Supporting Information

Silica Nanoparticle/Fluorescent Dye Assembly Capable of Ultrasensitively Detecting Airborne Triacetone Triperoxide: Proof-of-Concept Detection of Improvised Explosive Devices in the Workroom.

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General methods
N-(N'-Boc-piperidin-4-yl)-4-bromonaphthalene-1,8-dicarboxylmonoimide (AR43p)S03-S05
N-(Piperidin-4-yl)-4-bromonaphthalene-1,8-dicarboxylmonoimide (AR43d)S05-S07
N-(N'-Boc-piperidin-4-yl)-4-[2-(4-Boc-piperazin-1-yl)pyrimidin-5-yl]naphthalene-1,8-
dicarboxylmonoimide (AR82p)S08-S14
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noimide (AR82s)S15-S66
N-(Piperidin-4-yl)-4-[2-(piperazin-1-yl)pyrimidin-5-yl]naphthalene-1,8-dicarboxylmonoimide
(AR82d)
N-(N'-Boc-piperidin-4-yl)-4-[2-(4-Boc-piperazin-1-yl)pyridin-5-yl]naphthalene-1,8-dicar-
boxylmonoimide (AR83p)
N-(Piperidin-4-yl)-4-[2-(4-Boc-piperazin-1-yl)pyridin-5-yl]naphthalene-1,8-dicarboxylmono-
imide (AR83s)
N-(Piperidin-4-yl)-4-[2-(piperazin-1-yl)pyridin-5-yl]naphthalene-1,8-dicarboxylmonoimide
(AR83d)
N-(N'-Boc-piperidin-4-yl)-4-[(4-Boc-piperazin-1-yl)phenyl]naphthalene-1,8-dicarboxyl-
monoimide (AR90p)
N-(Piperidin-4-yl)-4-[(4-Boc-piperazin-1-yl)phenyl]naphthalene-1,8-dicarboxylmonoimide
(AR90s)S134-S148
N-(Piperidin-4-yl)-4-[(4-piperazin-1-yl)phenyl]naphthalene-1,8-dicarboxylmonoimide
(AR90d)

Synthesis of chemical probes and additional experiments.

General methods: Melting points were determined on an electrothermal melting point Gallenkamp apparatus and are uncorrected. Infrared Spectra were recorded with the potassium bromide pellet method, with a JASCO FT/IR-4200 spectrometer. Nuclear magnetic resonance (NMR) spectra were recorded with Varian Mercury-300 and Varian Unity Inova-400 spectrometers at room temperature (25°C), with CDCl₃, CD₃CN and CD₃OD as solvents. Chemical shifts (δ) were reported in parts per million (*ppm*) relative to the residual solvent peaks, rounded to the nearest 0.01 for ¹H-NMR and 0.1 for ¹³C-NMR. Spin-spin coupling constants (J) in ¹H-NMR were given in Hz, rounded to the nearest 0.1 Hz. Peak multiplicity was indicated as follows: s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet) and br (broad). Relative integrals were given in ¹H-NMR too. All ¹³C NMR were recorded with complete proton decoupling. Carbon types, structure assignments and attribution of peaks were determined from ¹³C-DEPT-NMR. NMR spectra were analyzed using MestReNova NMR data processing software. MALDI-TOF mass spectra were measured with a MALDI-TOF Bruker Autoflex Mass Spectrometry instrument, using DCTB (trans-2-[3-(4-tert-butylphenyl)-2methyl-2-propenylidene]malononitrile) or DIT (dithranol) as matrixes, in modes positive or negative. The atomic mass of the molecular ion (and/or fragments) per elementary charge were reported in dimensionless quantities. Absorption spectra were acquired with a Hitachi U-3900 spectrometer, in one-centimeter quartz cells at 25°C. Emission spectra were recorded with a Hitachi F-7000 FL or a modular Edinburgh Instruments FLS980 spectrofluorometers, in onecentimeter quartz cells at 25°C. Solvatochromism Tests: All samples were freshly prepared in each different solvent. Photos were taken with a Canon (EOS M3) camera with a 22 mm lens. Absorption spectra were acquired with a Hitachi U-3900 spectrometer, in one-centimeter quartz cells at 25°C. Emission spectra were recorded with Hitachi F-7000 FL spectrofluorometer, in one-centimeter quartz cells at 25°C. The solvents were, if it is not said otherwise: 1: H₂O, 2: MeOH (methanol), 3: DMSO (dimethyl sulfoxide), 4: DMF (N,N'-dimethylformamide), 5: MeCN (acetonitrile), 6: Acetone, 7: AcOEt (ethyl acetate), 8: THF (tetrahydrofuran), 9: CHCl₃, 10: CH₂Cl₂ (dichloromethane), 11: Toluene, 12: Et₂O (diethyl ether), 13: n-Hx (hexane), 14: c-Hx (cyclohexane). Fluorescence Lifetime Decays: Chromophore solutions were freshly prepared in a concentration of 1-10 µM in the corresponding solvent. The decay was fitted (black) with a sum of two exponentials by convolution with the instrumental response function (red). The quality of the fit was judged by χ^2 values and the plot of the weighted residues. Measurements were made with a modular spectrometer Edinburgh Instruments FLS980, with a source excitation pulsed lasers of picosecond diode: 366-380 nm, 398-410 nm, 437-446 nm, 470-478 nm, 505-515 nm and 635 nm. One-centimeter quartz cells, at 25 °C, were employed. Quantum Yields: Fresh solutions made from high-purity dyes (or mixtures of dye with an additive) and solvent were used. Integrating sphere was used as the method and three measurements were performed for each sample in order to calculate the average. Solutions were prepared in a concentration of 1-10 µM in the corresponding solvent. The excitation wavelength is indicated in each case. Measurements were made with a modular spectrometer Edinburgh Instruments FLS980. One-centimeter quartz cells, at 25°C, were employed.

