfurization process in TSCs depending on the factors involved: pH induced desulfurizations, desulfurization in electrochemical processes and desulfurization by oxidant reactants and others (radiation-, thermal-, solvent- and coordination-induced desulfurizations). Also, a section of biological implications of these processes is reported.

2. A survey for desulfurization reactions in thiosemicarbazone metal complexes

2.1. pH induced desulfurizations

Extreme pH values have been proved to provoke ruptures in the thiosemicarbazone chain of metal complexes, leading to different products coming from the ligand desulfurization. Notwithstanding, some effects of the medium are detected even at neutral pH values. The present section summarizes some key reactions dealing with desulfurization induced by acid or basic media.

a) Conversion of thiosemicarbazones into carbonitriles in basic medium

Aqueous solutions of the CuL(NO₃) compound (HL = pyridine-2-carbaldehyde thiosemicarbazone, HPTSC) basified by NaOH addition to pH = 9–11 undergo partial desulfurization to yield brown precipitates containing, as a major component of the mixture of phases, the [CuL(L^{CN})] compound [134], where HL^{CN} = (pyridine-2-ylmethylene)hydrazinecarbonitrile, following the general reaction depicted in Fig. 4. The crystal structure of [CuL(L^{CN})] reveals its square pyramidal geometry. The carbonitrile ligand acts as bidentate and the bond distances in the NCN terminal moiety fall in intermediate values between cyanamide (N–C=N) and carbodiimide (N=C=N) character. The presence of the carbonitrile can be easily checked by a characteristic infrared band at 2108 cm⁻¹ corresponding to v(CN).



Fig. 4 Transformation of pyridine–2–carbaldehyde thiosemicarbazonatocopper(II) (PTSC-Cu) into the (pyridine-2-ylmethylene)hydrazinecarbonitrilecopper(II) complex

An extensive study has been carried out to elucidate the conditions and mechanism of this reaction, whose transcendence will be discussed in other sections. The main results are given below.

1. Trials at different pH values (7.4, 9.0, 11.0 and 13.0), temperatures (40, 50 and 80 °C) and times (1 and 5 h) allowed to establish the best conditions to achieve mixtures with the highest $[CuL(L^{CN})]$ content at pH 9.0, 80 °C and 1 h. Excess of base (pH 11–13), heating and time led to mixtures rich in inorganic matrix where sulfide and sulfate anions were qualitatively detected.

2. No desulfurization reactions are observed for the free HPTSC ligand in the same experimental conditions.

3. The reaction is not sensible to the co-ligand, and suspensions of the CuLCl compound, less soluble than CuL(NO₃), behave in a similar way. CuLCl and CuL(NO₃) are, actually, the dinuclear [{CuLCl}₂] and the 2D [CuL(NO₃)]_n compounds, respectively [135,136].

4. The same results are obtained with different bases, as KOH, triethylamine (NEt₃) and a 0.5 M $Na_2CO_3/NaHCO_3$ buffer.

5. The process strongly depends on the addition order, because only affects to preformed $[CuL]^+$ entities. Note that the addition of Cu(II) solution to an aqueous HL solution at pH 13.0 affords the attainment of the $[CuL_2]$ complex, retaining the TSC integrity.

6. No evidences of TSC breakage have been observed for Pb(II), Fe(II,III), Co(III), Ni(II) and Zn(II) ions in experiments carried out at pH 11.0 for 1 h (Fig. 5) [137]. Attempts of desulfurization of the HPTSC ligand using HgCl₂ and Pb(AcO)₂ were also unsuccessful [138].



Fig. 5 Summary of the behavior of different metal ions coordinated to PTSC at basic pH values

In order to unveil the features of the reaction, solids precipitated from reactions between equimolar amounts of HL and Cu(NO₃)₂ at pH values of 2.0 (no addition of base), 4.0, 7.4 and 9.0 were analysed [139]. IR spectra recorded on the dark brown solids revealed the presence of medium to intense bands in the 2116–2106 cm⁻¹ region in the samples obtained at pH 7.4 and 9.0, which suggests ruptures in the TSC chain. In the mass spectra of these compounds were also detected peaks at m/z 628.99 and 630.99 attributed to $[Cu_2L_2(L^{CN})]^+$ and $[Cu_2(HL)_2(L^{CN})]^+$ ions, whose intensity increased in those experiments performed at 80 °C. These species could be related to the partial desulfurization of dinuclear $[Cu_2L_3]^+$ cations. These dinuclear entities have been isolated and crystallized, from the reaction of $Cu(ClO_4)_2$ with thio- and selenocarbazones in soft basic media by addition of NaOH (pH = 7.6, HL = HPTSC [139]) NaAcO (HL = 2-acetylpyridine 4-N-phenylthiosemicarbazone [140]) and NEt₃ (HL = 2-acetylpyridine 4,4-dimethyl-3-selenosemicarbazone) [141]. Their stabilization as solids containing $[Cu_2L_3]^+$ cations seems to require the presence of low coordinating co-ligands, as perchlorate or nitrate [139].

Mass spectrometry studies carried out on aqueous solutions at pH 4.0, 7.4, 9.0, 11.0 and 13.0 showed the presence of **peaks attributed** to $[Cu_2L_3]^+$ and $[Cu_2(HL)_2L]^+$ species only at pH 7.4. The detection of peaks attributed to $[Cu_2L_3]^+$ species at physiological pH values have been reported for the Cu(II) derivative of the antitumour drug Triapine[®], the 3-aminopyridine-2-carbaldehyde thiosemicarbazonecopper(II) complex [142,143].

Taking these results into account, it can be proposed that the desulfurization may proceed even at physiological pH values, at least in a certain extent. Considering the key role that $[Cu_2L_3]^+$ entities could play, a mechanism has been proposed and its validity checked by quantum mechanical calculations (see Fig. 6) [139]. The theoretical study identifies a highly exergonic ($\Delta G = -146.1 \text{ kcal} \cdot \text{mol}^{-1}$) nucleophilic attack of a hydroxide anion to one of the thioamide carbon in these $[Cu_2L_3]^+$ species, which could trigger the process. It must be emphasized that all the stages of the mechanistic sequence proposed are built with monomer or dinuclear metal complexes whose crystal structures have been previously reported as $[CuL(OH_2)]^+$ entities in $[{CuL(OH_2)}_2](SiF_6)\cdot 4H_2O$ [144], $[CuL_2]$ [134], $[CuL(L^{CN})]$ [134], and $[{CuL(SH)}_2]$ [137]. The latter could arise from dehydration of the $[{CuL(OH_2)(SH)}_2]$ (VIII) dimers acting the crystal packing as driving force through the strong non-covalent interactions (hydrogen bonding and π - π stacking) present in the lattice.

The complexity of this reaction could hide other important influences, as the concentration of $[CuL]^+$ cations, which could be dramatic for the formation of $[Cu_2L_3]^+$ species, or the coexistence of simultaneous redox processes involving the Cu(II) ions. The latter could involve the participation of the TSC ligands, in the same way that the self-reduction processes described for the $[FeL_2]^+$ complexes at pH > 6 to give $[FeL_2]$ [145-147].



Fig. 6 Proposal for a mechanism of the desulfurization of HPTSC to carbonitriles in basic media. Values of the Gibbs free energy arisen from DFT calculations are indicated for each step. The dynamic of transformations is represented by arrows

Apart from mechanistic proposals, the possibility of visualizing the terminal NCN fragment as cyanamide (N-C=N) or carbodiimide (N=C=N) connects this process with broader desulfurization reactions dealing with thioamide groups, as the reduction of thioamide to imine by triethylsilane at 80 °C with Fe catalyst [148], the conversion of thioamide into nitrile in the presence of a Rh catalyst in benzene basified by NEt₃ [149], or even the use of base to synthesize $1\frac{H}{-1}$,2,4-triazol-3-amines from condensation of amidines, isothiocyanates and hydrazines [150].

b) Appearance of sulfate from thiosemicarbazone breakage in acid and basic media

In other processes, often less understood, sulfate ions were identified, the origin of which only can be the release of the thioamide sulfur atom.

Due to the strong similarities with the system shown in Section 2.1.a), we start describing the behaviour of the [{Cu(L')(NO₃)}₂] compound when is exposed to basic media (HL' = pyridine-2-carbaldehyde 4-N-methylthiosemicarbazone, HPTSC4m) [137,151]. Experiments performed on this compound at pH 9.0 and 80 °C for 1 h yielded a dark brown compound with no evidence of TSC breakage. However, an increase in pH to 11.0, even at lower temperatures (50 °C) provoked the release of irritant gases, whose analysis by gas chromatography-mass spectrometry revealed the presence of methylisothiocyanate (S=C=N-CH₃), pyridine 2-carbaldehyde and N-methylthiosemicarbazide (H₂N-NH-C(=S)-NH-CH₃). It must be pointed out that these chemicals are starting materials for the synthesis of HPTSC4m. Once filtered the suspension, slow evaporation of the mother liquors resulted in the attainment of single crystals of the [{Cu(L')(OH₂)} {Cu(L')(OSO₃)}]·5H₂O derivative, where sulfate anions arise from the loss and oxidation of the sulfur atom of the thiosemicarbazone ligand. Notwithstanding, when the addition of base to pH 11.0 was carried out in a cold water bath (T < 15 °C) and the solution was kept at 5 °C inside a freezer for 2 months, crystals of [{Cu(L')₂NO](OH)·5H₂O were obtained, which contain unmodified HPTSC4m ligand. No desulfurization process is observed for the free HL' ligand in the experimental conditions reported above.

The heterogeneous content of the identified products suggests that a very complex set of processes take place simultaneously in this reaction, which precludes a clear and univocal mechanistic proposal. These facts, put all together, demonstrate that coordination to Cu(II) ions is required for the transformation of TSCs into carbonitrile triggered by basic medium.

c) Other desulfurization processes of thiosemicarbazone metal complexes in basic medium

A very interesting combination of desulfurization and cyclization processes in TSCs was reported by Castiñeiras and García-Santos in reactions of pyridine-2-formamide thiosemicarbazone (HFTSC) with $Mn(ClO_4)_2 \cdot 6H_2O$ [152]. They attained the $[Mn(HFTSC)_2](ClO_4)_2$ compound, which contained the unaltered HFTSC ligand, when the reaction proceeded in ethanol under reflux for 2 h. However, if drops of NEt₃ were added in the presence of 2,2'-bipyridine (bpy), a yellowish-brown precipitate was obtained. After removing it, crystals of $[Mn(bpy)_2(NCS)_2]$ could be isolated from the mother liquors by slow evaporation, where thiocyanato ligands came from the breakage of HFTSC.

In addition, when aqueous solutions of $Mn(ClO_4)_2 \cdot 6H_2O$ -bpy-HFTSC (in a 1:2:1 molar ratio) were basified with NaOH and refluxing for 2 h, a brown precipitate identified as $[Mn(bpy)(pdo)_{0.5}(pta)](SO_4)$ was recovered, where pta = 3-(pyridine-2-yl)1H-1,2,4-triazol-5-amine, and pdo = 2,4-pentanediol. Further recrystallization of this solid in pyridine (py) yielded crystals of $[Mn(py)_4Mn(py)_2(OH_2)_2(\mu-SO_4)_2]\cdot 4H_2O$, whereas the recrystallization from DMSO/CHCl₃ or DMF/diethyl ether mixtures afforded yellow crystals of $[Mn(pdo)(pta)_2]_4(SO_4)_2 \cdot 4H_2O \cdot S_8$. Note that pta, pdo, S_8 , and sulfate anions only could arise from desulfurization and decomposition of the TSC ligand, that authors attributed, at least in part, to the role of the Mn(II) ion as redox catalyst. The formation of pta is depicted in Fig. 7.



Fig. 7 Transformation of HFTSC into pta induced by coordination to Mn(II) in aqueous basic medium

d) Conversion of thiosemicarbazones into thiocyanate in acid medium

Slow addition of 1-8 drops of $HClO_4(c)$ over mixtures of HPTSC and $Cu(ClO_4)_2$ in ethanol yielded, at a brown compound identified as the 1D first stage, а complex $\{[Cu(PTSC)(OH_2)][Cu(PTSC)(OClO_3)]\}_n$ nClO₄·2nH₂O. Once filtered off and days after, bright green prismatic crystals of [Cu(HPTSC)(NCS)](ClO₄) could be gathered from the mother liquors, whose crystal building exhibited an incommensurate modulated structure [139]. Analogous results were serendipitously obtained by reaction of $Cu(NO_3)_2$ and HPTSC in a water: methanol mixture (1:1) in the presence of Na_2ATP (adenosine-5'-triphosphate disodium salt), which spontaneously evolved to pH 0.6 by slow evaporation for two months, to yield the $[Cu(HPTSC)(NCS)][Cu(HPTSC)(NCS)_{0.72}(NO_3)_{0.28}](NO_3)_2$ compound [137]. The stabilization of the complex in so extremely acid medium ratifies the high affinity between Cu(II) ions and HPTSC. The bands at 2136 cm⁻¹ and 2097 cm⁻¹ in the infrared spectra of the perchlorate and nitrate derivatives, respectively, allowed to easily identify the presence of the pseudohalide. No traces of thiocyanate were detected in solids arisen from acid treatments of the free HPTSC ligand.

A proposal for the desulfurization process is provided in Fig. 8. Coordination of Cu(II) ions to the thioamide group in the neutral TSC would trigger a nucleophilic attack by a water molecule. The formation of intermediates as 2-(hydrazinomethyl)pyridine (RNH₂) and carbamothioic O-acid, together with further dehydration and deprotonation of the latter, would generate the thiocyanate groups.



Fig. 8 Proposal for a mechanism of the desulfurization of HPTSC to thiocyanate in acid media

Formation of thiocyanate in chemical systems similar to TSCs, however, has been also described in basic media, as that reported for bis(N-alkyldithiocarbamato)cadmium(II) complexes in the presence of Et₃N to give alkyl isothiocyanate Fig. 9 [153].



Fig. 9 Formation of alkyl isothiocyanate from dithiocarbamatocadmium(II) complexes in basic medium

Desulfurization reactions in other sulfur-containing ligands have been reported, as the conversion from thiourea to urea-derivatives induced by basic-neutral media of NaAcO in ethanol [154].

e) Influence of pH in other reactions: from TSC to picolinate and formation of disulfide

Single crystals of the [Cu(PTSC)Cl]₂[Cu(pic)₂]·2H₂O compound were serendipitously obtained after slow evaporation of aqueous suspensions of CuCl₂, HPTSC and guanine at pH 5.6. The bis(picolinato)copper(II) species arose from the breakage of the TSC ligand [137], as will be discussed later for other reactions.

Chalcogenide elements, sulfur in particular, are of substantial relevance considering potential biomimetic applications. In biological environments, thione/thiol - disulfide exchange reactions have an important role on enzymatic processes involved in cellular functions, including redox activity, protein folding, DNA expression/repair or apoptosis. Disulfide formation induced by a redox process implies a weakening of the C=S thioamide bond in TSCs and this process could underlie desulfurization reactions as an intermediate

step. For that reason, a better compression of the factors involved in the S-S bond cleavage and formation is required. In this sense Hong and co-authors studied factors affecting to the interconversion between Cu(II) and Zn(II) thione-disulfide dinuclear thiosemicarbazone complexes, thus demonstrating feasible interconversion with acid-base or solvent changes (Fig. 10), whereas interconversion did not take place in the absence of metal ions [155].



Fig. 10 Disulfide formation in TSC-Cu(II) complexes

2.2. Desulfurization in electrochemical processes

Electrochemical synthesis has emerged as an interesting synthetic methodology leading to compounds usually different from those obtained from starting metal salts. Despite of the electrochemical conditions has not been identified as crucial for desulfurization reactions to occur, the great number of examples obtained from electrochemical mother liquors justified a section an in this review.

of During electrochemical reactions the thiosemicarbazone N-{2-([4-Nethylthiosemicarbazone]methyl)phenyl}-p-toluenesulfonamide, H_2L^1 , with manganese and copper metals, interesting catalysed processes were found to occur, with remarkable consequences regarding the ligand skeleton structure. In synthesising the manganese complex, it was obtained an unexpected dithiolate thiosemicarbazone tosyl ligand, H₂L², as a side-product. The disulfide ligand H₂L² presented here was formed by an oxidation process of the initial thiosemicarbazone ligand H_2L^1 during experiments on the electrochemical synthesis of the manganese complex. The proposed mechanism could start with a thionethiol equilibrium in solution, followed by coordination of manganese atoms to two different doublydeprotonated ligand units and coordination of the thiolate sulfur atom as a μ_2 -bridge between the two Mn(II) ions. This step was followed by a reductive elimination process resulting in the coupling of two thiolate units, thereby creating the disulfide bond (Fig. 11). The oxidation of the thiosemicarbazone to disulfide under physiological conditions could lead to a reinterpretation of the biological properties of some thiosemicarbazone systems, primarily those aspects related to their possible therapeutic uses [156].



Fig. 11 Mechanism proposed for the formation of the disulfide ligand H_2L^2

In the case of copper, the solid complex was $[CuL^1]_2$, but the crystallized product showed the copper atoms bound to a new cyclized thiosemicarbazone ligand, H_2L^3 , as was shown in the structures of the complexes $[Cu(L^3)]_2 \cdot CH_3 CN$ and $[Cu(L^3)(H_2O)]_2 \cdot CH_3 CN \cdot H_2O$. Oxidative cyclization of the original ligand H_2L^1 by Cu(II) ions followed by the addition of an acetamide fragment, accompanied by a reductive elimination process led to the formation of the new tetradentate ligand H_2L^3 . The new ligand features a five-membered 1,2,4-triazole ring, formed by nucleophilic attack of the thioamide nitrogen on the imine carbon (Fig. 12), followed by the addition of an acetamide fragment to a Cu(II) ion and the sulfur atom. The presence of an

acetamide residue in the reaction medium may possibly be explained in terms of copper-catalysed hydrolysis of acetonitrile whereas the attachment of an acetamide fragment to the formed 1,2,4-triazole-3-thione ring could take place by coordination of the metal followed by a reductive elimination process.



Fig. 12 Proposed mechanism for the formation of the complexes $[Cu(L^3)]_2 \cdot CH_3 CN$ and $[Cu(L^3)(H_2O)]_2 \cdot CH_3 CN \cdot H_2O$ by copper-catalysed oxidative cyclization of the ligand H_2L^1 followed by addition of an acetamide group

Another case of sulfate generated by thiosemicarbazone desulfurization was found in the complex $[Cu_2(LEt)_2(SO_4)]$, isolated by slow recrystallization of the mother liquors obtained after separation from the expected solid complex $Cu(L^{Et})_2$ (HL^{Et}= pyridine-2-carbaldehyde-4-*N*-ethyl-thiosemicarbazone). The complex consists of a neutral dinuclear Cu(II) entity, acting the sulfate group as bidentate ligand to achieve electroneutrality [157]. Some other Cu(II) dimer complexes derived from 2-pyridincarbaldehyde thiosemicarbazones and incorporating sulfate groups arising from desulfurization processes have been published before but, in these cases, the sulfate group acted as monodentate ligand. The mechanism explaining this desulfurization process starts with a nucleophilic attack, probably of a water molecule to the thioamide carbon atom (Fig. 13) and the subsequent release of copper sulfide to the media during the electrochemical synthesis. Slow oxidation of copper sulfide by oxygen from the water or the air could convert the copper(II) sulfide into copper(II) sulfate. Finally, the assembly of complex [Cu₂(LEt)₂(SO₄)] would take place by coordination of the ligands H₂L^{Et} and sulfate to the Cu(II) metal ions.





Fig. 13 Proposed mechanism for the desulfurization of pyridine-2-carboxaldehyde-4-*N*-ethyl-thiosemicarbazonecopper(II) complexes to form sulfate ions

Metal-free thiosemicarbazones have also been electrochemically studied, in particular isatin derivatives. Thus, upon oxidation, it was proposed the breakage the of the C=N bond, generating isatin and thiourea fragments for all evaluated molecules (Fig. 14). Regarding the reduction, the cleavage of the N-N bond as well as the generation of 3-aminoindolin-2-one and thiourea moieties was proposed [158].



Fig. 14 Electrochemically induced cleavage of isatin thiosemicarbazone

Electrochemistry, coupled to high resolution mass spectrometry (HRMS), was also employed to investigate the possible relationship between the structure of α -N-heterocyclic thiosemicarbazones and their metabolic behaviour. To this end, the metabolites of ten different Triapine[®] derivatives with a wide range of antitumour activities were analysed. In general, for all the investigated thiosemicarbazones, the identified processes of the metabolic reactions were hydroxylation, oxidative desulfurization (formation of the amidrazone and, for some derivatives, also the semicarbazone) and disulfide dimer formation and dehydrogenation in some cases (Fig. 15). In general, desulfurization was detected for all the investigated compounds, thus confirming that the study of desulfurization process is crucial for a better understanding of the thiosemicarbazones biological activity [159].



Fig. 15 Key metabolites of Triapine®: the dehydrogenated ring-closed thiadiazole and hydroxylated species

2.3. Desulfurizations by oxidant reactants

The interaction between thiosemicarbazone complexes and oxidants can give rise to desulfurization by reactions often involving oxidative cyclization processes. Some of them are described in the present section.

a) Conversion of thiosemicarbazones into 1,3,4-oxadiazoles by halate ions

The addition of bromate to aqueous solutions of HL (where HL = HPTSC and HPTSC4m) triggered a complex process whose main reaction led to 1,3,4-oxadiazole derivatives [160,161], that could be regarded as is shown in Fig. 16.