Synthesis of *N*-(*N'*-Boc-piperidin-4-yl)-4-bromonaphthalene-1,8-dicarboxylmonoimide (AR43p)



In a 100 ml flask, provided with a magnetic stirrer, 2 g of 4-bromo-1,8-naphtalic anhydride (7.22 mmol) and 2.9 g of 4-amino-1-Boc-piperidine (14.44 mmol) were dissolved in 40 ml of ethanol and the mixture was stirred 8 hours at 60°C. After that, the mixture was filtered, cooling to room temperature and the solvent was evaporated under reduced pressure. The organic solid was purified by column chromatography (SiO₂, CH₂Cl₂), getting 3.15 g of AR43p, white solid, 95% yield. m.p.: 148 – 150°C. IR (ATR, cm⁻¹): 2977 – 2849, 1689 (C=O), 1649 (C=O), 1419 (C-O), 1359, 1344, 1244, 1173 (C-N), 1143 (-CH₃), 1101, 1032 (C-Br), 981, 894, 774, 747, 424. ¹H-NMR (300 MHz, CDCl₃) δ (ppm): 8.48 (d, *J* = 7.1 Hz, 1H, H_{Ar}), 8.35 (d, *J* = 8.4 Hz, 1H, H_{Ar}), 8.23 (d, *J* = 7.9 Hz, 1H, H_{Ar}), 7.86 (d, *J* = 7.9 Hz, 1H, H_{Ar}), 7.70 (t, *J* = 7.9 Hz, 1H, H_{Ar}), 5.17 – 5.02 (m, 1H, CH), 4.26 (s, 2H, CH₂), 2.69 (m, 4H, 2×CH₂), 1.64 (m, 2H, CH₂), 1.45 (s, 9H, 3×CH₃). ¹³C-NMR (75MHz, CDCl₃) δ (ppm): 163.3 (C=O), 163.2 (C=O), 154.3 (C=O), 132.4 (C_{Ar}H), 131.5 (C_{Ar}H), 130.7 (C_{Ar}H), 130.6 (C_{Ar}H), 129.8 (C_{Ar}), 129.5 (C_{Ar}), 128.3 (C_{Ar}), 127.6 (C_{Ar}H), 122.9 (C_{Ar}), 122.0 (C_{Ar}), 79.2 (Cq), 51.6 (CH), 43.8 (CH₂), 28.2 (CH₃), 27.9 (CH₂). HRMS (MALDI, DCTB-) m/z: calculated for C₂₂H₂₃BrN₂O4: 458.0836 (M⁻); found 458.0802.



Figure S1. ¹H-NMR (CDCl₃, 300 MHz) of AR43p.







Figure S4. HRMS (MALDI, DCTB-) of AR43p.

Synthesis of N-(piperidin-4-yl)-4-bromonaphthalene-1,8-dicarboxylmonoimide (AR43d)



In a 50 ml flask with a magnetic stirrer, 150 mg of N-(N'-Boc-piperidin-4-yl)-4bromonaphthalene-1,8-dicarboxylmonoimide (AR43, 0.33 mmol) were dissolved in 15 ml of chloroform and 2 ml of trifluoroacetic acid (26 mmol) were added, and the mixture was stirred for 4 hours at room temperature. Afterwards, 10 ml of water were poured into the flask and the mixture was neutralized employing a 40% sodium hydroxide solution until it reached a neutral pH. Then, 20 ml of water were added and the mixture was extracted with chloroform (3×30 ml). The organic extracts were combined, dried over anhydrous Na₂SO₄ and the solvent was evaporated under reduced pressure. The product AR43d was obtained quantitatively as a white solid, 100% yield (117 mg). m.p.: 88 – 89°C. IR (ATR, cm⁻¹): 3359 (N-H), 2973 – 2830, 1686 (C=O), 1579, 1477, 1417, 1390, 1357, 1238, 1162 (C-N), 1121, 1097, 1076, 1049, 998 (C-Br), 932, 861, 807, 761, 642, 543, 511, 462, 408. ¹H-NMR (300 MHz, CD₃OD) δ (ppm): 8.61 – 8.55 (m, 2H, $2 \times H_{Ar}$), 8.36 (d, J = 7.9 Hz, 1H, H_{Ar}), 8.10 (d, J = 7.9 Hz, 1H, H_{Ar}), 7.90 (dd, J = 7.9 Hz, 1H, H8.5, 7.3 Hz, 1H, H_{Ar}), 5.31 (m, 1H, CH), 3.60 – 3.52 (m, 2H, NH y ½CH₂), 3.27 – 2.93 (m, 5H, 2×CH₂ y ¹/₂CH₂), 2.04 – 1.95 (m, 2H, CH₂). ¹³C-NMR (75MHz, CDCl₃) δ (ppm): 163.7 (C=O), 133.1 (CArH), 132.1 (CArH), 131.3 (CArH), 131.1 (CArH), 128.9 (CAr), 128.1 (CArH), 123.3 (C_{Ar}), 122.4 (C_{Ar}), 53.4 (Cq), 50.6 (CH), 45.5 (CH₂), 27.7 (CH₂). HRMS (MALDI, DCTB+) m/z: calculated for $C_{17}H_{15}BrN_2O_2$: 359.0390 (M⁺ + H); found 359.0391.



Figure S6. ¹³C-NMR (CDCl₃, 75 MHz) of AR43d.



Figure S7. DEPT NMR (CDCl₃, 75 MHz) of AR43d.



Figure S8. HRMS (MALDI, DCTB+) of AR43d.

Synthesis of *N*-(*N*'-Boc-piperidin-4-yl)-4-[2-(4-Boc-piperazin-1-yl)pyrimidin-5-yl]naphthalene-1,8-dicarboxylmonoimide (AR82p)