Fig. 16 Transformation of thiosemicarbazones into 1,3,4-oxadiazoles. Thiosemicarbazones: pyridine–2– carbaldehyde thiosemicarbazone (R = H, HL = HPTSC) and pyridine-2-carbaldehyde 4-Nmethylthiosemicarbazone ($R = CH_3$, HL = HPTSC4m). Oxadiazoles: 2-amino-5-pyridin-2-yl-1,3,4oxadiazole (R = H, $L_{oxad} = 134OXAD$) and 2-methylamino-5-pyridin-2-yl-1,3,4-oxadiazole ($R = CH_3$, $L_{oxad} = 134OXADm$)

The study of different well-characterized solids isolated from this process allowed to distinguish several steps, which are drawn in Fig. 17, despite some stages in the mechanism remain unclear. The reaction proceeded smoothly with the addition of bromate to aqueous solutions of preformed CuL(NO₃) complexes at pH ~ 6 (Step 3). An olive-green precipitate corresponding to the centrosymmetric S-bridged [{CuLBr}₂] dimer appeared (Step 3) and, after filtering it, the initial dark green colour of the solution gradually became lighter. Simultaneusly, the pH decreased to 3–4 and an irritant gas was released. Two days later, single crystals of [{Cu(L_{oxad})(OH₂)₂(OSO₃)}₂] were obtained (Step 4). The addition of K₄[Fe(CN)₆]·3H₂O led to the coprecipitation of purple K₂Cu[Fe(CN)₆]·H₂O and L_{oxad}, whose particle sizes were different enough to eliminate the complex in the filtrate while the white organic compound was retained in the filter and, afterwards, recrystallized in ethanol (5). The use of NaHCO₃ instead of K₄[Fe(CN)₆]·3H₂O also allowed to isolate the oxadiazole ligand. The rate of the reaction was increased with heating and addition of small

amounts of acid. However, brown suspensions were attained in the excess of acid or strong heating, whose filtration allowed to isolate crystals of bis(pyridine-2-carboxylate)copper(II) (6).

The reaction also took place without Cu(II) ions, but strong acid media was required (pH = 0–1) and, after filtration of a dark red unidentified solid, further treatment with NaOH to pH = 3–4. In the presence of acid media, mixtures of compounds were invariably obtained and purification was needed. The use of KIO₃ in acid media yielded the same results than KBrO₃, but the formation of iodine, probably due to comproportation between I⁻ and IO₃⁻, made more difficult the purification of the product. Furthermore, the addition of KIO₃ to the CuL(NO₃) complexes at pH ~ 6 led, firstly, to the attainment of a green compound identified as CuL(IO₃), which evolved when the suspension was kept with stirring to yield [{CuLI₂] dinuclear compounds in a final step, but oxadiazole derivatives were not attained. The analogous CuL(BrO₃) compound could not be isolated in these experiments. On the contrary, none of the attempts carried out by using chlorate as oxidant gave any evidence for oxidative cyclization, which could be due to kinetic factors. Finally, no oxidation compounds were attained by using pyridine–2–carbaldehyde 4,4'-N-dimethylthiosemicarbazone (HPTSC44m) neither free nor coordinated to Cu(II) ions, which suggests steric influences in the process.



Fig. 17 Reaction pathways identified for the different reactants and conditions

A plausible mechanism can be proposed taking into account the experimental evidences, which is depicted in Fig. 18 for the free ligand, but it could be extrapolate, at least in part, for the Cu(II) complexes.



Fig. 18 Mechanism suggested for the halate-induced oxidative cyclization of TSCs to 1,3,4-oxadiazoles

These processes allow to prepare 2-amino-1,3,4-oxadiazoles as the only product through easy and unexpensive reactions, sometimes in less than an hour.

Oxidations of acyl thiosemicarbazides, acyl carbodithioates and acyl thioureas to oxadiazoles, often promoted by metal ions, have been frecuently described in the literature [162-183].

b) Other desulfurization of thiosemicarbazones by oxidant reactants

The use of other oxidants to desulfurize and cyclize TSCs has been described for a long time, as it was reported by Landquist in the attainment of triazolinones by reaction with MnO_2 [184].

From a broader point of view apart from TSCs, the use of oxidants as H_2O_2 , 1O_2 and I_2 has been described for the desulfurization of other sulfur-containing molecules, as 8-thioguanosine to guanosine [185] or dithiocarbamato-Ru(II) complexes [186,187], among others.

Desulfurization of TSCs through processes involving redox-active metal ions is discussed in Section 2.4.d.

2.4. Others (radiation-, thermal-, solvent- and coordination-induced desulfurizations)

a) Solvent-induced desulfurizations

Another factors like light, pressure or solvent must be considered to have a relevant role in thiosemicarbazone desulfurization reactions. In this sense, recently it was presented the dinuclear helicate $[Ag_2(H_2L)_2]SO_4$ obtained by crystallization of the bisthiosemicarbazone cluster helicate $[Ag_4L_2]_2$ in the absence of any sulfate source, after a rare desulfurization process that takes place only in chloroform (Fig. 19). Three factors were investigated in this conversion: solvent, light and time. Only those recrystallizations performed in chloroform in the presence of light led to the formation of the sulfate helicate crystals. Regarding to time, the appearance of the sulfate crystals took place after a long crystallization period of 3-4 weeks. Some ¹H NMR studies mimicking the recrystalization process by protonation of the hydrazine NH groups, thus resulting on the dihelicate unit $[Ag_2(H_2L)_2]^{2+}$. It is relevant to mention that two side-products were identified along the desulfurization: silver hydrogen sulfide/sulfide, that easily evolved to the sulfate counterion by oxidation by moisture oxygen, and the organic fragment semicarbazone released

from the oxidative desulfurization. Finally, the remaining dinuclear complex combined with the sulfate thus generating the final crystallized sulfate dihelicate $[Ag_2(H_2L)_2]SO_4[188]$.



Fig. 19 Formation of the dinuclear helicate $[Ag_2(H_2L)_2](SO_4)$ by desulfurization of the doubletetranuclear cluster helicate $[Ag_4L_2]_2$

Notwithstanding, it must be admitted that not always coordination to metal ions is required to provoke solvent-induced desulfurization in TSCs. For instance, Peach and Dilworth et al. [189] described intramolecular cyclization reactions in recrystallization experiments of bisthiosemicarbazones that involved the loss of one of the thioamide sulfur atoms. The isolated product depended on the kind of solvent used, as is shown in Fig. 20.



Fig. 20 Benzil bis(4-phenyl-3-thiosemicarbazone) and related cyclized products of solvent-induced desulfurization processes

b) The influence of the electromagnetic radiation

Microwave radiation is known to promote desulfurization in organo-sulfur compounds being this process of great importance for reducing harmful emissions during the combustion process [190]. Microwave irradiation of thiosemicarbazones gave the corresponding isothiocyanates, which on addition of either activated nitriles or aldehydes furnished various types of azines (Fig. 21) [191]. This process was also reported for thiosemicarbazone analogues, as thiocarbohydrazones [192].



Fig. 21 Azine fragments formed after microwave irradiation of thiosemicarbazones

Visible light has been used as an agent for desulfurization of thiols and disulfides [193]. However, as far as we are aware, none of these processes has been reported for TSCs.

c) Thermal induced desulfurizations

Heating is another point to be considered in desulfurization processes experienced by TSC complexes. Thus, prolonged treatment in refluxing aqueous solutions of the complex $CuL(NO_3)$, being HL= pyridine-2-carbaldehyde thiosemicarbazone, induced breakage of a portion of the thiosemicarbazone molecules to give HS⁻ ligands that were identified in the structure of [{CuL(SH)}₂], whose crystals were collected from the mother liquors (Fig. 22). In contrast, the free ligands remain unaltered under the same experimental conditions. Extending the studies to other metal ions, as Fe(III), Co(III), Zn(II) and Pb(II), it was found that only bis(thiosemicarbazonato)iron(III) species underwent breakage whereas there was no evidence for desulfurization processes in the Co(III), Zn(II) and Pb(II) derivatives. The IR data suggested that breakage of HL ligands gave rise to thiocyanato ligands in the above mentioned Cu(II) derivatives, while a non-coordinated thiocyanate anion was present in the Fe(III) decomposition product [137].



Fig. 22 Summary of desulfurization processes by refluxing on $[ML]^+$ and $[ML_2]_{n+}$ entities, HL = pyridine-2-carbaldehyde thiosemicarbazone

Ghosh et al. [194] reported the conversion of methyl 2-pyridyl ketone 4-(4-tolyl)thiosemicarbazone (HL) into methyl(2-pyridyl)methyleneimine (L') by refluxing in ethanol for 24 h a mixture of the preformed $[RuCl(L)(PPh_3)_2]$ compound and a six-fold excess of the HL ligand (Fig. 23).



Fig. 23 Attainment of methyl(2-pyridyl)methyleneimine (L') by transformation of $[RuCl(L)(PPh_3)_2]$ into $[Ru(HL)(L')(PPh_3)]Cl_2$

Thermally activated desulfurizations, for instance by refluxing solvents with high boiling temperature, have been applied for synthetic methods using sulfur derivatives analogous to TSCs, as bisthioureas [195].

d) Oxidative desulfurization of thiosemicarbazones induced by metal ions

The mere coordination of TSC to metal ions can promote desulfurization. Sometimes, this kind of breakage reveals the existence of redox processes. For instance, the reaction of Na[AuCl₄] with the disulfured TSC derived from N-[N',N'- dialkylamino(thiocarbonyl)]benzimidoyl chloride gives rise to a partial reduction of Au(III) to Au(I), as Abram et al. have reported [196]. As a result, apart from the major product I, a certain amount of the ligands undergo oxidative cyclization, through an intermediate thiatriazine-Au(III), leading to the loss of one of their sulfur atoms to yield N-(hexamethylene)-N'-1-(5-diethylamino-3-phenyl-1,2,4-triazolyl)thiourea, that acts as monodentate ligand linking through the remaining sulfur atom to Au(I) ions in the minor product II (Fig. 24).



Fig. 24 Reduction of Au(III) to Au(I) by partial desulfurization of a TSC to give a triazole derivative

In the same way, the reaction of an aqueous solution of $[Cu(HPTSC)(ox)(OH_2)]$, acidified with HNO₃ to pH 0.8 in the presence of an excess of VOSO₄ (molar ratio 1:10), and further addition of base while vigorous stirring to pH 3.7 yielded, after filtration and slow evaporation of the mother liquors, crystals of the $\{[Cu(HPTSC)(OH_2)]_2[Cu(PTSC)S]_2(H_4V_{10}O_{28})\}_n$ compound [197]. The sulfido ligands, arisen from partial desulfurization of the TSC and probably caused by oxidation of V(IV) to V(V), play the role of μ_2 -S²⁻ bridges between $[Cu(PTSC)]^+$ entities to build $[\{Cu(PTSC)S\}_2]^{2-}$ dimers that connect the decavanadate clusters.

Other complex redox process was reported by Dilworth et al. in the reaction of pyridine-2-carbaldehyde thiosemicarbazone (HL) and analogues with the Re(V)-containing $[ReOCl_3(PPh_3)_2]$ compound [198]. Surprisingly, the reaction usually gave rise to $[ReL_2]Cl$ products, where Re(V) had been reduced to Re(III) probably by the released PPh₃ ligands. However, one of the resulting complexes, derived from 2-acetylpyridine thiosemicarbazone, contained a methyl(2-pyridyl)-methyleneimine ligand (L') as a result of a reductive cleavage of the hydrazinic N–N bond in the TSC (Fig. 25). The product, of formula $[ReCl_2L(PPh_3)_2][ReO_4]$ excluding the solvent molecules, contained both Re(III) and Re(VII) ions, the later formed through the redox processes that provoke the breakage of the ligand. In fact, the product in this reaction had a precedent in that reported by Ghosh et al [194] discussed in a previous section.



Fig. 25 The breakage of 2-acetylpyridine thiosemicarbazone to methyl(2-pyridyl)-methyleneimine by redox reaction and further coordination to Re(III) ions

Regarding to the influence of the metal ion coordination in these processes, Souza et al. [199] described that reaction of 3,5-diacetyl-1,2,4-triazole bis(4-ethylthiosemicarbazone) (H₅L) with PdCl₂ and LiCl in methanol led to the tetranuclear [Pd₄(μ_2 - η^2 -S₂)(H₂L)₂] compound, which exhibits a disulfide bridge between the four metal ions whose origin is the partial breakage of some of the TSCs.

Due to the strong analogies with TSCs, we include in this review the paper reported by Duan, He et al. about the fluorescent sensor tetra-2-pyridylthiocarbazone (H₂L), which desulfurized and cyclized to HL' in the presence of Hg(NO₃)₂·0.5H₂O or HgCl₂ [200], and a new pyridazine ring arose upon desulfurization (Fig. 26). The reaction was performed in H₂O/CH₃OH (90:10, v/v) mixtures and led to the attainment of crystals of the [{Hg(L')(SH)}₂] and [Hg(HL')Cl₂] compounds, whose structures were solved. Note that, in the first case, SH⁻ ligands coming from thioamide were coordinated to Hg(II) ions. An increase in the fluorescence intensity at 530 nm accompanied the reaction. No significant changes were observed for other metal ions, as Mg(II), Ca(II), Ba(II), Cr(III), Mn(II), Fe(II), Co(II), Ni(II), Cu(II), Ag(I) and Pb(II). However, the addition Zn(II) and Cd(II) also provoked an increase in the luminescence.



Fig. 26 Tetra-2-pyridylthiocarbazone (H_2L) and corresponding cyclization product (HL'), where the nitrogen positions able to deprotonate are marked with dotted boxes

Another Hg(II)-based chemodosimetric system based on thiosemicarbazone was investigated (Fig. 27). In this work, the conversion of the thiocarbonyl into a carbonyl group selectively exerted by Hg(II) ions and the dimerization of semicarbazone resulted in a pronounced OFF/ON-type fluorescent signalling behaviour [201].



Fig. 27 Desulfurization of 5-(4-nitro)phenyl-2-furaldehyde thiosemicarbazone in the presence of Hg(II) ions and further dimerization of the resulting semicarbazone

Coordination-induced desulfurizations have been reported for acetylthioureas by coordination to Ag^+ ions [202], for 1,3-bis(3-methylpyridin-2-yl)thiourea after reaction with $[PtCl_2(dmso)_2]$ in methanol [203] or, in the same solvent, thiocarboxylates linked to Cu(II) ions to give carboxylates [204], for nickel dithiolenes to give 5,5'-bis(1,3-dialkyl-4-imidazolidine-2-thione-4-thiolate) [205], and thiosemicarbazide coordinated to Cu(II) to provide thiocyanate [206]. It has also been described the Cu(II) assisted desulfurization of 1-R-tetrazole-5-thiols to tetrazoles in ethanol or acetonitrile [207].

Finally, it is noteworthy that different chemical treatments sometimes give rise to the same product. As an example, Marchio et al. reported how a thioxo-1,2,4-triazolecopper(II) compound underwent desulfurization towards the 1,2,4-triazole derivative through different chemical pathways [208], as represented in Fig. 28.



Fig. 28 Chemical versatility of the desulfurization of a thioxo-1,2,4-triazolecopper(II) complex

3. Biological implications

3.1. The physiological relevance of the triggering factors

As mentioned before, several factors have been probed to trigger desulfurization in thiosemicarbazones like pH, oxidants, temperature, radiation or solvents. In this context, it is important to mention that some of these factors intrinsically are crucial in cells, especially the pH and oxidation environments caused by ROS.

Physiologically normal intracellular fluid pH is commonly between 7.0 and 7.4, although there is variability between tissues [209]. However, the pH within organelles is tailored for its specific function. For instance, lysosomes are degradative organelles that need high internal acidity (4.5-5.0 range) to successfully perform their intended function [210]. In contrast, mitochondria have an internal pH of around 8.0, specifically pH 7.6–8.3, which is approximately 0.9 pH units higher than that of intermembrane space to generate large quantities of ATP [211,212]. Regarding the extreme pH values found in the diverse organs of mammals, they range from the gastric pH 1.5–3.5 to the pH 7.35–7.45 in blood [213,214].

Malignant tumour cells have acid pH values, in the 5.6-6.8 range. Thus, every tumour needs to change its metabolism to obtain the energy levels required for its high proliferative rates, and these adaptations lead to alterations in the extra- and intracellular pH. The changes in pH are common to all solid tumours, and can be used either as therapeutic targets, blocking the cell proton transporters and reversing the pH changes, or as means to specifically deliver anticancer drugs [215]. In short, taking all this into account, it can be deduced that the experimental conditions at which TSC-metal complexes show pH-induced desulfurization processes in the laboratory can be easily reached *in vivo*.

The presence of different oxidants in the biological media, mainly in aerobic organisms, is ubiquitous [216,217]. Therein, the attack of these species to the TSC backbone provoking alterations as those described in the previous section is feasible. Oxidant reactants are more likely produced in the chloroplasts, mitochondria, and peroxisomes surroundings [218,219]. The fighting against these entities to maintain the redox balance inside the cell requires the presence of reductant counterparts [220,221]. One of them is glutathione, the most abundant thiol in mammalian cells. The reduced/oxidized glutathione (GSH/GSSG)

couple is the main intracellular regulator of redox homeostasis in animals and plants. GSH controls the thiol oxidation state of proteins and acts as defence against oxidative stress, by directly scavenging ROS and by repairing their damage via enzymatic processes. Thus, GSH is a crucial compound for living cells, and targeting GSH metabolism is of wide interest for therapeutic purposes, in particular for fighting against cancer [222]. The activity of GSH strongly depends on different factors, as the presence of metal ions [223,224] or the pH [225]. In particular, the reactivity against reduced glutathione and, therefore, the production of ROS, seems to be notably enhanced with acidity in thiosemicarbazonecopper(II) complexes [226]. So, the interplay between the TSCs breakage and the increase in acidity of the medium could drastically modify the biological activity of these compounds.

The normal body temperature value about 36-37 °C could be non-innocent in several processes when it combines with other factors, as neutral-basic media, in the way we have shown in Section 2.1.

3.2. Detection of TSC desulfurization in in vitro, in vivo and clinical assays

The actual way of assessing the extent of the desulfurization of TSCs during their biological activity is the detection of these processes *in vivo*. One of the first works in this field suggested the transformation of 1-methylisatin 3-thiosemicarbazone into *syn* and *anti* isomers of 1-methylisatin 3-semicarbazone in incubation medium of the fungus Cladosporium resinae [227].

Few years after, it was published the fundamental paper by Sartorelli et al. dealing with the antineoplastic activity of 5-hydroxypyridine-2-carbaldehyde thiosemicarbazone (5-HP, Fig. 3) and isoquinoline-1-carbaldehyde thiosemicarbazone (IQ-1) on mice and dogs [228]. They used ¹⁴C and ³⁵S isotopically labelled thiosemicarbazones to verify both the tissue distribution and the nature of the metabolites excreted in urine. The results showed that metabolic pathways for IQ-1 led to extensive desulfurization with liberation of sulfate as the major contribution, 59.6 % of the excreted derivatives. A second contribution was the side-chain cleavage with release of CO₂ (up to 20 % in mice), urea and thiourea (6.4 %) and the hydrolysis to semicarbazide and thiosemicarbazide (6.0 %). In fact, the breakage of the TSC chain actually seemed to obey to two types of cleavage. An attack on the hydrazine =N–NH– moiety, to yield thiourea and urea, was tentatively attributed to the action of azoreductase enzymes. In this case, the amount of urea was found to be about 4 % of urinary radioactivity, suggesting that desulfurization does not need to precede the TSC chain cleavage. In addition, there was hydrolysis of the azomethine –CH=N– fragment yielding thiosemicarbazide and semicarbazide fragments. Finally, a 13.3 % was apparently in form of glucuronides formed by ring hydroxylation and further reaction of the hydroxylated IQ-1 derivatives with glucuronic acid.

Analogous studies were performed by the same research team with 5-HP during the first Phase I clinical trials on TSCs reported in the literature [229]. They showed that glucuronide of 5-HP was the major component of the urinary label (41-62 %) because of the reaction of the hydroxyl substituent. No measurable amounts of urea, thiourea or semicarbazide could be found, indicating that the integrity of the TSC chain in 5-HP was unaffected by the metabolism in humans.

Precisely the discouraging results obtained in those first clinical trials caused, at least in part, a thirty-year hiatus until a new member of the family of TSCs was tested in humans. It was the 3-aminopyridine-2-carbaldehyde thiosemicarbazone (3-AP, Fig. 29). Kowol et al. carried out an extensive study on the metabolism of 3-AP by electrochemical techniques that simulated oxidative liver reactions, together with analysis of human liver microsomes and *in vivo* distribution experiments in mice [230]. As a result, several metabolites were identified, among them small amounts of the desulfurated formamidine derivative, depicted as (II) in Fig. 29. This oxidative desulfurization would proceed through the oxidation of the thioamide to give $-NH-C(=SO_2)-NH_2$, and further release of SO₂ and attainment of the formamidine $-N=CH-NH_2$.



Fig. 29 Chemical structures of 3-AP (I) and the related formamidine (II)

The above-mentioned investigation was extended to 10 derivatives of 3-AP [159]. The panoplia of metabolic pathways involving oxidative desulfurization processes are represented in Fig. 30, and includes semicarbazone (II), amidrazone (IV) and the formation of ring-closed 1,3,4-oxadiazole (III) and triazole (V). The presence of semicarbazone (II) and amidrazone (IV) was detected for all the studied TSCs, however in some cases the amount of them was low. Dehydrogenation after the oxidative desulfurization led to the oxadiazole and triazole rings, which were not always observed. It is worth mentioning that the value of the m/z peak attributed to (V), about 190.11, entirely coincides with the possible nitrile-derivative of (IV) analogous to that described in Section 2.1.



Fig. 30 Metabolites of 3-AP derivatives

Kovarikova et al. [231] performed an *in vivo* and *in vitro* liquid chromatography tandem mass spectrometry analysis (LC-MS/MS) to characterize the metabolites of 2-benzoylpyridine 4-ethyl-3-thiosemicarbazone (Bp4eT, Fig. 31). The studies *in vitro* were carried out using subcellular fractions of rat microsomes and cytosol, and male human liver samples. Male rats were used for the experiments *in vivo*. They found two metabolites of Bp4eT *in vitro* arisen from desulfurization: benzoylpyridine 4-ethylsemicarbazone (II) and N3-ethyl-N1-[phenyl(pyridine-2yl)methylene] formamidrazone (III). Both metabolites were also discovered *in vivo*, being present in plasma, urine, and feces, together with a new metabolite tentatively identified as a hydroxylated form of the amidrazone (III). More recent studies performed by Richardson and Kovarikova on DpC (di(2-pyridyl)ketone 4-cyclohexyl-4-methyl-3-thiosemicarbazone) and Dp44mT (di(2-pyridyl)ketone 4,4-dimethyl-3-thiosemicarbazone) found negligible oxidation of the thioamide to give formamidrazone metabolites, discarding a relevant role in the toxicity or efficacy of the products arisen from slow hydrolytic processes [232].



Fig. 31 Metabolites of 2-benzoylpyridine 4-ethyl-3-thiosemicarbazone (Bp4eT, I): 2-benzoylpyridine 4ethylsemicarbazone (II) and N3-ethyl-N1-[phenyl(pyridine-2yl)methylene] formamidrazone (III)

3.3. The biological activity of the products arisen from the desulfurization reactions

It is well known since the early studies in the 60s of the last century that semicarbazones, which sometimes derived from desulfurization processes of TSCs *in vivo*, use to exhibit a lesser biological activity than the analogous TSCs [81, 233]. Forty years after, Richardson et al. compared the anti-proliferative activity of

different iron chelators derived from di-2-pyridylketone isonicotinoyl hydrazone [234]. They found that NNS thiohydrazones were 103-134-fold more active than their corresponding NNO carbonyl ligands. Surprisingly, the antiproliferative effects of the NNO ligands were 2-8-fold greater than the parent SNS compounds. Their work suggested that the antiproliferative activity of the NNS thiohydrazones were related to the low redox potentials of the NNS-Fe complexes, facilitating the generation of ROS. In fact, thioamide group was found to facilitate reversible Fe(III/II) reactions, in contrast to the amide moiety [235].

In the same way, Simunek et al. [236] described that the amidrazone and the semicarbazone metabolites arisen from desulfurization reactions of 2-benzoylpyridine 4-ethyl-3-thiosemicarbazone (Bp4eT, Fig. 31) showed more than 300-fold less cytotoxic activity than Bp4eT towards both cancer and normal cell lines.

The triazole-Au(I) (II) derivative described by Abram et al. [196] (Fig. 24), arisen from cyclization and desulfurization of TSC after reduction of Au(III), showed a much less activity against human MCF-7 breast cancer cells than its TSC-Au(III) counterpart (I).

It is, however, well established that 1,3,4-oxadiazoles show interesting biological properties [237]. Notwithstanding, as far as we are aware, no comparison has been carried out among them and their parent TSCs, in spite of the synthetic relationships due to the use of acetylthiosemicarbazides to prepare oxadiazole derivatives [176,182].

Finally, the inorganic sulfur species generated in these processes, as elemental sulfur, hydrogen sulfide and sulfate, play different biological roles in living beings. Sulfur is an essential element for life, being present in peptides, as amino acids and proteins, vitamins (thiamin, biotin, and coenzyme A) and several biomolecules as glycosaminoglycans, chondroitin sulfate, dermatan sulfate, and hyaluronic acid, among others [238]. However, elemental S_8 sulfur is toxic, and it is oxidized during the metabolic pathways to generate sulfite and sulfate [239]. Sulfate homeostasis in human body is maintained, at least in part, through renal clearance mechanisms. This oxoanion acts as detoxification agent and is necessary for the biosynthesis of numerous biomolecules [240]. On the other hand, despite its toxicity, hydrogen sulfide is involved in many physiological processes, including relaxation of vascular smooth muscles, mediation of neurotransmission, inhibition of insulin signalling and regulation of inflammation [241]. It is known long time ago that methemoglobine interacts with H₂S in the presence of oxygen to give a green compound, which has been modelled through the preparation and structural resolution of an analogous complex [242]. Notwithstanding the variety of biological processes and metabolic pathways involving sulfur species, the amounts of these arisen from TSCs desulfurization and the influence in the therapeutic activity of TSCs is far to be understood.

4. Conclusions

In summary, different chemical factors can trigger desulfurization reactions in TSCs. Among them, the most thoroughly studied are (not always) extreme pH values, the presence of oxidizing agents or the establishment of redox processes as those electrochemically induced, the influence of the solvent, the temperature and the electromagnetic radiation. Many of these reactions only occur when TSCs are bound to metal ions. Taking into account that coordination to intracellular essential metal elements, in particular Cu and Fe, seems to be pivotal for the biological activity of TSCs, the possibility for the desulfurization of the ligand *in vivo* becomes relevant. In fact, some physiological conditions reproduce the environments that make possible the desulfurization process (pH, temperature, chemicals...) and, actually, desulfurizations of TSCs have been detected in experiments performed *in vitro* and *in vivo*. These desulfurization processes lead to different products, as semicarbazone and semicarbazide, amidrazone, urea, sulfate, sufide and elemental sulfur, among others. The products often exhibit less activity than intact TSCs. In any case, despite the early detection of some of the species generated from such processes *in vivo*, further studies should be performed to identify new products and to elucidate the actual transcendence of these reactions at a cellular level. These results could shed light into pharmacokinetics leading to improvements in the use of TSCs as drugs.

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The authors declare they have no financial interests.

Author contributions

RP and JGT contributed to the idea, conception, and design of the present work. All the authors performed the literature search and analysis, and equally contributed to the writing of this manuscript. All the authors read and approved the final manuscript.

Declarations

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References

1. Gingras BA, Somorjai RI, Bayley CH (1961) The preparation of some thiosemicarbazones and their copper complexes. Can J Chem 39:973-985. https://doi.org/10.1139/v61-122

2. Benns BG, Gingras BA, Bayley CH (1960) Antifungal activity of some thiosemicarbazones and their copper complexes. Appl Microbiol 8:353-356. https://doi: 10.1128/am.8.6.353-356.1960

3. Singh RB, Ishii H (1991) Analytical potentialities of thiosemicarbazones and semicarbazones. Crit Rev Anal Chem 22:381-409. https://doi.org/10.1080/10408349108051640

4. Arion VB (2019) Coordination chemistry of S-substituted isothiosemicarbazides and isothiosemicarbazones. Coord Chem Rev 387:348-397. https://doi.org/10.1016/j.ccr.2019.02.013

5. González-Barcia LM, Romero MJ, González-Noya AM, Bermejo MR, Maneiro M, Zaragoza G, Pedrido R (2016)

"The Golden Method": Electrochemical synthesis is an efficient route to gold complexes. Inorg Chem 55:7823-7825. https://doi.org/10.1021/acs.inorgchem.6b01362

6. González-Barcia LM, Fernández-Fariña S, Rodríguez-Silva L, Bermejo MR, González-Noya AM, Pedrido R (2020) Comparative study of the antitumoral activity of phosphine-thiosemicarbazone gold(I) complexes obtained by different methodologies. J Inorg Biochem 203:110931-110940. https://doi.org/10.1016/j.jinorgbio.2019.110931

7. Lobana TS, Sharma R, Bawa G, Khanna S (2009) Bonding and structure trends of thiosemicarbazone derivatives of metals-an overview. Coord Chem Rev 253:977-1055. https://doi.org/10.1016/j.ccr.2008.07.004

 Osman UM, Silvarajoo S, Kamarudin KH, Tahir MIM, Kwong HC (2021) Ni(II) complex containing a thiosemicarbazone ligand: synthesis, spectroscopy, single-crystal X-ray crystallographic and conductivity studies. J Mol Struct 1223:128994. https://doi.org/10.1016/j.molstruc.2020.128994

9. Macalik L, Pyrkosz-Bulska M, Małecki G, Hermanowicz K, Solarz P, Janczak J, Hanuza J (2021) Synthesis, structural and spectroscopic properties of [N'-[(2,4-dihydroxyphenyl) methylidene]-4-(4-fluorophenyl) piperazine-1-carbothiohydrazide] thiosemicarbazone and its terbium complex. Inorg Chem Commun 123:108351. https://doi.org/10.1016/j.inoche.2020.108351

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10. Yousef TA, Abu El-Reash GM (2020) Synthesis, and biological evaluation of complexes based on thiosemicarbazone ligand. J Mol Struct 1201:127180. https://doi.org/10.1016/j.molstruc.2019.127180

11. West DX, Liberta AE, Yerande RG (1993) Thiosemicarbazone complexes of copper(II): structural and biological studies. Coord Chem Rev 123:49-71.

12. Beiles RH, Calvin M (1947) The oxygen-carrying synthetic chelate compounds. VII. Preparation. J Am Chem Soc 69:1886-1893. https://doi.org/10.1021/ja01200a013

13. Al-Jeboori M, Dawood AH (2008) Synthesis and structural studies of novel 2,6-diformyl-p-cresol bis-(thiosemicarbazone) ligand and their binuclear complexes with Ni⁺², Pd⁺², Zn⁺², Cd⁺² and Hg⁺² metal ions. J Kerbala Univ, 6:133.

14. Chandra S, Sangeetika C, Rathi A (2001) Magnetic and spectral studies on copper (II) complexes of NO and NS donor ligands. J Saudi Chem Soc 5:175-182.

15. Sahin M, Bal-Demirci T, Pozan-Soylu G, Ülküseven B (2009) Synthesis, characterization and thermal decomposition of dioxouranium(VI) complexes with *N*¹,*N*⁴-diarylidene-*S*-propyl-thiosemicarbazone: crystal structure of [UO₂(L¹)(C₄H₉OH)]. Inorg Chim Acta 362:2407-2412. https://doi.org/10.1016/j.ica.2008.10.036

16. Ferrari MB, Bisceglie F, Pelosi G, Tarasconi P, Albertini R, Dall'Aglio PP, Pinelli S, Bergamo A, Sava G (2004)
Synthesis, characterization and biological activity of copper complexes with pyridoxal thiosemicarbazone derivatives.
X-ray crystal structure of three dimeric complexes. J Inorg Biochem, 98:301-312.
https://doi.org/10.1016/j.jinorgbio.2003.09.011

17. Christlieb M, Dilworth JR (2016) Ligands for molecular imaging: the synthesis of bis(thiosemicarbazone) ligands. Chem Eur J 12:6194-6206. https://doi.org/10.1002/chem.200501069

18. Hałdys K, Latajka R (2019) Thiosemicarbazones with tyrosinase inhibitory activity. MedChemComm 10:378–389. https://doi.org/10.1039/c9md00005d

19. Paterson BM, Donnelly PS (2011) Copper complexes of bis(thiosemicarbazones): from chemotherapeutics to diagnostic and therapeutic radiopharmaceuticals. Chem Soc Rev 40:3005-3018. https://doi.org/10.1039/c0cs00215a

20. Gupta S, Singh N, Khan T, Joshi S (2022) Thiosemicarbazone derivatives of transition metals as multi-target drugs: a review. Results Chem 4:100459. https://doi.org/10.1016/j.rechem.2022.100459

21. French FA, Blanz Jr E J (1965) The carcinostatic activity of α-(N) heterocyclic carboxaldehyde thiosemicarbazones:
I. Isoquinoline-1-carboxaldehyde thiosemicarbazone. Cancer research, 25:1454-1458.

22. Campbell MJM (1975) Transition metal complexes of thiosemicarbazide and thiosemicarbazones. Coord Chem Rev 15:279-319. https://doi.org/10.1016/S0010-8545(00)80276-3

23. Padhyé S, Kauffman GB (1985) Transition metal complexes of semicarbazones and thiosemicarbazones. Coord Chem Rev 63:127-160. https://doi.org/10.1016/0010-8545(85)80022-9

24. Casas JS, García-Tasende MS, Sordo J (2000) Main group metal complexes of semicarbazones and thiosemicarbazones. A structural review. Coord Chem Rev 209:197-261. https://doi.org/10.1016/S0010-8545(00)00363-5

25. Quiroga AG, Ranninger CN (2004) Contribution to the SAR field of metallated and coordination complexes: studies of the palladium and platinum derivatives with selected thiosemicarbazones as antitumoral drugs. Coord Chem Rev 248:119-133. https://doi.org/10.1016/j.cct.2003.11.004.

26 Devi J, Kumar B, Taxak B (2022) Recent advancements of organotin(IV) complexes derived from hydrazone and thiosemicarbazone ligands as potential anticancer agents. Inorg Chem Commun 139:109208. https://doi.org/10.1016/j.inoche.2022.109208.

27. Kalinowski DS, Quach P, Richardson DR (2009) Thiosemicarbazones: The new wave in cancer treatment. Future Med Chem 1:1143-1151. https://doi.org/10.4155/fmc.09.80.

73. Qi J, Liang S, Gou Y, Zhang Z, Zhou Z, Yang F, Liang H (2015) Synthesis of four binuclear copper(II) complexes: Structure, anticancer properties and anticancer mechanism. Eur J Med Chem 96:360-368. https://doi.org/10.1016/j.ejmech.2015.04.031

74. Menezes SV, Sahni S, Kovacevic Z, Richardson DR (2017) Interplay of the iron-regulated metastasis suppressor NDRG1 with epidermal growth factor receptor (EGFR) and oncogenic signaling. J Biol Chem 292:12772-12782. https://doi.org/10.1074/jbc.R117.776393

75. Zou BQ, Lu X, Qin QP, Bai YX, Zhang Y, Wang M, Liu YC, Chen ZF, Liang H (2017) Three novel transition metal complexes of 6-methyl-2-oxo-quinoline-3-carbaldehyde thiosemicarbazone: synthesis, crystal structure, cytotoxicity, and mechanism of action. RSC Adv 7:17923-17933. https://doi.org/10.1039/C7RA00826K

76. Brodowska K, Correia I, Garribba E, Marques F, Klewicka E, Lodyga-Chruscińska E, Pessoa JC, Dzeikala A, Chrusciński L (2016) Coordination ability and biological activity of a naringenin thiosemicarbazone. J Inorg Biochem 165:36-48. https://doi.org/10.1016/j.jinorgbio.2016.09.014

77. Basha MT, Bordini J, Richardson DR, Martinez M, Bernhardt PV (2016) Kinetico-mechanistic studies on methemoglobin generation by biologically active thiosemicarbazone iron(III) complexes. J Inorg Biochem 162:326333. https://doi.org/10.1016/j.jinorgbio.2015.12.004

78. Pape VFS, Tóth S, Füredi A, Szebényi K, Lovrics A, Szabó P, Wiese M, Szakács G (2016) Design, synthesis, and biological evaluation of thiosemicarbazones, hydrazinobenzothiazoles and arylhydrazones as anticancer agents with a potential to overcome multidrug resistance. Eur J Med Chem 117:335-354. https://doi.org/10.1016/j.ejmech.2016.03.078

79. Myers CR (2016) Enhanced targeting of mitochondrial peroxide defense by the combined use of thiosemicarbazones and inhibitors of thioredoxin reductase. Free Radic Biol Med 91:81-92. https://doi.org/10.1016/j.freeradbiomed.2015.12.008

80. Fu Y, Liu Y, Wang J, Li C, Zhou S, Yang Y, Zhou P, Lu C, Li C (2017) Calcium release induced by 2pyridinecarboxaldehyde thiosemicarbazone and its copper complex contributes to tumor cell death. Oncol Rep 37:1662-1670. https://doi.org/10.3892/or.2017.5395

81. French FA, Blanz EJ (1966) The carcinostatic activity of thiosemicarbazones of formyl heteroaromatic compounds.1 III. Primary correlation. J Med Chem 9:585-589. https://doi.org/10.1021/jm00322a032

82. Agrawal KC, Sartorelli AC (1975) Alpha-(N)-heterocyclic carboxaldehyde thiosemicarbazones. In: Sartorelli AC, Johns DG (eds) Handbook of Experimental Pharmacology. Springer-Verlag, Berlin, pp 793-807.

83. Sartorelli AC, Agrawal KC, Tsiftsoglou AS, Moore EC (1977) Characterization of the biochemical mechanism of action of α -(N)-heterocyclic carboxaldehyde thiosemicarbazones. Adv Enzyme Regul 15:117-139. https://doi.org/10.1016/0065-2571(77)90012-7

 Kunos CA, Ivy SP (2018) Triapine radiochemotherapy in advanced stage cervical cancer. Front. Oncol 8:149-155. https://doi.org/10.3389/fonc.2018.00149

85. Kolesar J, Brundage RC, Pomplun M, Alberti D, Holen K, Traynor A, Ivy P, Wilding G (2011) Population pharmacokinetics of 3-aminopyridine-2-carboxaldehyde thiosemicarbazone (Triapine[®]) in cancer patients. Cancer Chemother Pharmacol 67:393-400. https://doi.org/10.1007/s00280-010-1331-z

86. Miah AB, Harrington KJ and Nutting CM (2010) Triapine® in clinical practice. Eur J Clin Med Oncol 2:87-92.

87. Guo ZL, Richardson DR, Kalinowski DS, Kovacevic Z, Tan-Un KC, Chan GCF (2016) The novel thiosemicarbazone, di-2-pyridylketone 4-cyclohexyl-4-methyl-3-thiosemicarbazone (DpC), inhibits neuroblastoma growth in vitro and in vivo via multiple mechanisms. J Hematol Oncol 9:98-114. https://doi.org/10.1186/s13045-016-0330-x

88. Serda M, Kalinowski DS, Rasko N, Potučková E, Mrozek-Wilczkiewicz A, Musiol R, Małecki JG, Sajewicz M, Ratuszna A, Muchowicz A, Gołąb J, Šimunek T, Richardson DR, Polanski J (2014) Exploring the anti-cancer activity



Fig. 13 Proposed mechanism for the desulfurization of pyridine-2-carboxaldehyde-4-*N*-ethyl-thiosemicarbazonecopper(II) complexes to form sulfate ions

Metal-free thiosemicarbazones have also been electrochemically studied, in particular isatin derivatives. Thus, upon oxidation, it was proposed the breakage the of the C=N bond, generating isatin and thiourea fragments for all evaluated molecules (Fig. 14). Regarding the reduction, the cleavage of the N-N bond as well as the generation of 3-aminoindolin-2-one and thiourea moieties was proposed [158].



Fig. 14 Electrochemically induced cleavage of isatin thiosemicarbazone

Electrochemistry, coupled to high resolution mass spectrometry (HRMS), was also employed to investigate the possible relationship between the structure of α -N-heterocyclic thiosemicarbazones and their metabolic behaviour. To this end, the metabolites of ten different Triapine[®] derivatives with a wide range of antitumour activities were analysed. In general, for all the investigated thiosemicarbazones, the identified processes of the metabolic reactions were hydroxylation, oxidative desulfurization (formation of the amidrazone and, for some derivatives, also the semicarbazone) and disulfide dimer formation and dehydrogenation in some cases (Fig. 15). In general, desulfurization was detected for all the investigated compounds, thus confirming that the study of desulfurization process is crucial for a better understanding of the thiosemicarbazones biological activity [159].



Fig. 15 Key metabolites of Triapine®: the dehydrogenated ring-closed thiadiazole and hydroxylated species

2.3. Desulfurizations by oxidant reactants

The interaction between thiosemicarbazone complexes and oxidants can give rise to desulfurization by reactions often involving oxidative cyclization processes. Some of them are described in the present section.

a) Conversion of thiosemicarbazones into 1,3,4-oxadiazoles by halate ions

The addition of bromate to aqueous solutions of HL (where HL = HPTSC and HPTSC4m) triggered a complex process whose main reaction led to 1,3,4-oxadiazole derivatives [160,161], that could be regarded as is shown in Fig. 16.