250 mg of N-(N'-Boc-piperidin-4-yl)-4-bromonaphthalene-1,8-dicarboxylmonoimide (AR43p, 0.54 mmol), 223 mg of 2-(4-Boc-piperazin-1-yl)pyrimidine-5-boronic acid pinacol ester (0.57 mmol) and 752 mg of potassium carbonate (5.44 mmol) were dissolved in a mixture of toluene:butanol:water (4:1:2 ml) under nitrogen atmosphere and the catalyst, Pd(PPh₃)₄ (5% mmol) was added. The resulting mixture was stirred overnight at 110°C. After that, the solvent was evaporated under reduce pressure, the solid was dissolved in CH₂Cl₂ and washed with water (3×100 ml). The organic extracts were combined, dried over anhydrous Na₂SO₄ and the solvent was evaporated under reduce pressure. The organic solid was purified by column chromatography (SiO₂, DCM:MeOH 50:1) to obtain 280 mg of AR82p, yellow solid, 80% yield. m.p.: 226 – 229°C (decomposition). IR (ATR, cm⁻¹): 3437 (N-H), 2977 – 2860 (C-H), 1699 (C=O), 1657 (C=O), 1585, 1513, 1450,1420, 1352, 1244, 1169 (C-N), 1106, 999, 948, 780, 759. ¹H-NMR (300 MHz, CDCl₃) δ (ppm): 8.57 (d, J = 7.6 Hz, 2H, 2×H_{Ar}), 8.44 (m, 2H, $2 \times H_{Ar}$, 8.21 (d, J = 8.4 Hz, 1H, H_{Ar}), 7.70 (dd, J = 8.4, 7.3 Hz, 1H, H_{Ar}), 7.60 (d, J = 7.6 Hz, 1H, H_{Ar}), 5.16 (m, 1H, CH), 4.26 (s, 2H, CH₂), 3.89 (m, 4H, 2×CH₂), 3.53 (m, 4H, 2×CH₂), 2.72 (m, 4H, 2×CH₂), 1.65 (m, 2H, CH₂), 1.47 (m, 18H, 6×CH₃). ¹³C-NMR (75MHz, CDCl₃) δ (ppm): 164.5 (C=O), 164.3 (C=O), 161.2 (C=O), 158.2 (C_{Ar}H), 140.7 (C_{Ar}), 132.3 (C_{Ar}), 132.2 (CAr), 132.1 (CAr), 131.6 (CArH), 131.1 (CArH), 130.1 (CAr), 128.7 (CArH), 128.5 (CArH), 127.7 (C_{Ar}H), 127.3 (C_{Ar}H), 123.6 (C_{Ar}), 121.0 (C_{Ar}), 80.3 (Cq), 79.6 (Cq), 52.0 (CH), 43.9 (CH₂), 28.6 (CH₃), 28.5 (CH₃), 28.4 (CH₂). HRMS (MALDI, DIT-) m/z: calculated for C₃₅H₄₂N₆O₆: 642.3160 (M⁻); found 642.3181. UV-Vis (CHCl₃), λ_{max} nm (log ε): 380 (4.3). Ø (MCH, %): 92.87. τ (375 nm, MCH, ns): 2.853 (χ^2 : 1.080).



Figure 1. ¹³C-NMR (CDCl₃, 75 MHz) of AR82p.



Figure S11. DEPT NMR (CDCl₃, 75 MHz) of AR82p.



Figure S12. HRMS (MALDI, DIT-) of AR82p.

Solvatochromism:

The concentration of the compound was 20 μ M and the excitation wavelength was 380 nm.



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Figure S13. Solvatochromism of AR82p.
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Figure S14. Solvatochromism of AR82p under visible and 366 nm light.



Fluorescence decay lifetime (τ):

The solvent used was MCH and the concentration of the compound was 20 μ M.



Figure S15. Fluorescence decay lifetime of AR82p.

Water acceptance test:

The solvent used was MeCN and THF and the concentration of the compound was 20 μ M.

Figure 2. Water acceptance test of AR82p in MeCN under visible (up) and UV (down) light.



Figure S16. Water acceptance test of AR82p in THF under visible (up) and UV (down) light.



There was an increase in emission intensity from 80% water (AIE). This phenomenon was also studied using THF as solvent and the same amounts of water. In THF, an increase in the intensity of fluorescent emission was observed from 70% water. In both cases, there was a shift of the emission maximum from yellow-brown (535–545 nm) to green (about 475 nm).

Figure S17. Emission intensity graph (left) and graph of relationship between the emission intensity and the wavelength of the maximum of emission (right) as a function of the amount of water added for AR82p in THF.



pH test:

The proportion of water that the compound could accept was 40%. The concentration of the compound was 20 μ M and the solvent used was MeCN.