Fig. 16 Transformation of thiosemicarbazones into 1,3,4-oxadiazoles. Thiosemicarbazones: pyridine–2– carbaldehyde thiosemicarbazone (R = H, HL = HPTSC) and pyridine-2-carbaldehyde 4-Nmethylthiosemicarbazone ($R = CH_3$, HL = HPTSC4m). Oxadiazoles: 2-amino-5-pyridin-2-yl-1,3,4oxadiazole (R = H, $L_{oxad} = 134OXAD$) and 2-methylamino-5-pyridin-2-yl-1,3,4-oxadiazole ($R = CH_3$, $L_{oxad} = 134OXADm$)

The study of different well-characterized solids isolated from this process allowed to distinguish several steps, which are drawn in Fig. 17, despite some stages in the mechanism remain unclear. The reaction proceeded smoothly with the addition of bromate to aqueous solutions of preformed CuL(NO₃) complexes at pH ~ 6 (Step 3). An olive-green precipitate corresponding to the centrosymmetric S-bridged [{CuLBr}₂] dimer appeared (Step 3) and, after filtering it, the initial dark green colour of the solution gradually became lighter. Simultaneusly, the pH decreased to 3–4 and an irritant gas was released. Two days later, single crystals of [{Cu(L_{oxad})(OH₂)₂(OSO₃)}₂] were obtained (Step 4). The addition of K₄[Fe(CN)₆]·3H₂O led to the coprecipitation of purple K₂Cu[Fe(CN)₆]·H₂O and L_{oxad}, whose particle sizes were different enough to eliminate the complex in the filtrate while the white organic compound was retained in the filter and, afterwards, recrystallized in ethanol (5). The use of NaHCO₃ instead of K₄[Fe(CN)₆]·3H₂O also allowed to isolate the oxadiazole ligand. The rate of the reaction was increased with heating and addition of small

amounts of acid. However, brown suspensions were attained in the excess of acid or strong heating, whose filtration allowed to isolate crystals of bis(pyridine-2-carboxylate)copper(II) (6).

The reaction also took place without Cu(II) ions, but strong acid media was required (pH = 0–1) and, after filtration of a dark red unidentified solid, further treatment with NaOH to pH = 3–4. In the presence of acid media, mixtures of compounds were invariably obtained and purification was needed. The use of KIO₃ in acid media yielded the same results than KBrO₃, but the formation of iodine, probably due to comproportation between I⁻ and IO₃⁻, made more difficult the purification of the product. Furthermore, the addition of KIO₃ to the CuL(NO₃) complexes at pH ~ 6 led, firstly, to the attainment of a green compound identified as CuL(IO₃), which evolved when the suspension was kept with stirring to yield [{CuLI₂] dinuclear compounds in a final step, but oxadiazole derivatives were not attained. The analogous CuL(BrO₃) compound could not be isolated in these experiments. On the contrary, none of the attempts carried out by using chlorate as oxidant gave any evidence for oxidative cyclization, which could be due to kinetic factors. Finally, no oxidation compounds were attained by using pyridine–2–carbaldehyde 4,4'-N-dimethylthiosemicarbazone (HPTSC44m) neither free nor coordinated to Cu(II) ions, which suggests steric influences in the process.



Fig. 17 Reaction pathways identified for the different reactants and conditions

A plausible mechanism can be proposed taking into account the experimental evidences, which is depicted in Fig. 18 for the free ligand, but it could be extrapolate, at least in part, for the Cu(II) complexes.



Fig. 18 Mechanism suggested for the halate-induced oxidative cyclization of TSCs to 1,3,4-oxadiazoles

These processes allow to prepare 2-amino-1,3,4-oxadiazoles as the only product through easy and unexpensive reactions, sometimes in less than an hour.

Oxidations of acyl thiosemicarbazides, acyl carbodithioates and acyl thioureas to oxadiazoles, often promoted by metal ions, have been frecuently described in the literature [162-183].

b) Other desulfurization of thiosemicarbazones by oxidant reactants

The use of other oxidants to desulfurize and cyclize TSCs has been described for a long time, as it was reported by Landquist in the attainment of triazolinones by reaction with MnO_2 [184].

From a broader point of view apart from TSCs, the use of oxidants as H_2O_2 , 1O_2 and I_2 has been described for the desulfurization of other sulfur-containing molecules, as 8-thioguanosine to guanosine [185] or dithiocarbamato-Ru(II) complexes [186,187], among others.

Desulfurization of TSCs through processes involving redox-active metal ions is discussed in Section 2.4.d.

2.4. Others (radiation-, thermal-, solvent- and coordination-induced desulfurizations)

a) Solvent-induced desulfurizations

Another factors like light, pressure or solvent must be considered to have a relevant role in thiosemicarbazone desulfurization reactions. In this sense, recently it was presented the dinuclear helicate $[Ag_2(H_2L)_2]SO_4$ obtained by crystallization of the bisthiosemicarbazone cluster helicate $[Ag_4L_2]_2$ in the absence of any sulfate source, after a rare desulfurization process that takes place only in chloroform (Fig. 19). Three factors were investigated in this conversion: solvent, light and time. Only those recrystallizations performed in chloroform in the presence of light led to the formation of the sulfate helicate crystals. Regarding to time, the appearance of the sulfate crystals took place after a long crystallization period of 3-4 weeks. Some ¹H NMR studies mimicking the recrystalization process by protonation of the hydrazine NH groups, thus resulting on the dihelicate unit $[Ag_2(H_2L)_2]^{2+}$. It is relevant to mention that two side-products were identified along the desulfurization: silver hydrogen sulfide/sulfide, that easily evolved to the sulfate counterion by oxidation by moisture oxygen, and the organic fragment semicarbazone released

from the oxidative desulfurization. Finally, the remaining dinuclear complex combined with the sulfate thus generating the final crystallized sulfate dihelicate $[Ag_2(H_2L)_2]SO_4[188]$.



Fig. 19 Formation of the dinuclear helicate $[Ag_2(H_2L)_2](SO_4)$ by desulfurization of the doubletetranuclear cluster helicate $[Ag_4L_2]_2$

Notwithstanding, it must be admitted that not always coordination to metal ions is required to provoke solvent-induced desulfurization in TSCs. For instance, Peach and Dilworth et al. [189] described intramolecular cyclization reactions in recrystallization experiments of bisthiosemicarbazones that involved the loss of one of the thioamide sulfur atoms. The isolated product depended on the kind of solvent used, as is shown in Fig. 20.



Fig. 20 Benzil bis(4-phenyl-3-thiosemicarbazone) and related cyclized products of solvent-induced desulfurization processes

b) The influence of the electromagnetic radiation

Microwave radiation is known to promote desulfurization in organo-sulfur compounds being this process of great importance for reducing harmful emissions during the combustion process [190]. Microwave irradiation of thiosemicarbazones gave the corresponding isothiocyanates, which on addition of either activated nitriles or aldehydes furnished various types of azines (Fig. 21) [191]. This process was also reported for thiosemicarbazone analogues, as thiocarbohydrazones [192].



Fig. 21 Azine fragments formed after microwave irradiation of thiosemicarbazones

Visible light has been used as an agent for desulfurization of thiols and disulfides [193]. However, as far as we are aware, none of these processes has been reported for TSCs.

c) Thermal induced desulfurizations

Heating is another point to be considered in desulfurization processes experienced by TSC complexes. Thus, prolonged treatment in refluxing aqueous solutions of the complex $CuL(NO_3)$, being HL= pyridine-2-carbaldehyde thiosemicarbazone, induced breakage of a portion of the thiosemicarbazone molecules to give HS⁻ ligands that were identified in the structure of [{CuL(SH)}₂], whose crystals were collected from the mother liquors (Fig. 22). In contrast, the free ligands remain unaltered under the same experimental conditions. Extending the studies to other metal ions, as Fe(III), Co(III), Zn(II) and Pb(II), it was found that only bis(thiosemicarbazonato)iron(III) species underwent breakage whereas there was no evidence for desulfurization processes in the Co(III), Zn(II) and Pb(II) derivatives. The IR data suggested that breakage of HL ligands gave rise to thiocyanato ligands in the above mentioned Cu(II) derivatives, while a non-coordinated thiocyanate anion was present in the Fe(III) decomposition product [137].



Fig. 22 Summary of desulfurization processes by refluxing on $[ML]^+$ and $[ML_2]_{n+}$ entities, HL = pyridine-2-carbaldehyde thiosemicarbazone

Ghosh et al. [194] reported the conversion of methyl 2-pyridyl ketone 4-(4-tolyl)thiosemicarbazone (HL) into methyl(2-pyridyl)methyleneimine (L') by refluxing in ethanol for 24 h a mixture of the preformed $[RuCl(L)(PPh_3)_2]$ compound and a six-fold excess of the HL ligand (Fig. 23).



Fig. 23 Attainment of methyl(2-pyridyl)methyleneimine (L') by transformation of $[RuCl(L)(PPh_3)_2]$ into $[Ru(HL)(L')(PPh_3)]Cl_2$

Thermally activated desulfurizations, for instance by refluxing solvents with high boiling temperature, have been applied for synthetic methods using sulfur derivatives analogous to TSCs, as bisthioureas [195].

d) Oxidative desulfurization of thiosemicarbazones induced by metal ions

The mere coordination of TSC to metal ions can promote desulfurization. Sometimes, this kind of breakage reveals the existence of redox processes. For instance, the reaction of Na[AuCl₄] with the disulfured TSC derived from N-[N',N'- dialkylamino(thiocarbonyl)]benzimidoyl chloride gives rise to a partial reduction of Au(III) to Au(I), as Abram et al. have reported [196]. As a result, apart from the major product I, a certain amount of the ligands undergo oxidative cyclization, through an intermediate thiatriazine-Au(III), leading to the loss of one of their sulfur atoms to yield N-(hexamethylene)-N'-1-(5-diethylamino-3-phenyl-1,2,4-triazolyl)thiourea, that acts as monodentate ligand linking through the remaining sulfur atom to Au(I) ions in the minor product II (Fig. 24).



Fig. 24 Reduction of Au(III) to Au(I) by partial desulfurization of a TSC to give a triazole derivative

In the same way, the reaction of an aqueous solution of $[Cu(HPTSC)(ox)(OH_2)]$, acidified with HNO₃ to pH 0.8 in the presence of an excess of VOSO₄ (molar ratio 1:10), and further addition of base while vigorous stirring to pH 3.7 yielded, after filtration and slow evaporation of the mother liquors, crystals of the $\{[Cu(HPTSC)(OH_2)]_2[Cu(PTSC)S]_2(H_4V_{10}O_{28})\}_n$ compound [197]. The sulfido ligands, arisen from partial desulfurization of the TSC and probably caused by oxidation of V(IV) to V(V), play the role of μ_2 -S²⁻ bridges between $[Cu(PTSC)]^+$ entities to build $[\{Cu(PTSC)S\}_2]^{2-}$ dimers that connect the decavanadate clusters.

Other complex redox process was reported by Dilworth et al. in the reaction of pyridine-2-carbaldehyde thiosemicarbazone (HL) and analogues with the Re(V)-containing $[ReOCl_3(PPh_3)_2]$ compound [198]. Surprisingly, the reaction usually gave rise to $[ReL_2]Cl$ products, where Re(V) had been reduced to Re(III) probably by the released PPh₃ ligands. However, one of the resulting complexes, derived from 2-acetylpyridine thiosemicarbazone, contained a methyl(2-pyridyl)-methyleneimine ligand (L') as a result of a reductive cleavage of the hydrazinic N–N bond in the TSC (Fig. 25). The product, of formula $[ReCl_2L(PPh_3)_2][ReO_4]$ excluding the solvent molecules, contained both Re(III) and Re(VII) ions, the later formed through the redox processes that provoke the breakage of the ligand. In fact, the product in this reaction had a precedent in that reported by Ghosh et al [194] discussed in a previous section.

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of novel thiosemicarbazones generated through the combination of retro-fragments: dissection of critical structureactivity relationships. PLoS One 9:e110291. https://doi.org/10.1371/journal.pone.0110291

89. https://clinicaltrials.gov/ct2/show/NCT02688101?term=DpC&draw=2&rank=1 (access 21.06.2023)

90. https://clinicaltrials.gov/ct2/show/NCT02433626?term=thiose-micarbazone&draw=2&rank=9 (access 21.06.2023)

91. https://classic.clinicaltrials.gov/ct2/show/NCT02466971?term=thiosemicarbazone&draw=2 (access 21.06.2023).

92. Sartorelli AC, Booth BA (1967) Inhibition of the growth of sarcoma 180 ascites cells by combinations of inhibitors of nucleic acid biosynthesis and the cupric chelate of kethoxal bis-(thiosemicarbazone). Cancer Research 27:1614-1619.

93. Nutting CM, van Herpen, CML, Miah, AB (2009) Phase II study of 3-AP Triapine in patients with recurrent or metastatic head and neck squamous cell carcinoma. Annals Oncol 20:1275-1279. https://doi.org/10.1093/annonc/mdn775

94. Ma B, Goh BC, Tan EH, Lam KC, Soo R, Leong SS, Wang LZ, Mo F, Chan ATC, Zee B, Mok T (2008) A multicenter phase II trial of 3-aminopyridine-2-carboxaldehyde thiosemicarbazone (3-AP, Triapine[®]) and gemcitabine in advanced non-small-cell lung cancer with pharmacokinetic evaluation using peripheral blood mononuclear cells. Investigational New Drugs 26:169-173. https://doi.org/10.1007/s10637-007-9085-0

95. Priyarega S, Haribabu J, Karvembu R (2022) Development of thiosemicarbazone-based transition metal complexes as homogeneous catalysts for various organic transformations. Inorganica Chim Acta 532:120742. https://doi.org/10.1016/j.ica.2021.120742

96. Gil-García R, Ugalde M, Busto N, Lozano HJ, Leal JM, Pérez B, Madariaga G, Insausti M, Lezama L, Sanz R, Gómez-Sainz LM, García B, García-Tojal J (2016) Selectivity of a thiosemicarbazonatocopper(II) complex towards duplex RNA. Relevant noncovalent interactions both in solid state and solution. Dalton Trans 45:18704-18718. https://doi.org/10.1039/C6DT02907H

97. Ruiz R, García B, García-Tojal J, Busto N, Ibeas S, Leal JM, Martins C, Gaspar J, Borras J, Gil-García R, González-Álvarez M (2010) Biological assays and noncovalent interactions of pyridine-2-carbaldehyde thiosemicarbazonecopper(II) drugs with [poly(dA–dT)]2, [poly(dG–dC)]2, and calf thymus DNA. J Biol Inorg Chem 15:515-532. https://doi.org/10.1007/s00775-009-0620-7

98. Saswati, Chakraborty A, Dash SP, Panda AK, Acharyya R, Biswas A, Mukhopadhyay S, Crochet A, Patil YP, Nethaji M, Dinda R (2015) Synthesis, X-ray structure and in vitro cytotoxicity studies of Cu(I/II) complexes of thiosemicarbazone: special emphasis on their interactions with DNA. Dalton Trans 44:6140-6157. https://doi.org/10.1039/C4DT03764B

99. Muralisankar M, Bhuvanesh NSP, Sreekanth A (2016) Synthesis, X-ray crystal structure, DNA/protein binding and DNA cleavage studies of novel copper(II) complexes of N-substituted isatin thiosemicarbazone ligands. New J Chem 40:2661-2679. https://doi.org/10.1039/C5NJ02806J

100. Lessa JA, Guerra JC, Miranda LF, Romeiro CFD, Da Silva JG, Mendes IC, Speziali NL, Souza-Fagundes EM, Beraldo H (2011) Gold(I) complexes with thiosemicarbazones: cytotoxicity against human tumor cell lines and inhibition of thioredoxin reductase activity. J Inorg Biochem 105:1729-1739. https://doi.org/10.1016/j.jinorgbio.2011.09.008

101. Myers JM, Cheng Q, Antholine WE, Kalyanaraman B, Filipovska A, Arnér ESJ, Myers CR (2013) Redox activation of Fe(III)–thiosemicarbazones and Fe(III)–bleomycin by thioredoxin reductase: specificity of enzymatic redox centers and analysis of reactive species formation by ESR spin trapping. Free Radic Biol Med 60:183-194. https://doi.org/10.1016/j.freeradbiomed.2013.02.016

102. Leigh M, Raines DJ, Castillo CE, Duhme-Klair AK (2011) Inhibition of xanthine oxidase by thiosemicarbazones, hydrazones and dithiocarbazates derived from hydroxy-substituted benzaldehydes. ChemMedChem 6:1107-1118. https://doi.org/10.1002/cmdc.201100054

103. Kaska WC, Carrano C, Michalowski J, Jackson J, Levinson W (1978) Inhibition of the RNA dependent DNA polymerase and the malignant transforming ability of Rous sarcoma virus by thiosemicarbazone-transition metal complexes. Bioinorg Chem 3:245-254. https://doi.org/10.1016/S0006-3061(00)80198-2

104. Bacher F, Enyedy EA, Nagy NV, Rockenbauer A, Bognár GM, Trondl R, Novak MS, Klapproth E, Kiss T, Arion VB (2013) Copper(II) complexes with highly water-soluble L- and D-proline–thiosemicarbazone conjugates as potential inhibitors of topoisomerase IIα. Inorg Chem 52:8895-8908. https://doi.org/10.1021/ic401079w

105. Bisceglie F, Pinelli S, Alinovi R, Goldoni M, Mutti A, Camerini A, Piola L, Tarasconi P, Pelosi G (2014) Cinnamaldehyde and cuminaldehyde thiosemicarbazones and their copper(II) and nickel(II) complexes: a study to understand their biological activity. J Inorg Biochem 140:111-125. https://doi.org/10.1016/j.jinorgbio.2014.07.014

106. Bisceglie F, Musiari A, Pinelli S, Alinovi R, Menozzi I, Polverini E, Tarasconi P, Tavone M, Pelosi G (2015) Quinoline-2-carboxaldehyde thiosemicarbazones and their Cu(II) and Ni(II) complexes as topoisomerase IIa inhibitors. J Inorg Biochem 152:10-19. https://doi.org/10.1016/j.jinorgbio.2015.08.008

107. Djoko KY, Paterson BM, Donnelly PS, McEwan AG (2014) Antimicrobial effects of copper(II) bis(thiosemicarbazonato) complexes provide new insight into their biochemical mode of action. Metallomics 6:854-863. https://doi.org/10.1039/c3mt00348e.

108. Moore EC, Zedeck MS, Agrawal KC, Sartorelli AC (1970) Inhibition of ribonucleoside diphosphate reductase by 1-formylisoquinoline thiosemicarbazone and related compounds. Biochemistry 9:4492-4498. https://doi.org/10.1021/bi00825a005

109. Moore EC, Booth BA, Sartorelli AC (1971) Inhibition of deoxyribonucleotide synthesis by pyridine carboxaldehyde thiosemicarbazones. Cancer Res 31:235-238.

111. Thelander L, Gräslund A (1983) Mechanism of inhibition of mammalian ribonucleotide reductase by the iron chelate of 1-formylisoquinoline thiosemicarbazone. Destruction of the tyrosine free radical of the enzyme in an oxygen-requiring reaction. J Biol Chem 258:4063-4066. https://doi.org/10.1016/S0021-9258(18)32582-1

112. Shao J, Zhou B, Di Bilio AJ, Zhu L, Wang T, Qi C, Shih J, Yen Y (2006) A Ferrous-Triapine[®] complex mediates formation of reactive oxygen species that inactivate human ribonucleotide reductase. Mol Cancer Ther 5:586-592. https://doi.org/10.1158/1535-7163.MCT-05-0384

113. Popović-Bijelić A, Kowol CR, Lind MES, Luo J, Himo F, Enyedy ÉA, Arion VB, Gräslund A (2011) Ribonucleotide reductase inhibition by metal complexes of Triapine[®] (3-aminopyridine-2-carboxaldehyde thiosemicarbazone): a combined experimental and theoretical study. J Inorg Biochem 105:1422-1431. https://doi.org/10.1016/j.jinorgbio.2011.07.003

114. Akladios FN, Andrew SD, Parkinson CJ (2015) Selective induction of oxidative stress in cancer cells via synergistic combinations of agents targeting redox homeostasis. Bioorganic Med Chem 23:3097-3104. https://doi.org/10.1016/j.bmc.2015.05.006

115. Kalinowski DS, Stefani C, Toyokuni S, Ganz T, Anderson GJ, Subramaniam NV, Trinder D, Olynyk JK, Chua A, Jansson PJ, Sahni S, Lane DJR, Merlot AM, Kovacevic Z, Huang MLH, Lee CS, Richardson DR (2016) Redox cycling metals: pedaling their roles in metabolism and their use in the development of novel therapeutics. Biochim Biophys Acta-Mol Cell Res 1863:727-748. https://doi.org/10.1016/j.bbamcr.2016.01.026

116. Akladios FN, Andrew SD, Parkinson CJ (2016) Increased generation of intracellular reactive oxygen species initiates selective cytotoxicity against the MCF-7 cell line resultant from redox active combination therapy using copper–thiosemicarbazone complexes. J Biol Inorg Chem 21:407-419. https://doi.org/10.1007/s00775-016-1350-2

117. Whitnall M, Howard J, Ponka P, Richardson DR (2006) A class of iron chelators with a wide spectrum of potent antitumor activity that overcomes resistance to chemotherapeutics. Proc Natl Acad Sci 103:14901-14906. https://doi.org/10.1073/pnas.0604979103

118. Kolesar JM, Sachidanandam K, Schelman WR, Eickhoff J, Holen KD, Traynor AM, Alberti DB, Thomas JP, Chitambar CR, Wilding G, Antholine WE (2011) Cytotoxic evaluation of 3-aminopyridine-2-carboxaldehyde thiosemicarbazone in peripheral blood lymphocytes of patients with refractory solid tumors using electron paramagnetic resonance. Exp Ther Med 2:119-123. https://doi.org/10.3892/etm.2010.165

119. Lovejoy DB, Jansson PJ, Brunk UT, Wong J, Ponka P, Richardson DR (2011) Antitumor activity of metalchelating compound Dp44mT is mediated by formation of a redox-active copper complex that accumulates in lysosomes. Cancer Res 71:5871-5880. https://doi.org/10.1158/0008-5472.CAN-11-1218