Figure S18. pH test of AR82p under visible (up) and UV (down) light.



Quenching of emission at very acidic pH and yellow fluorescence at very basic pH were seen. **Cations and anions test:**

The concentration of the compound was 20 µM and the solvent used was MeCN. Figure S19. Cations test of AR82p under visible (up) and UV (down) light.





Under visible light, a response to Pd^{2+} was observed, while under UV light, a fluorescence change to blue was taken place to Au^{3+} .

The concentration of the compound was 20 μ M and the solvent used was MeCN.



Figure S20. Anions test of AR82p under visible (up) and UV (down) light.

No significant changes were observed.

Oxidants and reductants test:

The solvent used was MeCN and concentration of the compound was 20 μ M.

Figure S21. Oxidants and reductants test of AR82p under visible (up) and UV (down) light.



*HCl is included in all experiments in a close position to HNO₃ to distinguish between the redox action of HNO₃ and a possible effect due only to the concomitant acidity of nitric acid.

No significant changes were observed.

Synthesis of *N*-(piperidin-4-yl)-4-[2-(4-Boc-piperazin-1-yl)pyrimidin-5-yl]naphthalene-1,8-dicarboxylmonoimide (AR82s)



150 mg of N-(piperidin-4-yl)-4-bromonaphthalene-1,8-dicarboxylmonoimide (AR43d, 0.42 mmol), 172 mg of 2-(4-Boc-piperazin-1-yl)pyrimidin-5-yl boronic acid pinacol ester (0.44 mmol) and 451 mg of potassium carbonate (3.26 mmol) were dissolved in a mixture of toluene:butanol:water (4:1:2 ml) under nitrogen atmosphere and, then, the Pd(PPh₃)₄ (5% mmol) was added. The resulting mixture was stirred overnight at 110°C, the solvent was evaporated under reduce pressure. The solid was dissolved in CH2Cl2 and washed with water (3×100 ml). The organic extracts were combined, dried over anhydrous Na₂SO₄ and the solvent was evaporated under reduce pressure. The organic solid was purified by column chromatography (SiO₂, CH₂Cl₂:MeOH 50:1) getting 87 mg of AR82s, yellow solid, 38% yield. m.p.: 302 – 304°C (decomposition). IR (ATR, cm⁻¹): 3440 (N-H), 2922 – 2480, 1693 (C=O), 1650 (C=O), 1585, 1509, 1450 (C-O), 1402, 1351, 1238, 1162 (C-N), 1121, 998, 947, 780, 753, 687, 642, 508. ¹H-NMR (300 MHz, CDCl₃) δ (ppm): 8.62 (dd, J = 7.4, 1.7 Hz, 2H, 2×H_{Ar}), 8.50 (s, 2H, $2 \times H_{Ar}$), 8.25 (dd, J = 8.5, 1.1 Hz, 1H, H_{Ar}), 7.73 (dd, J = 8.5, 7.4 Hz, 1H, H_{Ar}), 7.64 (d, J = 7.4 Hz, 1H, H_{Ar}), 5.26 – 5.13 (m, 1H, CH), 3.93 (m, 4H, 2×CH₂), 3.56 (m, 4H, 2×CH₂), 3.29 (m, 2H, CH₂), 2.89 – 2.65 (m, 4H, 2×CH₂), 2.13 (s, 1H, NH), 1.73 (m, 2H, CH₂), 1.51 (s, 9H, 3×CH₃). ¹³C-NMR (75 MHz, CDCl₃) δ (ppm): 164.5 (C=O), 164.3 (C=O), 164.1 (C=O), 158.2 (C_{Ar}H), 154.9 (C_{Ar}), 140.7 (C_{Ar}), 132.1 (C_{Ar}), 131.9 (C_{Ar}H), 131.8 (C_{Ar}), 131.7 (CAr), 131.5 (CAr), 131.4 (CAr), 130.1 (CAr), 129.0 (CAr), 127.8 (CArH), 127.5 (CArH), 123.3 (CAr), 122.2 (CAr), 121.0 (CAr), 80.4 (Cq), 44.1 (CH₂), 28.6 (CH₃), 25.8 (CH₂). HRMS (MALDI, DCTB+) m/z: calculated for $C_{30}H_{34}N_6O_4$: 543.2714 (M⁺ + H); found 543.2689. UV-Vis (CHCl₃), λ_{max} nm (log ε): 385 (4.9). Ø (CHCl₃, %): 73.00. τ (375 nm, CHCl₃, ns): 3.962 (χ^2 : 1.072).