120. Ishiguro K, Lin ZP, Penketh PG, Shyam K, Zhu R, Baumann RP, Zhu YL, Sartorelli AC, Rutherford TJ, Ratner ES (2014) Distinct mechanisms of cell-kill by triapine, and its terminally dimethylated derivative Dp44mT due to a loss or gain of activity of their copper(II) complexes. Biochem Pharmacol 91:312-322. https://doi.org/10.1016/j.bcp.2014.08.006

121. Haeili M, Moore C, Davis CJC, Cochran JB, Shah S, Shrestha TB, Zhang Y, Bossmann SH, Benjamin WH, Kutsch O, Wolschendorf F (2014) Copper complexation screen reveals compounds with potent antibiotic properties against methicillin-resistant staphylococcus aureus. Antimicrob Agents Ch 58:3727-3736. https://doi.org/10.1128/aac.02316-13

122. Stacy AE, Palanimuthu D, Bernhardt PV, Kalinowski DS, Jansson PJ, Richardson DR (2016) Zinc(II)thiosemicarbazone complexes are localized to the lysosomal compartment where they transmetallate with copper ions to induce cytotoxicity. J Med Chem 59:4965-4984. https://doi.org/10.1021/acs.jmedchem.6b00238

123. Jansson PJ, Sharpe PC, Bernhardt PV, Richardson DR (2010) Novel thiosemicarbazones of the ApT and DpT series and their copper complexes: identification of pronounced redox activity and characterization of their antitumor activity. J Med Chem 53:5759-5769.https://doi.org/10.1021/jm100561b

124. French FA, Blanz EJ, Shaddix SC, Brockman RW (1974) α-(N)-Formylheteroaromatic thiosemicarbazones. inhibition of tumor-derived ribonucleoside diphosphate reductase and correlation with in vivo antitumor activity. J Med Chem 17:172-181. https://doi.org/10.1021/jm00248a006

125. Lovejoy DL, Richardson DR (2002) Novel "hybrid" iron chelators derived from aroylhydrazones and thiosemicarbazones demonstrate selective antiproliferative activity against tumor cells. Blood 100:666-675. https://doi.org/10.1182/blood.V100.2.666

126. Bacher F, Dömötör O, Kaltenbrunner M, Mojović M, Popović-Bijelić A, Gräslund A, Ozarowski A, Filipovico L, Radulovićo S, Enyedy ÉA, Arion VA (2014) Effects of terminal dimethylation and metal coordination of proline-2-formylpyridine thiosemicarbazone hybrids on lipophilicity, antiproliferative activity, and hR2 RNR inhibition. Inorg Chem 53:12595-12609. https://doi.org/10.1021/ic502239u

127. Stefani C, Punnia-Moorthy G, Lovejoy DB, Jansson PJ, Kalinowski DS, Sharpe PC, Bernhardt PV, Richarrdson DR (2011) Halogenated 2'-benzoylpyridine thiosemicarbazone (XBpT) chelators with potent and selective antineoplastic activity: relationship to intracellular redox activity. J Med Chem 54:6936-6948. https://doi.org/10.1021/jm200924c

128. Wang L, He W, Yu Z (2013) Transition-metal mediated carbon–sulfur bond activation and transformations. Chem Soc Rev 42:599-621. https://doi.org/10.1039/C2CS35323G

Journal of Biological Inorganic Chemistry

Ren P, Pike SD, Pernik I, Weller AS, Willis MC (2015) Rh–POP pincer xantphos complexes for C–S and C–H activation. Implications for carbothiolation catalysis. Organometallics 34:711-723. https://doi.org/10.1021/om500984y
 Munjanja L, Brennessel WW, Jones WD (2015) Room-temperature carbon–sulfur bond activation by a reactive (dippe)Pd fragment. Organometallics 34:1716-1724. https://doi.org/10.1021/acs.organomet.5b00194

131. Li Y, Rauchfuss TB (2016) Synthesis of diiron(I) dithiolato carbonyl complexes. Chem Rev 116:7043-7077. https://doi.org/10.1021/acs.chemrev.5b00669

132. Kumar S, Guyon F, Knorr M, Labat S, Miqueu K, Golz C, Strohmann C (2017) Experimental and theoretical studies on the mechanism of the C–S bond activation of Pd^{II} thiolate/thioether complexes. Organometallics 36:1303-1321. https://doi.org/10.1021/acs.organomet.7b00039

133. Lou J, Wang Q, Wu P, Wang H, Zhou YG, Yu Z (2020) Transition-metal mediated carbon–sulfur bond activation and transformations: an update. Chem Soc Rev 49:4307-4359. https://doi.org/10.1039/C9CS00837C

134. Gómez-Saiz P, Gil-García R, Maestro MA, Pizarro JL, Arriortua MI, Lezama L, Rojo T, García Tojal J (2005) Unexpected behaviour of pyridine-2-carbaldehyde thiosemicarbazonatocopper(II) entities in aqueous basic medium-Partial transformation of thioamide into nitrile. Eur J Inorg Chem 3409-3413. https://doi.org/10.1002/ejic.200500326

135. García-Tojal J, Urtiaga MK, Cortés R, Lezama L, Arriortua MI, Rojo T (1994) Synthesis, structure, spectroscopic and magnetic properties of two copper(II) dimers containing pyridine-2-carbaldehyde thiosemicarbazonate (L), [{CuL(X)}₂] (X = Cl or Br). J Chem Soc Dalton Trans 2233-2238. https://doi.org/10.1039/DT9940002233

136. García-Tojal J, Gil-García R, Fouz VI, Madariaga G, Lezama L, Galletero MS, Borrás J, Nollmann FI, García-Girón C, Alcaraz R, Cavia-Saiz M, Muñiz P, Palacios Ò, Samper KG, Rojo T (2018) Revisiting the thiosemicarbazonecopper(II) reaction with glutathione. Activity against colorectal carcinoma cell lines. J Inorg Biochem 180:69-79. https://doi.org/10.1016/j.jinorgbio.2017.12.005

137. Gil-García R, Fraile R, Donnadieu B, Madariaga G, Januskaitis V, Rovira J, González L, Borrás J, Arnáiz FJ, García-Tojal J (2013) Desulfurization processes of thiosemicarbazonecopper(II) derivatives in acidic and basic aqueous media. New J Chem 37:3568-3580. https://doi.org/10.1039/C3NJ00321C

138. Ainscough EW, Brodie AM, Denny WA, Finlay GJ, Ranford JD (1998) Nitrogen, sulfur and oxygen donor adducts with copper(II) complexes of antitumor 2-formylpyridinethiosemicarbazone analogs: physicochemical and cytotoxic studies. J Inorg Biochem 70:175-185. https://doi.org/10.1016/S0162-0134(98)10011-9

139. Gil–García R, Madariaga G, Jiménez-Pérez A, Herrán-Torres, I, Gago-González A, Ugalde M, Januskaitis V, Barrera-García J, Insausti M, Galletero MS, Borrás J, Cuevas JV, Pedrido R, Gómez-Saiz P, Lezama L, García-Tojal J (2023) Perchlorate-induced structural diversity in thiosemicarbazone-copper(II) complexes provides insights to understand the reactivity in acid and basic media. CrystEngComm 25:2213-2226. https://doi.org/10.1039/D3CE00119A

140. Wang YT, Fang Y, Zhao M, Li MX, Ji YM, Han QX (2017) Cu(II), Ga(III) and In(III) complexes of 2acetylpyridine N(4)-phenylthiosemicarbazone: synthesis, spectral characterization and biological activities. MedChemComm 8:2125-2132. https://doi.org/10.1039/C7MD00415J

141. Al-Eisawi Z, Stefani C, Jansson PJ, Arvind A, Sharpe PC, Basha MT, Iskander GM, Kumar N, Kovacevic Z, Lane DJR, Sahni S, Bernhardt PV, Richardson DR, Kalinowski DS (2016) Novel mechanism of cytotoxicity for the selective selenosemicarbazone, 2-acetylpyridine 4,4-dimethyl-3-selenosemicarbazone (Ap44mSe): lysosomal membrane permeabilization. J Med Chem 59:294-312. https://doi.org/10.1021/acs.jmedchem.5b01399

142. Enyedy ÉA, Nagy NV., Zsigó É, Kowol CR, Arion VB, Keppler BK, Kiss T (2010) Comparative solution equilibrium study of the interactions of copper(II), iron(II) and zinc(II) with triapine[®] (3-aminopyridine-2-carbaldehyde thiosemicarbazone) and related ligands. Eur J Inorg Chem 2010:1717-1728. https://doi.org/10.1002/ejic.200901174

143. Dömötör O, May NV., Pelivan K, Kiss T, Keppler BK, Kowol CR, Enyedy EA (2018) A comparative study of α -N-pyridyl thiosemicarbazones: spectroscopic properties, solution stability and copper(II) complexation. Inorg Chim Acta 472:264-275. https://doi.org/10.1016/j.ica.2017.07.001

144. Gil-García R, Gómez-Saiz P, Díez-Gómez V, Madariaga G, Insausti M, Lezama L, Cuevas JV, García-Tojal J (2014) Thiosemicarbazonecopper(II) compounds with halide/hexafluorosilicate anions: structure, water clusters, non-covalent interactions and magnetism. Polyhedron 81:675-686. https://doi.org/10.1016/j.poly.2014.07.032

145. Abras A, Beraldo H, Fantini EO, Borges RHU, Da Rocha MA, Tosi L (1990) Spectroscopic studies of metal complexes containing π -delocalized sulfur ligands. Mössbauer and kinetic studies of iron(II) and iron(III) complexes of the antitumor agent 2-formylpyridine thiosemicarbazone. Inorg Chim Acta 172:113-117. https://doi.org/10.1016/S0020-1693(00)80459-4

146. Borges RHU, Paniago E, Beraldo H (1997) Equilibrium and kinetic studies of iron(II) and iron(III) complexes of some α (N)-heterocyclic thiosemicarbazones. Reduction of the iron(III) complexes of 2-formylpyridine thiosemicarbazone and 2-acetylpyridine thiosemicarbazone by cellular thiol-like reducing agents. J Inorg Biochem 65:267-275. https://doi.org/10.1016/S0162-0134(96)00142-0

147. García-Tojal J, Donnadieu B, Costes JP, Serra JL, Lezama L, Rojo T (2002) Spectroscopic properties of iron thiosemicarbazone compounds. Structure of $[Fe(C_7H_7N_4S)_2] \cdot 1.25H_2O$. Inorg Chim Acta 333:132-137. https://doi.org/10.1016/S0020-1693(02)00802-2

148. Fukumoto K, Sakai A, Hayasaka K, Nakazawa H (2013) Desulfurization and H-migration of secondary thioamides catalyzed by an iron complex to yield imines and their reaction mechanism. Organometallics 32:2889-2892. https://doi.org/10.1021/om400304v

149. Mutoh Y, Sakigawara M, Niiyama I, Saito S, Ishii Y (2014) Synthesis of rhodium–primary thioamide complexes and their desulfurization leading to rhodium sulfido cubane-type clusters and nitriles. Organometallics 33:5414-5422. https://doi.org/10.1021/om500714c

150. Guo W, Liu G, Deng L, Mei W, Zou X, Zhong Y, Zhou X, Fan X, Zheng L (2021) Metal- and oxidant-free green three-component desulfurization and deamination condensation approach to fully substituted 1 H-1,2,4-triazol-3-amines and their photophysical properties. J Org Chem 86:17986-18003. https://doi.org/10.1021/acs.joc.1c02313

151. Gómez-Saiz P, García-Tojal J, Diez-Gómez V, Gil-García R, Pizarro JL, Arriortua MI, Rojo T (2005) Indirect evidences of desulfurization of a thiosemicarbazonecopper(II) system in aqueous basic medium. Inorg Chem Commun 8:259-262. https://doi.org/10.1016/j.inoche.2004.12.016

152. Castiñeiras A, Garcia-Santos I (2008) Desulfuration and cyclization of (Z)-2-[amino(pyridine-2-yl)methylene]hydrazonecarbothioamide in the presence of manganese(II). Zeitschrift fur Anorg und Allg Chemie 634:2907-2916. https://doi.org/10.1002/zaac.200800326

153. Van Poppel LH, Groy TL, Caudle MT (2004) Carbon-Sulfur bond cleavage in bis(N-alkyldithiocarbamato)cadmium(II) complexes: heterolytic desulfurization coupled to topochemical proton transfer. Inorg Chem 43:3180-3188. https://doi.org/10.1021/ic035135v

154. Al-Mutairi AA, Al-Alshaikh MA, Al-Omary FAM, Hassan HM, El-Mahdy AM, El-Emam AA (2019) Synthesis, antimicrobial, and anti-proliferative activities of novel 4-(adamantan-1-yl)-1-arylidene-3- thiosemicarbazides, 4- arylmethyl N-(adamantan-1-yl) piperidine-1-carbothioimidates, and related derivatives. Molecules 24:4308. https://doi.org/10.3390/molecules24234308

155. Jeong H, Kang Y, Kim J, Kim BK, Hong S (2019) Factors that determine thione(thiol)-disulfide interconversion in a bis(thiosemicarbazone) copper(II) complex. RSC Adv 9:9049-9052. https://doi.org/10.1039/c9ra01115c

156. Pedrido R, Romero MJ, Bermejo MR, González-Noya AM, García-Lema I, Zaragoza G (2008) Metal-catalysed oxidation processes in thiosemicarbazones: new complexes with the ligand N-{2-([4-N-ethylthiosemicarbazone]-methyl)phenyl}-p- toluenesulfonamide. Chem Eur J 14:500-512. https://doi.org/10.1002/chem.200700867

157. Pedrido R, Romero MJ, Bermejo MR, Martínez-Calvo M, González-Noya AM, Zaragoza G (2009) Coordinative trends of a tridentate thiosemicarbazone ligand: synthesis, characterization, luminescence studies and desulfurization processes. Dalton Trans 39:8329-8340. https://doi.org/10.1039/b908782f

158. Da Rosa Justim J, Correa Bohs LM, Barreto Martins B, Tribuzy Bandeira KC, Lopes de Melo AP, Carratu Gervini V, Bresolin L, Godoi M, de Menezes Peixoto CR (2021) Electrochemical characterization of isatin-thiosemicarbazone derivatives. J Chem Sci 133:124. https://doi.org/10.1007/s12039-021-01970-x

159. Pelivan K, Frensemeier LM, Karst U, et al (2018) Comparison of metabolic pathways of different α -N-heterocyclic thiosemicarbazones. Anal Bioanal Chem 410:2343-2361. https://doi.org/10.1007/s00216-018-0889-x

160. Gómez-Saiz P, García-Tojal J, Maestro MA, Arnaiz FJ, Rojo T (2002) Evidence of desulfurization in the oxidative cyclization of thiosemicarbazones. Conversion to 1,3,4-oxadiazole derivatives. Inorg Chem 41:1345-1347. https://doi.org/10.1021/ic015625s

161. Gómez-Saiz P, García-Tojal J, Maestro MA, Mahía J, Arnaiz FJ, Lezama L, Rojo T (2003) New 1,3,4oxadiazolecopper(II) derivatives obtained from thiosemicarbazone complexes. Eur J Inorg Chem 2003:2639-2650. https://doi.org/10.1002/ejic.200200689

162. Hiremath SP, Goudar NN, Purohit MG (1981) Synthesis of substituted 1',3',4'-oxadiazolyl-indoles, thiadiazolyl-indoles and triazolyl-indoles. Indian J Chem Sect B 20:388-390.

163. Hiremath SP, Biradar JS, Kudari SM (1984) Synthesis of substituted oxadiazoles, thiadiazoles and triazoles and evaluation of their biological activity. J Indian Chem Soc 61:74-76. https://doi.org/10.5281/zenodo.6325527

164. Fernandes PS, Sonar TM (1986) Synthesis and biological activity of heterocyclic derivatives derived from ethyl-2-hydroxyquinoxaline-3-carboxylate. J Indian Chem Soc 63:427-429. https://doi.org/10.5281/zenodo.6273852

165. Hiremath SP, Sonar VN, Sekhar KR, Purohit MG (1989) Synthesis of substituted oxadiazolylindoles, thiadiazolylindoles and indolylthiazolidinones. Indian J Chem Sect B 28B:626-630.

Raman K, Singh HK, Salzman SK, Parmar SS (1993) Substituted thiosemicarbazides and corresponding cyclized
 1,3,4-oxadiazoles and their antiinflammatory activity. J Pharm Sci 82:167-169. https://doi.org/10.1002/jps.2600820210
 Kelarev VI, Karakhanov RA, Gassanov SS, Morozova GV, Kuatbekova KP (1993) Synthesis of 1,3,4-oxa(thia)diazole and 1,2,4-triazolederivatives containing 3-indolylmethyl radicals. Russ J Org Chem 29:388-395.

168. Albar HA, Makki MSI, Faidallah HM (1996) Synthesis of heterocyclic compounds from delta-unsaturated 1,3diketo-esters. Indian J Chem Sect B 35:23-29.

169. Vashi BS, Mehta DS, Shah VH (1996) Synthesis of 2,5-disubstituted-1,3,4-oxadiazole, 1,5-disubstituted-2mercapto-1,3,4-triazole and 2,5-disubstituted-1,3,4-thiadiazole derivatives as potential antimicrobial agents. Indian J Chem Sect B 35:111-115. https://doi.org/10.1002/chin.199617050

170. Rao GR, Mogilaiah K, Sreenivasulu B (1996) Synthesis and antimicrobial activity of 1',2',4'-triazolyl/1',3',4'-thiadiazolyl/1',3',4'-oxadiazolyl-1,8-naphthyridines and related compounds. Indian J Chem Sect B 35:339-344.

171. Omar FA, Mahfouz NM, Rahman MA (1996) Design, synthesis and antiinflammatory activity of some 1,3,4oxadiazole derivatives. Eur J Med Chem 31:819-825. https://doi.org/10.1016/0223-5234(96)83976-6

172. Tripathi P, Pal A, Jancik V, Pandey AK, Singh J, Singh NK (2007) Metal-assisted transformation of Nbenzoyldithiocarbazate to 5-phenyl-1,3,4-oxadiazole-2-thiol in the presence of ethylenediamine, and its first row transition metalcomplexes. Polyhedron 26:2597-2602. https://doi.org/10.1016/j.poly.2006.12.046

173. Hassan AA, Mourad AFE, Abou-Zied AH (2007) Reaction of 1-acylthiosemicarbazides with ethenetetracarbonitrile. J Heterocyclic Chem 44:1171-1176. https://doi.org/10.1002/jhet.5570440532

174. Hassan AA, Mourad AE, Abuo-Zied AH (2007) Benzo- and naphthoimidazoxadiazolediene, naphthobisthiazole as well as naphthothiazine derivatives from 1-acylthiosemicarbazides. Arkivoc 222-235. https://doi.org/10.3998/ark.5550190.0008.124 175. Mekuskiene G, Tumkevicius S, Vainilavicius P (2002) 5-(4,6-diphenyl-2-pyrimidinyl)-1,3,4-oxa(thia)diazoles and 1,2,4-triazoles. J Chem Res 213-215. https://doi.org/10.3184/030823402103171898

176. Dolman SJ, Gosselin F, O'Shea PD, Davies IW (2006) Superior reactivity of thiosemicarbazides in the synthesis of 2-amino-1,3,4-oxadiazoles. J Org Chem 71:9548-9551. https://doi.org/10.1021/j00618730

177. Singh NK, Bharty MK, Dulare R, Butcher RJ (2009) Synthesis and X-ray crystallographic studies of Ni(II) and Cu(II) complexes of [5-(4-pyridyl)-1,3,4] oxadiazole-2-thione/thiol formed by transformation of N-(pyridine-4-carbonyl)-hydrazine carbodithioate in the presence of ethylenediamine. Polyhedron 28:2443-2449. https://doi.org/10.1016/j.poly.2009.04.030

178. Li Z, Zhu A, Yang J (2012) One-pot three-component mild synthesis of 2-aryl-3-(9-alkylcarbazol-3-yl)thiazolin4-ones. J Heterocycl Chem 49:1458-1461.https://doi.org/10.1002/jhet.1047

179. Bharti A, Bharty MK, Kashyap S, Singh UP, Butcher RJ, Singh NK (2013) Hg(II) complexes of 4-phenyl-5-(3-pyridyl)-1,2,4-triazole-3-thione and 5-(4-pyridyl)-1,3,4-oxadiazole-2-thione and a Ni(II) complex of 5-(thiophen-2-yl)-1,3,4-oxadiazole-2-thione: synthesis and X-ray structural studies. Polyhedron 50:582-591. https://doi.org/10.1016/j.poly.2012.11.043

180. Yang Z, She M, Yin B, Hao L, Obst M, Li J (2015) Solvent-dependent turn-on probe for dual monitoring of Ag⁺ and Zn²⁺ in living biological samples. Anal Chim Acta 868:53-59. https://doi.org/10.1016/j.aca.2015.01.052

181. Bao W, Chen C, Yi N, Jiang J, Zeng Z, Deng W, Peng Z, Xiang J (2017) Synthesis of 2-amino-1,3,4-oxadiazoles through elemental sulfur promoted cyclization of hydrazides with isocyanides. Chin J Chem 35:1611-1618. https://doi.org/10.1002/cjoc.201700188