Figure S23. ¹³C-NMR (CDCl₃, 75 MHz) of AR82s.



Figure S24. DEPT NMR (CDCl₃, 75 MHz) of AR82s.



Figure S25. HRMS (MALDI, DCTB+) of AR82s.

Solvatochromism:

The concentration of the compound was 20 μ M and the excitation wavelength was 370 nm.





Figure S27. Solvatochromism of AR82s under visible and 366 nm light.



Fluorescence decay lifetime (τ):

The solvent used was CHCl₃. The concentration of the compound was 20 μ M.



Figure S28. Fluorescence decay lifetime of AR82s.

Water acceptance test:

The solvent used was acetone and the concentration of the compound was 20 μ M.

Figure S29. Water acceptance test of AR82s, acetone under visible (up) and UV (down) light.



Figure S30. Water aceptance test of AR82s in THF under visible (up) and UV (down) light.



There was a very slight increase in emission intensity at 80% and 90% of water (AIE) and the result using THF as a solvent was very similar. The emission maximum was shifted from brown – yellow region (530 - 555 nm), with intermediate amounts of water, to the green region (490 – 505 nm), with high water content.

Figure S31. Emission intensity graph (left) and graph of relationship between the emission intensity and the wavelength of the maximum of emission (right) as a function of the amount of water added for AR82s using THF as a solvent.



pH test:

The amount of water that the compound could accept was 20%. The concentration of the compound was 20 μ M and the solvent used was acetone.

Figure S32. pH test of AR82s under visible (up) and UV (down) light.



The most significative change was the quenching of the emission at very acidic pH and the fluorescence color change to white at very basic pH. For intermediate pH, the fluorescence was turned brownish.

Cations and anions test:

The concentration of the compound was 20 μ M and the solvent used was acetone.

Figure S33. Cations test of AR82s under visible (up) and UV (down) light.



The most significant changes under UV light were the turn of the fluorescence to white against Sn^{2+} and Au^{3+} .

The concentration of the compound was 20 μM and the solvent used was acetone.

Figure S34. Anions test of AR82s under visible (up) and UV (down) light.



The only response of the probe to the different anionic analytes was against HSO4-.

Oxidants and reductants test:

The solvent used was acetone and concentration of the compound was 20 μ M.

Figure S35. Oxidants and reductants test of AR82s under visible (up) and UV (down) light.

	R	HCI* HNO ₃ Oxn Hz H ₂ O ₂	R m-CPBA TNB TNT	R m-CPBA TATP HMTD
Visible light				~~~~
ngnv	~	****		日本大大日



*HCl is included in all experiments in a close position to HNO₃ to distinguish between the redox action of HNO₃ and a possible effect due only to the concomitant acidity of nitric acid.

The most significant change was the turn of the fluorescence to blue against oxone.

Preliminary solvents test:

In order to determine the fluorescence variation of the compound under study in the presence of TATP, some tests were carried out for the probe in 9 different solvents. The AR82s concentration was 20 μ M and the TATP was added in excess (7 mg in each vial). All the tests were performed at room temperature and the photographs were taken immediately after the addition of TATP and again after 24 hours.

Figure S36. Pairs of AR82s (left) and AR82s with excess of TATP (right) in different solvents at time 0 h (up) and 24 h (down).



The biggest changes were seen in DMF and MeCN with a fluorescence color change and in toluene and DCM with a decrease in the emission intensity.