182. Golmohammadi F, Balalaie S, Hamdan F, Maghari S (2018) Efficient synthesis of novel conjugated 1,3,4oxadiazole-peptides. New J Chem 42:4344-4351. https://doi.org/10.1039/c7nj04720g

183. Abu-Hashem AA (2021) Synthesis and antimicrobial activity of new 1,2,4-triazole, 1,3,4-oxadiazole, 1,3,4-thiadiazole, thiopyrane, thiazolidinone, and azepine derivatives. J Heterocycl Chem 58:74-92. https://doi.org/10.1002/jhet.4149

184. Landquist JK (1970) Oxidative cyclisation of ketone thiosemicarbazones. Part I. 4-methyl- and 4-aryl-thiosemicarbazones. J Chem Soc C 63-66. https://doi.org/10.1039/J39700000063

185. Xiao S, Lee W, Chen F, Zavalij PY, Gutierrez O, Davis JT (2020) Oxidation of 8-thioguanosine gives redoxresponsive hydrogels and reveals intermediates in a desulfurization pathway. Chem Commun 56:6981-6984. https://doi.org/10.1039/D0CC02926B

186. Baird IR, Cameron BR, Skerlj RT (2003) Unique chemistry of amino acid dithiocarbamates with Ru(III) bis-βdiketonates. Inorganica Chim Acta 353:107-118. https://doi.org/10.1016/S0020-1693(03)00233-0

187. Ng S, Ziller JW, Farmer PJ (2004) Multiple pathways for the oxygenation of a ruthenium(II) dithiocarbamate complex: S-oxygenation and S-extrusion. Inorg Chem 43:8301-8309. https://doi.org/10.1021/ic048661a

188. Fernández-Fariña S, González-Barcia LM, Romero MJ, García-Tojal J, Maneiro, M, Seco JM, Zaragoza G, Martínez-Calvo M, González-Noya AM, Pedrido R (2022) Conversion of a double-tetranuclear cluster silver helicate into a dihelicate via a rare desulfurization process. Inorg Chem Front 9:531-536. https://doi.org/10.1039/d1qi01308d

189. Alsop L, Cowley AR, Dilworth JR, Donnelly PS, Peach JM, Rider JT (2005) Investigations into some aryl substituted bis(thiosemicarbazones) and their copper complexes. Inorganica Chim Acta 358:2770-2780. https://doi.org/10.1016/j.ica.2005.03.027

190 Gooneh-Farahani S, Anbia M (2023) A review of advanced methods for ultra-deep desulfurization under mild conditions and the absence of hydrogen. J Environ Chem Eng 11:108997. https://doi.org/10.1016/j.jece.2022.108997 191. Aly AA, Hassan AA, Brown AB, Ibrahim MAA, AbdAl-Latif ESSM (2017) Azines from one-pot reaction of thiosemicarbazones. J Sulphur Chem 38:11-17. https://doi.org/10.1080/17415993.2016.1210146

1	
2	
3	
4	192. Kassim K, Hamali MA, Yamin B (2019) A New alternative synthesis of salicylaldazine via microwave irradiation
5	method. J Chem 2019:9546373. https://doi.org/10.1155/2019/9546373
7	193. Qiu W, Shi S, Li R, Lin X, Rao L, Sun Z (2021) A mild, general, metal-free method for desulfurization of thiols
8	and disulfides induced by visible-light. Chinese J Chem 39:1255-1258. https://doi.org/10.1002/cjoc.202000607
9	194. Maji M, Chatterjee M, Ghosh S, Chattopadhyay SK, Wu BM, Mak TCW (1999) Chemistry of ruthenium(II)
10	complexes of the tridentate NNS donor methyl 2-pyridyl ketone 4-(4-tolyl)thiosemicarbazone. Isolation and structural
11	characterisation of a novel ruthenium(II) complex containing a co-ordinated imine of an alpha N haterocyclic katone
12	characterisation of a nover runnehum (1) complex containing a co-ordinated mine of an appla-tv neterocyclic ketone.
13	J Chem Soc, Daiton Trans 135-140. https://doi.org/10.1039/A8063411
15	195. Hassan AA, El-Sheref EM (2010) Chemistry and heterocyclization of dithiobiurea and thioureidoalkylthiourea.
16	J Heterocycl Chem 47:764-784. https://doi.org/10.1002/jhet.406
17	196. Da S. Maia PI, Nguyen HH, Ponader D, Hagenbach A, Bergemann S, Gust R, Deflon VM, Abram U (2012)
18	Neutral gold complexes with tridentate SNS thiosemicarbazide ligands. Inorg Chem 51:1604-1613.
19	https://doi.org/10.1021/ic201905t
20	197 Gil-García R. Zichner R. Díez-Gómez V. Donnadieu B. Madariaga G. Insausti M. Lezama L. Vitoria P. Pedrosa
21	MR García-Toial I (2010) Polyovometallate thiosemicarbazone hybrid compounds. Fur I Inorg Chem 2010:1513-
23	4525 https://doi.org/10.1002/sijo.201000484
24	
25	198. Cowley AR, Dilworth JR, Donnelly PS, Woollard-Shore J (2003) Synthesis and characterisation of new
26	homoleptic rhenium thiosemicarbazone complexes. J Chem Soc Dalton Trans 3:748-754.
27	https://doi.org/10.1039/B210540N
20	199. Matesanz AI, Pastor C, Souza P (2007) Synthesis and structural characterization of a disulphide-bridged
30	tetranuclear palladium(II) complex derived from 3,5-diacetyl 1,2,4-triazole bis(4-ethylthiosemicarbazone). Inorg Chem
31	Commun 10:97-100. https://doi.org/10.1016/j.inoche.2006.09.016
32	200. Zhao Y. Lin Z. He C. Wu H. Duan C (2006) A "turn-on" fluorescent sensor for selective Hg(II) detection in
33	aqueous media based on metal-induced dye formation Inorg Chem 45:10013-10015
34 35	https://doi.org/10.1021/io061067b
36	$\frac{1}{1000} = \frac{1}{1000} = \frac{1}{10000000000000000000000000000000000$
37	201. Zhang P, Shi B, Zhang Y, Lin Q, Yao H, You XM, Wei TB (2013) A selective fluorogenic chemodosimeter for
38	Hg^{2+} based on the dimerization of desulfurized product. Tetrahedron 69:10292-10298.
39	https://doi.org/10.1016/j.tet.2013.10.024
40	202. Hwang KS, Park KY, Kim D Bin, Chang SK (2017) Fluorescence sensing of Ag ⁺ ions by desulfurization of an
41 42	acetylthiourea derivative of 2-(2-hydroxyphenyl)benzothiazole. Dyes Pigm 147:413-419.
43	https://doi.org/10.1016/j.dyepig.2017.08.041
44	203. Bulak E, Dogan I, Varnali T, Schwederski B, Gunal SE, Lönnecke P, Bubrin M, Kaim W (2021) An Acyclic
45	Diaminocarbene Complex of Platinum Formed by Desulfurization of 1 3-Bis(3-methylpyridin-2-yl)thiourea Eur I
46	$\frac{1}{2} = \frac{1}{2} = \frac{1}$
47	204 Singh & Chaterradi I. Dhattacharra & (2012) Studies of surthering structural features of Cu(1) this share 2
48 70	204. Singn S, Chaturvedi J, Bhattacharya S (2012) Studies of synthesis, structural features of Cu(1) thiophene-2-
50	thiocarboxylates and unprecedented desulfurization of Cu(II) thiocarboxylate complexes. Dalton Trans 41:424-431.
51	https://doi.org/10.1039/C1DT10629E
52	205. Bigoli F, Deplano P, Mercuri L, Pellinghelli MA, Pintus G, Trogu EF (1999) Unusual desulfurization of a nickel
53	dithiolene by bis(2-diphenylphosphinophenyl)phenylphosphine (tp) to produce $Ni(tp)(R4btimdt)$ [R4btimdt = 5,5'-
54	bis(1,3-dialkyl-4-imidazolidine-2-thione-4-thiolate], the first complex of this class of ligands. Chem Commun
55 56	698:2093-2094. https://hdl.handle.net/11381/1458647
57	206. Chawla SK. Arora M. Nättinen K. Rissanen K (2006) Unique conner ion catalyzed hydrolytic cleavage of C-
58	N(2) hond of thiosemicarbazide Polyhedron 25:627-634 https://doi.org/10.1016/j.poly.2005.05.021
59	1(2) cond of unosemical outlide. For ynder on 25.027-054. https://doi.org/10.1010/j.p01y.2005.05.021
60	

207. Voitekhovich SV, Lyakhov AS, Ivashkevich LS, Ivashkevich OA (2020) Copper-assisted desulfurization of 1-R-tetrazole-5-thiols under complexation. Inorg Chem Commun 114:107827-107830. https://doi.org/10.1016/j.inoche.2020.107827

208. Tardito S, Bussolati O, Maffini M, Tegoni M, Giannetto M, Dall'Asta V, Franchi-Gazzola R, Lanfranchi M, Pellinghelli MA, Mucchino C, Mori G, Marchiò L (2007) Thioamido coordination in a thioxo-1,2,4-triazole copper(II) complex enhances nonapoptotic programmed cell death associated with copper accumulation and oxidative stress in human cancer cells. J Med Chem 50:1916-1924. https://doi.org/10.1021/jm061174f

209. Madshus IH (1988) Regulation of intracellular pH in eukaryotic cells. Biochem J. 250:1-8. https://doi.org/10.1042/bj2500001

210. Nunes P, Guido D, Demaurex N (2015) Measuring phagosome pH by ratiometric fluorescence microscopy. J Vis Exp e53402. https://doi.org/10.3791/53402

211. Asokan A, Cho MJ (2002) Exploitation of intracellular pH gradients in the cellular delivery of macromolecules. J Pharm Sci 91:903-913. https://doi.org/10.1002/jps.10095

212. Porcelli AM, Ghelli A, Zanna C, Pinton P, Rizzuto R, Rugolo M (2005) pH difference across the outer mitochondrial membrane measured with a green fluorescent protein mutant. Biochem Biophys Res Commun 326:799-804. https://doi.org/10.1016/j.bbrc.2004.11.105

213. Wilson AF, Simmons DH (1970) Organ and Whole Body Cell pH. Proc Soc Exp Biol Med 134:127-130. https://doi:10.3181/00379727-134-34743

214. Park HJ, Lim CS, Kim ES, Han HJ, Lee TH, Chun HJ, Cho BR (2012) Measurement of pH values in human tissues by two-photon microscopy. Angew Chem Int Ed 51:2673-2676. https://doi.org/10.1002/anie.201109052

215. Persi E, Duran-Frigola M, Damaghi M, Roush WR, Aloy P, Cleveland JL, Gillies RJ, Ruppin E (2018) Systems analysis of intracellular pH vulnerabilities for cancer therapy. Nat Commun 9:2997. https://doi.org/10.1038/s41467-018-05261-x

216. Radi R (2018) Oxygen radicals, nitric oxide, and peroxynitrite: redox pathways in molecular medicine. Proc Natl Acad Sci U S A 115:5839-5848. https://doi.org/10.1073/pnas.1804932115

217. Hu X, Dong D, Xia M, Yang Y, Wang J, Su J, Sun L, Yu H (2020) Oxidative stress and antioxidant capacity: development and prospects. New J Chem 44:11405-11419. https://doi.org/10.1039/D0NJ02041A

218. Di Meo S, Reed TT, Venditti P, Victor VM (2016) Role of ROS and RNS sources in physiological and pathological conditions. Oxid Med Cell Longev 2016:ID 1245049. https://doi.org/10.1155/2016/1245049

219. Zhang Y, Wong HS (2021) Are mitochondria the main contributor of reactive oxygen species in cells. J Exp Biol 224:jeb221606. https://doi.org/10.1242/jeb.221606

220. Neha K, Haider MR, Pathak A, Yar MS (2019) Medicinal prospects of antioxidants: a review. Eur J Med Chem 178:687-704. https://doi.org/10.1016/j.ejmech.2019.06.010

221. Pisoschi AM, Pop A, Iordache F, Stanca L, Predoi G, Serban AI (2021) Oxidative stress mitigation by antioxidants - An overview on their chemistry and influences on health status. Eur J Med Chem 209:112891. https://doi.org/10.1016/j.ejmech.2020.112891

222. Jungwirth U, Kowol CR, Keppler BK, Hartinger CG, Berger W, Heffeter P (2011) Anticancer activity of metal complexes: involvement of redox processes. Antioxidan Redox Signal 15:1085-1127. https://doi.org/10.1089/ars.2010.3663

223. Eteshola EOU, Haupt DA, Koos SI, Siemer LA, Morris DL (2020) The role of metal ion binding in the antioxidant mechanisms of reduced and oxidized glutathione in metal-mediated oxidative DNA damage. Metallomics 79-91. https://doi.org/10.1039/C9MT00231F

224. Ouyang Y, Peng Y, Li J, Holmgren A, Lu (2018) Modulation of thiol-dependent redox system by metal ions *via* thioredoxin and glutaredoxin systems. Metallomics 10:218-228. https://doi.org/10.1039/C7MT00327G

225. Arunachalam V, Tummanapelli AK, Vasudevan S (2019) The multiple dissociation constants of glutathione disulfide: interpreting experimental pH-titration curves with: Ab initio MD simulations. Phys Chem Chem Phys 21:9212-9217. https://doi.org/10.1039/C9CP00761J

226. Falcone E, Ritacca AG, Hager S, Schueffl H, Vileno B, Khoury YE, Hellwig P, Kowol CR, Heffeter P, Sicilia E, Faller P (2022) Copper-catalyzed glutathione oxidation is accelerated by the anticancer thiosemicarbazone Dp44mT and further boosted at lower pH. J Am Chem Soc 144:14758-14768. https://doi.org/10.1021/jacs.2c05355

227. Nutting LA, Weber EM, Tryon JL (1967) Metabolic removal of sulfur from 1-methylisatin 3-thiosemicarbazone.J Virol 1:650-651. https://doi.org/10.1128/jvi.1.3.650-651.1967

228. Creasey, WA, Agrawal KC, Capizzi RL, Stinson KK, Sartorelli AC (1972) Studies of the antineoplastic activity and metabolism of α-(N)-heterocyclic carboxaldehyde thiosemicarbazones in dogs and mice. Cancer Res 32:565-572.
229. DeConti RC, Toftness BR, Agrawal KC, Tomchick R, Mead JA, Bertino JR, Sartorelli AC, Creasey WA (1972) Clinical and pharmacological studies with 5-hydroxy-2-formylpyridine thiosemicarbazone. Cancer Res 32:1455-1462.
230. Pelivan K, Frensemeier L, Karst U, Koellensperger G, Bielec B, Hager S, Heffeter P, Kepplerae BK, Kowol CR (2017) Understanding the metabolism of the anticancer drug Triapine[®]: Electrochemical oxidation, microsomal incubation and: In vivo analysis using LC-HRMS. Analyst 142:3165-3176. https://doi.org/10.1039/C7AN00902J

231. Stariat J, Šesták V, Vávrová K, Nobilis M, Kollárová Z, Klimeš J, Kalinowski DS, Richardson DR, Kovaříková P (2012) LC-MS/MS identification of the principal in vitro and in vivo phase I metabolites of the novel thiosemicarbazone anti-cancer drug, Bp4eT. Anal Bioanal Chem 403:309-321. https://doi.org/10.1007/s00216-012-5766-4

232. Sestak V, Stariat J, Cermanova J, Potuckova E, Chladek J, Roh J, Bures J, Jansova H, Prusa P, Sterba M, Micuda S, Simunek T, Kalinowski DS, Richardson DR, Kovarikova P (2015) Novel and potent anti-tumor and anti-metastatic di-2-pyridylketone thiosemicarbazones demonstrate marked differences in pharmacology between the first and second generation lead agents. Oncotarget 6:42411-42428. https://doi.org/10.18632/oncotarget.6389

233. Agrawal KC, Sartorelli AC (1969) Potential antitumor agents. II. Effects of modifications in the side chain of 1-formylisoquinoline thiosemicarbazone. J Med Chem 12:771-774. https://doi.org/10.1021/jm00305a011

234. Kalinowski DS, Sharpe PC, Bernhardt PV, Richardson DR (2007) Design, synthesis, and characterization of new iron chelators with anti-proliferative activity: Structure-activity relationships of novel thiohydrazone analogues. J Med Chem 50:6212-6225. https://doi.org/10.1021/jm070839q

235. Richardson DR, Sharpe PC, Lovejoy DB, Senaratne D, Kalinowski DS, Islam M, Bernhardt PV (2006) Dipyridyl thiosemicarbazone chelators with potent and selective antitumor activity form iron complexes with redox activity. J Med Chem 49:6510-6521. https://doi.org/10.1021/jm0606342

236. Potůčková E, Roh J, Macháček M, Sahni S, Stariat J, Šesták V, Jansová H, Hašková P, Jirkovská A, Vávrová K, Kovaříková P, Kalinowski DS, Richardson DR, Šimůnek T (2015) In vitro characterization of the pharmacological properties of the anti-cancer chelator, Bp4eT, and its Phase I metabolites. PLoS One 10:e0139929. https://doi.org/10.1371/journal.pone.0139929

237. Boström J, Hogner A, Llinàs A, Wellner E, Plowright AT (2012) Oxadiazoles in medicinal chemistry. J Med Chem 55:1817-1830. https://doi.org/10.1021/jm2013248

238. Raab A, Feldmann J (2019) Biological sulphur-containing compounds – Analytical challenges. Anal Chim Acta 1079:20-29. https://doi.org/10.1016/j.aca.2019.05.064

239. Komarnisky LA, Christopherson RJ, Basu TK (2003) Sulfur: Its clinical and toxicologic aspects. Nutrition 19:5461. https://doi.org/10.1016/S0899-9007(02)00833-X

240. Markovich D (2001) Physiological roles and regulation of mammalian sulfate transporters. Physiol Rev 81:1499-1533. https://doi.org/10.1152/physrev.2001.81.4.1499 241. Takano Y, Shimamoto K, Hanaoka K (2016) Chemical tools for the study of hydrogen sulfide (H₂S) and sulfane sulfur and their applications to biological studies. J Clin Biochem Nutr 58:7-15. https://doi.org/10.3164/jcbn.15-91 242. Pavlik JW, Noll BC, Oliver AG, Schulz CE, Scheidt WR (2010) Hydrosulfide (HS⁻) coordination in iron porphyrinates. Inorg Chem 49:1017-1026. https://doi.org/10.1021/ic901853p

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102. Leigh M, Raines DJ, Castillo CE, Duhme-Klair AK (2011) Inhibition of xanthine oxidase by thiosemicarbazones, hydrazones and dithiocarbazates derived from hydroxy-substituted benzaldehydes. ChemMedChem 6:1107-1118. https://doi.org/10.1002/cmdc.201100054

103. Kaska WC, Carrano C, Michalowski J, Jackson J, Levinson W (1978) Inhibition of the RNA dependent DNA polymerase and the malignant transforming ability of Rous sarcoma virus by thiosemicarbazone-transition metal complexes. Bioinorg Chem 3:245-254. https://doi.org/10.1016/S0006-3061(00)80198-2

104. Bacher F, Enyedy EA, Nagy NV, Rockenbauer A, Bognár GM, Trondl R, Novak MS, Klapproth E, Kiss T, Arion VB (2013) Copper(II) complexes with highly water-soluble L- and D-proline–thiosemicarbazone conjugates as potential inhibitors of topoisomerase IIα. Inorg Chem 52:8895-8908. https://doi.org/10.1021/ic401079w

105. Bisceglie F, Pinelli S, Alinovi R, Goldoni M, Mutti A, Camerini A, Piola L, Tarasconi P, Pelosi G (2014) Cinnamaldehyde and cuminaldehyde thiosemicarbazones and their copper(II) and nickel(II) complexes: a study to understand their biological activity. J Inorg Biochem 140:111-125. https://doi.org/10.1016/j.jinorgbio.2014.07.014

106. Bisceglie F, Musiari A, Pinelli S, Alinovi R, Menozzi I, Polverini E, Tarasconi P, Tavone M, Pelosi G (2015) Quinoline-2-carboxaldehyde thiosemicarbazones and their Cu(II) and Ni(II) complexes as topoisomerase IIa inhibitors. J Inorg Biochem 152:10-19. https://doi.org/10.1016/j.jinorgbio.2015.08.008

107. Djoko KY, Paterson BM, Donnelly PS, McEwan AG (2014) Antimicrobial effects of copper(II) bis(thiosemicarbazonato) complexes provide new insight into their biochemical mode of action. Metallomics 6:854-863. https://doi.org/10.1039/c3mt00348e.

108. Moore EC, Zedeck MS, Agrawal KC, Sartorelli AC (1970) Inhibition of ribonucleoside diphosphate reductase by 1-formylisoquinoline thiosemicarbazone and related compounds. Biochemistry 9:4492-4498. https://doi.org/10.1021/bi00825a005

109. Moore EC, Booth BA, Sartorelli AC (1971) Inhibition of deoxyribonucleotide synthesis by pyridine carboxaldehyde thiosemicarbazones. Cancer Res 31:235-238.

111. Thelander L, Gräslund A (1983) Mechanism of inhibition of mammalian ribonucleotide reductase by the iron chelate of 1-formylisoquinoline thiosemicarbazone. Destruction of the tyrosine free radical of the enzyme in an oxygen-requiring reaction. J Biol Chem 258:4063-4066. https://doi.org/10.1016/S0021-9258(18)32582-1

112. Shao J, Zhou B, Di Bilio AJ, Zhu L, Wang T, Qi C, Shih J, Yen Y (2006) A Ferrous-Triapine[®] complex mediates formation of reactive oxygen species that inactivate human ribonucleotide reductase. Mol Cancer Ther 5:586-592. https://doi.org/10.1158/1535-7163.MCT-05-0384

113. Popović-Bijelić A, Kowol CR, Lind MES, Luo J, Himo F, Enyedy ÉA, Arion VB, Gräslund A (2011) Ribonucleotide reductase inhibition by metal complexes of Triapine[®] (3-aminopyridine-2-carboxaldehyde thiosemicarbazone): a combined experimental and theoretical study. J Inorg Biochem 105:1422-1431. https://doi.org/10.1016/j.jinorgbio.2011.07.003

114. Akladios FN, Andrew SD, Parkinson CJ (2015) Selective induction of oxidative stress in cancer cells via synergistic combinations of agents targeting redox homeostasis. Bioorganic Med Chem 23:3097-3104. https://doi.org/10.1016/j.bmc.2015.05.006

115. Kalinowski DS, Stefani C, Toyokuni S, Ganz T, Anderson GJ, Subramaniam NV, Trinder D, Olynyk JK, Chua A, Jansson PJ, Sahni S, Lane DJR, Merlot AM, Kovacevic Z, Huang MLH, Lee CS, Richardson DR (2016) Redox cycling metals: pedaling their roles in metabolism and their use in the development of novel therapeutics. Biochim Biophys Acta-Mol Cell Res 1863:727-748. https://doi.org/10.1016/j.bbamcr.2016.01.026

116. Akladios FN, Andrew SD, Parkinson CJ (2016) Increased generation of intracellular reactive oxygen species initiates selective cytotoxicity against the MCF-7 cell line resultant from redox active combination therapy using copper–thiosemicarbazone complexes. J Biol Inorg Chem 21:407-419. https://doi.org/10.1007/s00775-016-1350-2

117. Whitnall M, Howard J, Ponka P, Richardson DR (2006) A class of iron chelators with a wide spectrum of potent antitumor activity that overcomes resistance to chemotherapeutics. Proc Natl Acad Sci 103:14901-14906. https://doi.org/10.1073/pnas.0604979103

118. Kolesar JM, Sachidanandam K, Schelman WR, Eickhoff J, Holen KD, Traynor AM, Alberti DB, Thomas JP, Chitambar CR, Wilding G, Antholine WE (2011) Cytotoxic evaluation of 3-aminopyridine-2-carboxaldehyde thiosemicarbazone in peripheral blood lymphocytes of patients with refractory solid tumors using electron paramagnetic resonance. Exp Ther Med 2:119-123. https://doi.org/10.3892/etm.2010.165

119. Lovejoy DB, Jansson PJ, Brunk UT, Wong J, Ponka P, Richardson DR (2011) Antitumor activity of metalchelating compound Dp44mT is mediated by formation of a redox-active copper complex that accumulates in lysosomes. Cancer Res 71:5871-5880. https://doi.org/10.1158/0008-5472.CAN-11-1218

120. Ishiguro K, Lin ZP, Penketh PG, Shyam K, Zhu R, Baumann RP, Zhu YL, Sartorelli AC, Rutherford TJ, Ratner ES (2014) Distinct mechanisms of cell-kill by triapine, and its terminally dimethylated derivative Dp44mT due to a loss or gain of activity of their copper(II) complexes. Biochem Pharmacol 91:312-322. https://doi.org/10.1016/j.bcp.2014.08.006

121. Haeili M, Moore C, Davis CJC, Cochran JB, Shah S, Shrestha TB, Zhang Y, Bossmann SH, Benjamin WH, Kutsch O, Wolschendorf F (2014) Copper complexation screen reveals compounds with potent antibiotic properties against methicillin-resistant staphylococcus aureus. Antimicrob Agents Ch 58:3727-3736. https://doi.org/10.1128/aac.02316-13

122. Stacy AE, Palanimuthu D, Bernhardt PV, Kalinowski DS, Jansson PJ, Richardson DR (2016) Zinc(II)thiosemicarbazone complexes are localized to the lysosomal compartment where they transmetallate with copper ions to induce cytotoxicity. J Med Chem 59:4965-4984. https://doi.org/10.1021/acs.jmedchem.6b00238

123. Jansson PJ, Sharpe PC, Bernhardt PV, Richardson DR (2010) Novel thiosemicarbazones of the ApT and DpT series and their copper complexes: identification of pronounced redox activity and characterization of their antitumor activity. J Med Chem 53:5759-5769.https://doi.org/10.1021/jm100561b

124. French FA, Blanz EJ, Shaddix SC, Brockman RW (1974) α-(N)-Formylheteroaromatic thiosemicarbazones. inhibition of tumor-derived ribonucleoside diphosphate reductase and correlation with in vivo antitumor activity. J Med Chem 17:172-181. https://doi.org/10.1021/jm00248a006

125. Lovejoy DL, Richardson DR (2002) Novel "hybrid" iron chelators derived from aroylhydrazones and thiosemicarbazones demonstrate selective antiproliferative activity against tumor cells. Blood 100:666-675. https://doi.org/10.1182/blood.V100.2.666

126. Bacher F, Dömötör O, Kaltenbrunner M, Mojović M, Popović-Bijelić A, Gräslund A, Ozarowski A, Filipovico L, Radulovićo S, Enyedy ÉA, Arion VA (2014) Effects of terminal dimethylation and metal coordination of proline-2-formylpyridine thiosemicarbazone hybrids on lipophilicity, antiproliferative activity, and hR2 RNR inhibition. Inorg Chem 53:12595-12609. https://doi.org/10.1021/ic502239u

127. Stefani C, Punnia-Moorthy G, Lovejoy DB, Jansson PJ, Kalinowski DS, Sharpe PC, Bernhardt PV, Richarrdson DR (2011) Halogenated 2'-benzoylpyridine thiosemicarbazone (XBpT) chelators with potent and selective antineoplastic activity: relationship to intracellular redox activity. J Med Chem 54:6936-6948. https://doi.org/10.1021/jm200924c

128. Wang L, He W, Yu Z (2013) Transition-metal mediated carbon–sulfur bond activation and transformations. Chem Soc Rev 42:599-621. https://doi.org/10.1039/C2CS35323G

Journal of Biological Inorganic Chemistry

Ren P, Pike SD, Pernik I, Weller AS, Willis MC (2015) Rh–POP pincer xantphos complexes for C–S and C–H activation. Implications for carbothiolation catalysis. Organometallics 34:711-723. https://doi.org/10.1021/om500984y
 Munjanja L, Brennessel WW, Jones WD (2015) Room-temperature carbon–sulfur bond activation by a reactive (dippe)Pd fragment. Organometallics 34:1716-1724. https://doi.org/10.1021/acs.organomet.5b00194

131. Li Y, Rauchfuss TB (2016) Synthesis of diiron(I) dithiolato carbonyl complexes. Chem Rev 116:7043-7077. https://doi.org/10.1021/acs.chemrev.5b00669

132. Kumar S, Guyon F, Knorr M, Labat S, Miqueu K, Golz C, Strohmann C (2017) Experimental and theoretical studies on the mechanism of the C–S bond activation of Pd^{II} thiolate/thioether complexes. Organometallics 36:1303-1321. https://doi.org/10.1021/acs.organomet.7b00039

133. Lou J, Wang Q, Wu P, Wang H, Zhou YG, Yu Z (2020) Transition-metal mediated carbon–sulfur bond activation and transformations: an update. Chem Soc Rev 49:4307-4359. https://doi.org/10.1039/C9CS00837C

134. Gómez-Saiz P, Gil-García R, Maestro MA, Pizarro JL, Arriortua MI, Lezama L, Rojo T, García Tojal J (2005) Unexpected behaviour of pyridine-2-carbaldehyde thiosemicarbazonatocopper(II) entities in aqueous basic medium-Partial transformation of thioamide into nitrile. Eur J Inorg Chem 3409-3413. https://doi.org/10.1002/ejic.200500326

135. García-Tojal J, Urtiaga MK, Cortés R, Lezama L, Arriortua MI, Rojo T (1994) Synthesis, structure, spectroscopic and magnetic properties of two copper(II) dimers containing pyridine-2-carbaldehyde thiosemicarbazonate (L), [{CuL(X)}₂] (X = Cl or Br). J Chem Soc Dalton Trans 2233-2238. https://doi.org/10.1039/DT9940002233

136. García-Tojal J, Gil-García R, Fouz VI, Madariaga G, Lezama L, Galletero MS, Borrás J, Nollmann FI, García-Girón C, Alcaraz R, Cavia-Saiz M, Muñiz P, Palacios Ò, Samper KG, Rojo T (2018) Revisiting the thiosemicarbazonecopper(II) reaction with glutathione. Activity against colorectal carcinoma cell lines. J Inorg Biochem 180:69-79. https://doi.org/10.1016/j.jinorgbio.2017.12.005

137. Gil-García R, Fraile R, Donnadieu B, Madariaga G, Januskaitis V, Rovira J, González L, Borrás J, Arnáiz FJ, García-Tojal J (2013) Desulfurization processes of thiosemicarbazonecopper(II) derivatives in acidic and basic aqueous media. New J Chem 37:3568-3580. https://doi.org/10.1039/C3NJ00321C

138. Ainscough EW, Brodie AM, Denny WA, Finlay GJ, Ranford JD (1998) Nitrogen, sulfur and oxygen donor adducts with copper(II) complexes of antitumor 2-formylpyridinethiosemicarbazone analogs: physicochemical and cytotoxic studies. J Inorg Biochem 70:175-185. https://doi.org/10.1016/S0162-0134(98)10011-9

139. Gil–García R, Madariaga G, Jiménez-Pérez A, Herrán-Torres, I, Gago-González A, Ugalde M, Januskaitis V, Barrera-García J, Insausti M, Galletero MS, Borrás J, Cuevas JV, Pedrido R, Gómez-Saiz P, Lezama L, García-Tojal J (2023) Perchlorate-induced structural diversity in thiosemicarbazone-copper(II) complexes provides insights to understand the reactivity in acid and basic media. CrystEngComm 25:2213-2226. https://doi.org/10.1039/D3CE00119A

140. Wang YT, Fang Y, Zhao M, Li MX, Ji YM, Han QX (2017) Cu(II), Ga(III) and In(III) complexes of 2acetylpyridine N(4)-phenylthiosemicarbazone: synthesis, spectral characterization and biological activities. MedChemComm 8:2125-2132. https://doi.org/10.1039/C7MD00415J

141. Al-Eisawi Z, Stefani C, Jansson PJ, Arvind A, Sharpe PC, Basha MT, Iskander GM, Kumar N, Kovacevic Z, Lane DJR, Sahni S, Bernhardt PV, Richardson DR, Kalinowski DS (2016) Novel mechanism of cytotoxicity for the selective selenosemicarbazone, 2-acetylpyridine 4,4-dimethyl-3-selenosemicarbazone (Ap44mSe): lysosomal membrane permeabilization. J Med Chem 59:294-312. https://doi.org/10.1021/acs.jmedchem.5b01399

142. Enyedy ÉA, Nagy NV., Zsigó É, Kowol CR, Arion VB, Keppler BK, Kiss T (2010) Comparative solution equilibrium study of the interactions of copper(II), iron(II) and zinc(II) with triapine[®] (3-aminopyridine-2-carbaldehyde thiosemicarbazone) and related ligands. Eur J Inorg Chem 2010:1717-1728. https://doi.org/10.1002/ejic.200901174

143. Dömötör O, May NV., Pelivan K, Kiss T, Keppler BK, Kowol CR, Enyedy EA (2018) A comparative study of α -N-pyridyl thiosemicarbazones: spectroscopic properties, solution stability and copper(II) complexation. Inorg Chim Acta 472:264-275. https://doi.org/10.1016/j.ica.2017.07.001

144. Gil-García R, Gómez-Saiz P, Díez-Gómez V, Madariaga G, Insausti M, Lezama L, Cuevas JV, García-Tojal J (2014) Thiosemicarbazonecopper(II) compounds with halide/hexafluorosilicate anions: structure, water clusters, non-covalent interactions and magnetism. Polyhedron 81:675-686. https://doi.org/10.1016/j.poly.2014.07.032

145. Abras A, Beraldo H, Fantini EO, Borges RHU, Da Rocha MA, Tosi L (1990) Spectroscopic studies of metal complexes containing π -delocalized sulfur ligands. Mössbauer and kinetic studies of iron(II) and iron(III) complexes of the antitumor agent 2-formylpyridine thiosemicarbazone. Inorg Chim Acta 172:113-117. https://doi.org/10.1016/S0020-1693(00)80459-4

146. Borges RHU, Paniago E, Beraldo H (1997) Equilibrium and kinetic studies of iron(II) and iron(III) complexes of some α (N)-heterocyclic thiosemicarbazones. Reduction of the iron(III) complexes of 2-formylpyridine thiosemicarbazone and 2-acetylpyridine thiosemicarbazone by cellular thiol-like reducing agents. J Inorg Biochem 65:267-275. https://doi.org/10.1016/S0162-0134(96)00142-0

147. García-Tojal J, Donnadieu B, Costes JP, Serra JL, Lezama L, Rojo T (2002) Spectroscopic properties of iron thiosemicarbazone compounds. Structure of $[Fe(C_7H_7N_4S)_2] \cdot 1.25H_2O$. Inorg Chim Acta 333:132-137. https://doi.org/10.1016/S0020-1693(02)00802-2

148. Fukumoto K, Sakai A, Hayasaka K, Nakazawa H (2013) Desulfurization and H-migration of secondary thioamides catalyzed by an iron complex to yield imines and their reaction mechanism. Organometallics 32:2889-2892. https://doi.org/10.1021/om400304v

149. Mutoh Y, Sakigawara M, Niiyama I, Saito S, Ishii Y (2014) Synthesis of rhodium–primary thioamide complexes and their desulfurization leading to rhodium sulfido cubane-type clusters and nitriles. Organometallics 33:5414-5422. https://doi.org/10.1021/om500714c

150. Guo W, Liu G, Deng L, Mei W, Zou X, Zhong Y, Zhou X, Fan X, Zheng L (2021) Metal- and oxidant-free green three-component desulfurization and deamination condensation approach to fully substituted 1 H-1,2,4-triazol-3-amines and their photophysical properties. J Org Chem 86:17986-18003. https://doi.org/10.1021/acs.joc.1c02313

151. Gómez-Saiz P, García-Tojal J, Diez-Gómez V, Gil-García R, Pizarro JL, Arriortua MI, Rojo T (2005) Indirect evidences of desulfurization of a thiosemicarbazonecopper(II) system in aqueous basic medium. Inorg Chem Commun 8:259-262. https://doi.org/10.1016/j.inoche.2004.12.016

152. Castiñeiras A, Garcia-Santos I (2008) Desulfuration and cyclization of (Z)-2-[amino(pyridine-2-yl)methylene]hydrazonecarbothioamide in the presence of manganese(II). Zeitschrift fur Anorg und Allg Chemie 634:2907-2916. https://doi.org/10.1002/zaac.200800326

153. Van Poppel LH, Groy TL, Caudle MT (2004) Carbon-Sulfur bond cleavage in bis(N-alkyldithiocarbamato)cadmium(II) complexes: heterolytic desulfurization coupled to topochemical proton transfer. Inorg Chem 43:3180-3188. https://doi.org/10.1021/ic035135v

154. Al-Mutairi AA, Al-Alshaikh MA, Al-Omary FAM, Hassan HM, El-Mahdy AM, El-Emam AA (2019) Synthesis, antimicrobial, and anti-proliferative activities of novel 4-(adamantan-1-yl)-1-arylidene-3- thiosemicarbazides, 4- arylmethyl N-(adamantan-1-yl) piperidine-1-carbothioimidates, and related derivatives. Molecules 24:4308. https://doi.org/10.3390/molecules24234308

155. Jeong H, Kang Y, Kim J, Kim BK, Hong S (2019) Factors that determine thione(thiol)-disulfide interconversion in a bis(thiosemicarbazone) copper(II) complex. RSC Adv 9:9049-9052. https://doi.org/10.1039/c9ra01115c

156. Pedrido R, Romero MJ, Bermejo MR, González-Noya AM, García-Lema I, Zaragoza G (2008) Metal-catalysed oxidation processes in thiosemicarbazones: new complexes with the ligand N-{2-([4-N-ethylthiosemicarbazone]-methyl)phenyl}-p- toluenesulfonamide. Chem Eur J 14:500-512. https://doi.org/10.1002/chem.200700867

157. Pedrido R, Romero MJ, Bermejo MR, Martínez-Calvo M, González-Noya AM, Zaragoza G (2009) Coordinative trends of a tridentate thiosemicarbazone ligand: synthesis, characterization, luminescence studies and desulfurization processes. Dalton Trans 39:8329-8340. https://doi.org/10.1039/b908782f

158. Da Rosa Justim J, Correa Bohs LM, Barreto Martins B, Tribuzy Bandeira KC, Lopes de Melo AP, Carratu Gervini V, Bresolin L, Godoi M, de Menezes Peixoto CR (2021) Electrochemical characterization of isatin-thiosemicarbazone derivatives. J Chem Sci 133:124. https://doi.org/10.1007/s12039-021-01970-x

159. Pelivan K, Frensemeier LM, Karst U, et al (2018) Comparison of metabolic pathways of different α -N-heterocyclic thiosemicarbazones. Anal Bioanal Chem 410:2343-2361. https://doi.org/10.1007/s00216-018-0889-x

160. Gómez-Saiz P, García-Tojal J, Maestro MA, Arnaiz FJ, Rojo T (2002) Evidence of desulfurization in the oxidative cyclization of thiosemicarbazones. Conversion to 1,3,4-oxadiazole derivatives. Inorg Chem 41:1345-1347. https://doi.org/10.1021/ic015625s

161. Gómez-Saiz P, García-Tojal J, Maestro MA, Mahía J, Arnaiz FJ, Lezama L, Rojo T (2003) New 1,3,4oxadiazolecopper(II) derivatives obtained from thiosemicarbazone complexes. Eur J Inorg Chem 2003:2639-2650. https://doi.org/10.1002/ejic.200200689

162. Hiremath SP, Goudar NN, Purohit MG (1981) Synthesis of substituted 1',3',4'-oxadiazolyl-indoles, thiadiazolyl-indoles and triazolyl-indoles. Indian J Chem Sect B 20:388-390.

163. Hiremath SP, Biradar JS, Kudari SM (1984) Synthesis of substituted oxadiazoles, thiadiazoles and triazoles and evaluation of their biological activity. J Indian Chem Soc 61:74-76. https://doi.org/10.5281/zenodo.6325527

164. Fernandes PS, Sonar TM (1986) Synthesis and biological activity of heterocyclic derivatives derived from ethyl-2-hydroxyquinoxaline-3-carboxylate. J Indian Chem Soc 63:427-429. https://doi.org/10.5281/zenodo.6273852

165. Hiremath SP, Sonar VN, Sekhar KR, Purohit MG (1989) Synthesis of substituted oxadiazolylindoles, thiadiazolylindoles and indolylthiazolidinones. Indian J Chem Sect B 28B:626-630.

166. Raman K, Singh HK, Salzman SK, Parmar SS (1993) Substituted thiosemicarbazides and corresponding cyclized
1,3,4-oxadiazoles and their antiinflammatory activity. J Pharm Sci 82:167-169. https://doi.org/10.1002/jps.2600820210
167. Kelarev VI, Karakhanov RA, Gassanov SS, Morozova GV, Kuatbekova KP (1993) Synthesis of 1,3,4-oxa(thia)diazole and 1,2,4-triazolederivatives containing 3-indolylmethyl radicals. Russ J Org Chem 29:388-395.

168. Albar HA, Makki MSI, Faidallah HM (1996) Synthesis of heterocyclic compounds from delta-unsaturated 1,3diketo-esters. Indian J Chem Sect B 35:23-29.

169. Vashi BS, Mehta DS, Shah VH (1996) Synthesis of 2,5-disubstituted-1,3,4-oxadiazole, 1,5-disubstituted-2mercapto-1,3,4-triazole and 2,5-disubstituted-1,3,4-thiadiazole derivatives as potential antimicrobial agents. Indian J Chem Sect B 35:111-115. https://doi.org/10.1002/chin.199617050

170. Rao GR, Mogilaiah K, Sreenivasulu B (1996) Synthesis and antimicrobial activity of 1',2',4'-triazolyl/1',3',4'-thiadiazolyl/1',3',4'-oxadiazolyl-1,8-naphthyridines and related compounds. Indian J Chem Sect B 35:339-344.

171. Omar FA, Mahfouz NM, Rahman MA (1996) Design, synthesis and antiinflammatory activity of some 1,3,4oxadiazole derivatives. Eur J Med Chem 31:819-825. https://doi.org/10.1016/0223-5234(96)83976-6

172. Tripathi P, Pal A, Jancik V, Pandey AK, Singh J, Singh NK (2007) Metal-assisted transformation of Nbenzoyldithiocarbazate to 5-phenyl-1,3,4-oxadiazole-2-thiol in the presence of ethylenediamine, and its first row transition metalcomplexes. Polyhedron 26:2597-2602. https://doi.org/10.1016/j.poly.2006.12.046

173. Hassan AA, Mourad AFE, Abou-Zied AH (2007) Reaction of 1-acylthiosemicarbazides with ethenetetracarbonitrile. J Heterocyclic Chem 44:1171-1176. https://doi.org/10.1002/jhet.5570440532

174. Hassan AA, Mourad AE, Abuo-Zied AH (2007) Benzo- and naphthoimidazoxadiazolediene, naphthobisthiazole as well as naphthothiazine derivatives from 1-acylthiosemicarbazides. Arkivoc 222-235. https://doi.org/10.3998/ark.5550190.0008.124 175. Mekuskiene G, Tumkevicius S, Vainilavicius P (2002) 5-(4,6-diphenyl-2-pyrimidinyl)-1,3,4-oxa(thia)diazoles and 1,2,4-triazoles. J Chem Res 213-215. https://doi.org/10.3184/030823402103171898

176. Dolman SJ, Gosselin F, O'Shea PD, Davies IW (2006) Superior reactivity of thiosemicarbazides in the synthesis of 2-amino-1,3,4-oxadiazoles. J Org Chem 71:9548-9551. https://doi.org/10.1021/j00618730

177. Singh NK, Bharty MK, Dulare R, Butcher RJ (2009) Synthesis and X-ray crystallographic studies of Ni(II) and Cu(II) complexes of [5-(4-pyridyl)-1,3,4] oxadiazole-2-thione/thiol formed by transformation of N-(pyridine-4-carbonyl)-hydrazine carbodithioate in the presence of ethylenediamine. Polyhedron 28:2443-2449. https://doi.org/10.1016/j.poly.2009.04.030

178. Li Z, Zhu A, Yang J (2012) One-pot three-component mild synthesis of 2-aryl-3-(9-alkylcarbazol-3-yl)thiazolin4-ones. J Heterocycl Chem 49:1458-1461.https://doi.org/10.1002/jhet.1047

179. Bharti A, Bharty MK, Kashyap S, Singh UP, Butcher RJ, Singh NK (2013) Hg(II) complexes of 4-phenyl-5-(3-pyridyl)-1,2,4-triazole-3-thione and 5-(4-pyridyl)-1,3,4-oxadiazole-2-thione and a Ni(II) complex of 5-(thiophen-2-yl)-1,3,4-oxadiazole-2-thione: synthesis and X-ray structural studies. Polyhedron 50:582-591. https://doi.org/10.1016/j.poly.2012.11.043

180. Yang Z, She M, Yin B, Hao L, Obst M, Li J (2015) Solvent-dependent turn-on probe for dual monitoring of Ag⁺ and Zn²⁺ in living biological samples. Anal Chim Acta 868:53-59. https://doi.org/10.1016/j.aca.2015.01.052

181. Bao W, Chen C, Yi N, Jiang J, Zeng Z, Deng W, Peng Z, Xiang J (2017) Synthesis of 2-amino-1,3,4-oxadiazoles through elemental sulfur promoted cyclization of hydrazides with isocyanides. Chin J Chem 35:1611-1618. https://doi.org/10.1002/cjoc.201700188

182. Golmohammadi F, Balalaie S, Hamdan F, Maghari S (2018) Efficient synthesis of novel conjugated 1,3,4oxadiazole-peptides. New J Chem 42:4344-4351. https://doi.org/10.1039/c7nj04720g

183. Abu-Hashem AA (2021) Synthesis and antimicrobial activity of new 1,2,4-triazole, 1,3,4-oxadiazole, 1,3,4-thiadiazole, thiopyrane, thiazolidinone, and azepine derivatives. J Heterocycl Chem 58:74-92. https://doi.org/10.1002/jhet.4149

184. Landquist JK (1970) Oxidative cyclisation of ketone thiosemicarbazones. Part I. 4-methyl- and 4-aryl-thiosemicarbazones. J Chem Soc C 63-66. https://doi.org/10.1039/J39700000063

185. Xiao S, Lee W, Chen F, Zavalij PY, Gutierrez O, Davis JT (2020) Oxidation of 8-thioguanosine gives redoxresponsive hydrogels and reveals intermediates in a desulfurization pathway. Chem Commun 56:6981-6984. https://doi.org/10.1039/D0CC02926B

186. Baird IR, Cameron BR, Skerlj RT (2003) Unique chemistry of amino acid dithiocarbamates with Ru(III) bis-βdiketonates. Inorganica Chim Acta 353:107-118. https://doi.org/10.1016/S0020-1693(03)00233-0

187. Ng S, Ziller JW, Farmer PJ (2004) Multiple pathways for the oxygenation of a ruthenium(II) dithiocarbamate complex: S-oxygenation and S-extrusion. Inorg Chem 43:8301-8309. https://doi.org/10.1021/ic048661a

188. Fernández-Fariña S, González-Barcia LM, Romero MJ, García-Tojal J, Maneiro, M, Seco JM, Zaragoza G, Martínez-Calvo M, González-Noya AM, Pedrido R (2022) Conversion of a double-tetranuclear cluster silver helicate into a dihelicate via a rare desulfurization process. Inorg Chem Front 9:531-536. https://doi.org/10.1039/d1qi01308d

189. Alsop L, Cowley AR, Dilworth JR, Donnelly PS, Peach JM, Rider JT (2005) Investigations into some aryl substituted bis(thiosemicarbazones) and their copper complexes. Inorganica Chim Acta 358:2770-2780. https://doi.org/10.1016/j.ica.2005.03.027

190 Gooneh-Farahani S, Anbia M (2023) A review of advanced methods for ultra-deep desulfurization under mild conditions and the absence of hydrogen. J Environ Chem Eng 11:108997. https://doi.org/10.1016/j.jece.2022.108997 191. Aly AA, Hassan AA, Brown AB, Ibrahim MAA, AbdAl-Latif ESSM (2017) Azines from one-pot reaction of thiosemicarbazones. J Sulphur Chem 38:11-17. https://doi.org/10.1080/17415993.2016.1210146

1	
2	
3	
4	192. Kassim K, Hamali MA, Yamin B (2019) A New alternative synthesis of salicylaldazine via microwave irradiation
5	method. J Chem 2019:9546373. https://doi.org/10.1155/2019/9546373
7	193. Qiu W, Shi S, Li R, Lin X, Rao L, Sun Z (2021) A mild, general, metal-free method for desulfurization of thiols
8	and disulfides induced by visible-light. Chinese J Chem 39:1255-1258. https://doi.org/10.1002/cjoc.202000607
9	194. Maji M, Chatterjee M, Ghosh S, Chattopadhyay SK, Wu BM, Mak TCW (1999) Chemistry of ruthenium(II)
10	complexes of the tridentate NNS donor methyl 2-pyridyl ketone 4-(4-tolyl)thiosemicarbazone. Isolation and structural
11	characterisation of a novel ruthenium(II) complex containing a co-ordinated imine of an alpha N haterocyclic katone
12	characterisation of a nover runnehum (1) complex containing a co-ordinated mine of an appla-tv neterocyclic ketone.
13	J Chem Soc, Daiton Trans 135-140. https://doi.org/10.1039/A8063411
15	195. Hassan AA, El-Sheref EM (2010) Chemistry and heterocyclization of dithiobiurea and thioureidoalkylthiourea.
16	J Heterocycl Chem 47:764-784. https://doi.org/10.1002/jhet.406
17	196. Da S. Maia PI, Nguyen HH, Ponader D, Hagenbach A, Bergemann S, Gust R, Deflon VM, Abram U (2012)
18	Neutral gold complexes with tridentate SNS thiosemicarbazide ligands. Inorg Chem 51:1604-1613.
19	https://doi.org/10.1021/ic201905t
20	197 Gil-García R. Zichner R. Díez-Gómez V. Donnadieu B. Madariaga G. Insausti M. Lezama L. Vitoria P. Pedrosa
21	MR García-Toial I (2010) Polyovometallate thiosemicarbazone hybrid compounds. Fur I Inorg Chem 2010:1513-
23	4525 https://doi.org/10.1002/sijo.201000484
24	
25	198. Cowley AR, Dilworth JR, Donnelly PS, Woollard-Shore J (2003) Synthesis and characterisation of new
26	homoleptic rhenium thiosemicarbazone complexes. J Chem Soc Dalton Trans 3:748-754.
27	https://doi.org/10.1039/B210540N
20	199. Matesanz AI, Pastor C, Souza P (2007) Synthesis and structural characterization of a disulphide-bridged
30	tetranuclear palladium(II) complex derived from 3,5-diacetyl 1,2,4-triazole bis(4-ethylthiosemicarbazone). Inorg Chem
31	Commun 10:97-100. https://doi.org/10.1016/j.inoche.2006.09.016
32	200. Zhao Y. Lin Z. He C. Wu H. Duan C (2006) A "turn-on" fluorescent sensor for selective Hg(II) detection in
33	aqueous media based on metal-induced dye formation Inorg Chem 45:10013-10015
34 35	https://doi.org/10.1021/io061067b
36	$\frac{1}{1000} = \frac{1}{1000} = \frac{1}{10000000000000000000000000000000000$
37	201. Zhang P, Shi B, Zhang Y, Lin Q, Yao H, You XM, Wei TB (2013) A selective fluorogenic chemodosimeter for
38	Hg^{2+} based on the dimerization of desulfurized product. Tetrahedron 69:10292-10298.
39	https://doi.org/10.1016/j.tet.2013.10.024
40	202. Hwang KS, Park KY, Kim D Bin, Chang SK (2017) Fluorescence sensing of Ag ⁺ ions by desulfurization of an
41 42	acetylthiourea derivative of 2-(2-hydroxyphenyl)benzothiazole. Dyes Pigm 147:413-419.
43	https://doi.org/10.1016/j.dyepig.2017.08.041
44	203. Bulak E, Dogan I, Varnali T, Schwederski B, Gunal SE, Lönnecke P, Bubrin M, Kaim W (2021) An Acyclic
45	Diaminocarbene Complex of Platinum Formed by Desulfurization of 1 3-Bis(3-methylpyridin-2-yl)thiourea Eur I
46	Inora Chem 2:2425 2422 https://doi.org/10.1002/ajjc.202100277
47	204 Singh & Chatumodi I. Dhattachama & (2012) Studies of surthering structural features of Cu(1) this share 2
48 70	204. Singn S, Chaturvedi J, Bhattacharya S (2012) Studies of synthesis, structural features of Cu(1) thiophene-2-
50	thiocarboxylates and unprecedented desulfurization of Cu(II) thiocarboxylate complexes. Dalton Trans 41:424-431.
51	https://doi.org/10.1039/C1DT10629E
52	205. Bigoli F, Deplano P, Mercuri L, Pellinghelli MA, Pintus G, Trogu EF (1999) Unusual desulfurization of a nickel
53	dithiolene by bis(2-diphenylphosphinophenyl)phenylphosphine (tp) to produce $Ni(tp)(R4btimdt)$ [R4btimdt = 5,5'-
54	bis(1,3-dialkyl-4-imidazolidine-2-thione-4-thiolate], the first complex of this class of ligands. Chem Commun
55 56	698:2093-2094. https://hdl.handle.net/11381/1458647
57	206. Chawla SK. Arora M. Nättinen K. Rissanen K (2006) Unique conner ion catalyzed hydrolytic cleavage of C-
58	N(2) hond of thiosemicarbazide Polyhedron 25:627-634 https://doi.org/10.1016/j.poly.2005.05.021
59	1(2) cond of unosemical outlide. For ynder on 25.027-054. https://doi.org/10.1010/j.p01y.2005.05.021
60	

207. Voitekhovich SV, Lyakhov AS, Ivashkevich LS, Ivashkevich OA (2020) Copper-assisted desulfurization of 1-R-tetrazole-5-thiols under complexation. Inorg Chem Commun 114:107827-107830. https://doi.org/10.1016/j.inoche.2020.107827

208. Tardito S, Bussolati O, Maffini M, Tegoni M, Giannetto M, Dall'Asta V, Franchi-Gazzola R, Lanfranchi M, Pellinghelli MA, Mucchino C, Mori G, Marchiò L (2007) Thioamido coordination in a thioxo-1,2,4-triazole copper(II) complex enhances nonapoptotic programmed cell death associated with copper accumulation and oxidative stress in human cancer cells. J Med Chem 50:1916-1924. https://doi.org/10.1021/jm061174f

209. Madshus IH (1988) Regulation of intracellular pH in eukaryotic cells. Biochem J. 250:1-8. https://doi.org/10.1042/bj2500001

210. Nunes P, Guido D, Demaurex N (2015) Measuring phagosome pH by ratiometric fluorescence microscopy. J Vis Exp e53402. https://doi.org/10.3791/53402

211. Asokan A, Cho MJ (2002) Exploitation of intracellular pH gradients in the cellular delivery of macromolecules. J Pharm Sci 91:903-913. https://doi.org/10.1002/jps.10095

212. Porcelli AM, Ghelli A, Zanna C, Pinton P, Rizzuto R, Rugolo M (2005) pH difference across the outer mitochondrial membrane measured with a green fluorescent protein mutant. Biochem Biophys Res Commun 326:799-804. https://doi.org/10.1016/j.bbrc.2004.11.105

213. Wilson AF, Simmons DH (1970) Organ and Whole Body Cell pH. Proc Soc Exp Biol Med 134:127-130. https://doi:10.3181/00379727-134-34743

214. Park HJ, Lim CS, Kim ES, Han HJ, Lee TH, Chun HJ, Cho BR (2012) Measurement of pH values in human tissues by two-photon microscopy. Angew Chem Int Ed 51:2673-2676. https://doi.org/10.1002/anie.201109052

215. Persi E, Duran-Frigola M, Damaghi M, Roush WR, Aloy P, Cleveland JL, Gillies RJ, Ruppin E (2018) Systems analysis of intracellular pH vulnerabilities for cancer therapy. Nat Commun 9:2997. https://doi.org/10.1038/s41467-018-05261-x

216. Radi R (2018) Oxygen radicals, nitric oxide, and peroxynitrite: redox pathways in molecular medicine. Proc Natl Acad Sci U S A 115:5839-5848. https://doi.org/10.1073/pnas.1804932115

217. Hu X, Dong D, Xia M, Yang Y, Wang J, Su J, Sun L, Yu H (2020) Oxidative stress and antioxidant capacity: development and prospects. New J Chem 44:11405-11419. https://doi.org/10.1039/D0NJ02041A

218. Di Meo S, Reed TT, Venditti P, Victor VM (2016) Role of ROS and RNS sources in physiological and pathological conditions. Oxid Med Cell Longev 2016:ID 1245049. https://doi.org/10.1155/2016/1245049

219. Zhang Y, Wong HS (2021) Are mitochondria the main contributor of reactive oxygen species in cells. J Exp Biol 224:jeb221606. https://doi.org/10.1242/jeb.221606

220. Neha K, Haider MR, Pathak A, Yar MS (2019) Medicinal prospects of antioxidants: a review. Eur J Med Chem 178:687-704. https://doi.org/10.1016/j.ejmech.2019.06.010

221. Pisoschi AM, Pop A, Iordache F, Stanca L, Predoi G, Serban AI (2021) Oxidative stress mitigation by antioxidants - An overview on their chemistry and influences on health status. Eur J Med Chem 209:112891. https://doi.org/10.1016/j.ejmech.2020.112891

222. Jungwirth U, Kowol CR, Keppler BK, Hartinger CG, Berger W, Heffeter P (2011) Anticancer activity of metal complexes: involvement of redox processes. Antioxidan Redox Signal 15:1085-1127. https://doi.org/10.1089/ars.2010.3663

223. Eteshola EOU, Haupt DA, Koos SI, Siemer LA, Morris DL (2020) The role of metal ion binding in the antioxidant mechanisms of reduced and oxidized glutathione in metal-mediated oxidative DNA damage. Metallomics 79-91. https://doi.org/10.1039/C9MT00231F

224. Ouyang Y, Peng Y, Li J, Holmgren A, Lu (2018) Modulation of thiol-dependent redox system by metal ions *via* thioredoxin and glutaredoxin systems. Metallomics 10:218-228. https://doi.org/10.1039/C7MT00327G

225. Arunachalam V, Tummanapelli AK, Vasudevan S (2019) The multiple dissociation constants of glutathione disulfide: interpreting experimental pH-titration curves with: Ab initio MD simulations. Phys Chem Chem Phys 21:9212-9217. https://doi.org/10.1039/C9CP00761J

226. Falcone E, Ritacca AG, Hager S, Schueffl H, Vileno B, Khoury YE, Hellwig P, Kowol CR, Heffeter P, Sicilia E, Faller P (2022) Copper-catalyzed glutathione oxidation is accelerated by the anticancer thiosemicarbazone Dp44mT and further boosted at lower pH. J Am Chem Soc 144:14758-14768. https://doi.org/10.1021/jacs.2c05355

227. Nutting LA, Weber EM, Tryon JL (1967) Metabolic removal of sulfur from 1-methylisatin 3-thiosemicarbazone.J Virol 1:650-651. https://doi.org/10.1128/jvi.1.3.650-651.1967

228. Creasey, WA, Agrawal KC, Capizzi RL, Stinson KK, Sartorelli AC (1972) Studies of the antineoplastic activity and metabolism of α-(N)-heterocyclic carboxaldehyde thiosemicarbazones in dogs and mice. Cancer Res 32:565-572.
229. DeConti RC, Toftness BR, Agrawal KC, Tomchick R, Mead JA, Bertino JR, Sartorelli AC, Creasey WA (1972) Clinical and pharmacological studies with 5-hydroxy-2-formylpyridine thiosemicarbazone. Cancer Res 32:1455-1462.
230. Pelivan K, Frensemeier L, Karst U, Koellensperger G, Bielec B, Hager S, Heffeter P, Kepplerae BK, Kowol CR (2017) Understanding the metabolism of the anticancer drug Triapine[®]: Electrochemical oxidation, microsomal incubation and: In vivo analysis using LC-HRMS. Analyst 142:3165-3176. https://doi.org/10.1039/C7AN00902J

231. Stariat J, Šesták V, Vávrová K, Nobilis M, Kollárová Z, Klimeš J, Kalinowski DS, Richardson DR, Kovaříková P (2012) LC-MS/MS identification of the principal in vitro and in vivo phase I metabolites of the novel thiosemicarbazone anti-cancer drug, Bp4eT. Anal Bioanal Chem 403:309-321. https://doi.org/10.1007/s00216-012-5766-4

232. Sestak V, Stariat J, Cermanova J, Potuckova E, Chladek J, Roh J, Bures J, Jansova H, Prusa P, Sterba M, Micuda S, Simunek T, Kalinowski DS, Richardson DR, Kovarikova P (2015) Novel and potent anti-tumor and anti-metastatic di-2-pyridylketone thiosemicarbazones demonstrate marked differences in pharmacology between the first and second generation lead agents. Oncotarget 6:42411-42428. https://doi.org/10.18632/oncotarget.6389

233. Agrawal KC, Sartorelli AC (1969) Potential antitumor agents. II. Effects of modifications in the side chain of 1-formylisoquinoline thiosemicarbazone. J Med Chem 12:771-774. https://doi.org/10.1021/jm00305a011

234. Kalinowski DS, Sharpe PC, Bernhardt PV, Richardson DR (2007) Design, synthesis, and characterization of new iron chelators with anti-proliferative activity: Structure-activity relationships of novel thiohydrazone analogues. J Med Chem 50:6212-6225. https://doi.org/10.1021/jm070839q

235. Richardson DR, Sharpe PC, Lovejoy DB, Senaratne D, Kalinowski DS, Islam M, Bernhardt PV (2006) Dipyridyl thiosemicarbazone chelators with potent and selective antitumor activity form iron complexes with redox activity. J Med Chem 49:6510-6521. https://doi.org/10.1021/jm0606342

236. Potůčková E, Roh J, Macháček M, Sahni S, Stariat J, Šesták V, Jansová H, Hašková P, Jirkovská A, Vávrová K, Kovaříková P, Kalinowski DS, Richardson DR, Šimůnek T (2015) In vitro characterization of the pharmacological properties of the anti-cancer chelator, Bp4eT, and its Phase I metabolites. PLoS One 10:e0139929. https://doi.org/10.1371/journal.pone.0139929

237. Boström J, Hogner A, Llinàs A, Wellner E, Plowright AT (2012) Oxadiazoles in medicinal chemistry. J Med Chem 55:1817-1830. https://doi.org/10.1021/jm2013248

238. Raab A, Feldmann J (2019) Biological sulphur-containing compounds – Analytical challenges. Anal Chim Acta 1079:20-29. https://doi.org/10.1016/j.aca.2019.05.064

239. Komarnisky LA, Christopherson RJ, Basu TK (2003) Sulfur: Its clinical and toxicologic aspects. Nutrition 19:5461. https://doi.org/10.1016/S0899-9007(02)00833-X

240. Markovich D (2001) Physiological roles and regulation of mammalian sulfate transporters. Physiol Rev 81:1499-1533. https://doi.org/10.1152/physrev.2001.81.4.1499 241. Takano Y, Shimamoto K, Hanaoka K (2016) Chemical tools for the study of hydrogen sulfide (H₂S) and sulfane sulfur and their applications to biological studies. J Clin Biochem Nutr 58:7-15. https://doi.org/10.3164/jcbn.15-91 242. Pavlik JW, Noll BC, Oliver AG, Schulz CE, Scheidt WR (2010) Hydrosulfide (HS⁻) coordination in iron porphyrinates. Inorg Chem 49:1017-1026. https://doi.org/10.1021/ic901853p

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