Work concentration test:

In order to choose an optimum concentration for the next quantitative studio, the absorbance and fluorescence of the probe was checked to be linear while concentration changes was small, even when working with an excess of m-CPBA (7000 μ M). The solvent used for this test was DCM and the excitation wavelength was 370 nm for all measurements.

Figure S37. Absorbance spectrum (up left), absorbance profile at 385 nm (up right), fluorescence spectrum (down left) and fluorescence profile at 497 nm (down right) of increasing concentrations of AR82s solution in DCM.



Figure S38. Absorbance spectrum (up left), absorbance profile at 385 nm (up right), fluorescence spectrum (down left) and fluorescence profile at 507 nm (down right) of increasing concentrations of AR82s with an excess of m-CPBA solution in DCM.





The ideal working concentration should stay at 0.1 of absorbance or less, to avoid inner filter effects, possible dynamic quenching or stacking processes. It was implied a concentration below 11 μ M. Taking the results into account, the chosen concentration was 2.5 μ M, value around which the Lambert – Beer Law (or pseudo – Lambert – Beer linear behavior for fluorescence) was fulfilled.

Solvents test with TATP:

For all solvents, the working concentration of AR82s was 2.5 μ M, the excitation wavelength was 370 nm and the temperature for all test was 25°C.

In the kinetic studies, TATP was in excess (20 mM) and the fluorescence emission measurements were made during 25 minutes in the case of DMF and 60 minutes in MeCN, toluene and DCM at 5 minutes time intervals. In the titration, the TATP concentration was between 0 and 35000 μ M and it was added directly as solid. The measurements were carried out immediately after the addition of the probe.

Figure S39. Study of AR82s with TATP in DMF. Kinetic study (up left) and profile as function as time at 415 nm and 550 nm simultaneously (up right) in presence of TATP excess. Titration (down left) and fluorescence profile at 550 nm (down right) under increasing concentrations of TATP.





There was a drop in the emission intensity band immediately after adding TATP. According to the fluorescence spectra, the LOD of TATP was somewhere between 15000 and 20000 μ M, too high for a good probe.

Figure S40. Kinetic study at different times under visible (up) and UV (down) light in DMF. In each photo, left vial was contained the probe and right vial was contained the probe and an excess of TATP.



Figure S41. Titration of AR82s with an excess of TATP under visible (up) and UV (down) light in DMF.

μΜ	0	250	500	1000	5000	10000	15000	20000	25000	30000	35000
Visible light											
UV light (366 nm)) ·	-	

Figure S42. Study of AR82s with TATP in MeCN. Kinetic study (up left) and profile as function as time at 535 nm (up right) in presence of TATP excess. Titration (down left) and fluorescence profile at 530 nm (down right) under increasing concentrations of TATP.



There was a drop in the emission intensity band immediately after adding TATP which was stabilized after 40 minutes. According to the fluorescence spectra, the LOD of TATP was somewhere between 1000 and 5000 μ M, a little high for a good probe.

Figure S43. Kinetic study at different times under visible (up) and UV (down) light in MeCN. In each photo, left vial was contained the probe and right vial was contained the probe and an excess of TATP.

	t=0	t=60 min	t=24 h
Visible light			
UV light (366 nm)			

Figure S44. Titration of AR82s with an excess of TATP under visible (up) and UV (down) light in MeCN.

μΜ	0	250	500	1000	5000	10000	15000	20000	25000	30000	35000
Visible light								the second			
UV light (366 nm)											

Figure S45. Study of AR82s with TATP in toluene. Kinetic study (up left) and profile as function as time at 460 nm (up right) in presence of TATP excess. Titration (down left) and fluorescence profile at 460 nm (down right) under increasing concentrations of TATP.



There was a decrease in the emission intensity band immediately after adding TATP which was stabilized after 5 minutes. According to the fluorescence spectra, the LOD of TATP was

somewhere between 5000 and 10000 μM with a small initial increase and then a considerable decrease in emission intensity.

Figure S46. Kinetic study at different times under visible (up) and UV (down) light in toluene. In each photo, left vial was contained the probe and right vial was contained the probe and an excess of TATP.



Figure S47. Titration of AR82s with an excess of TATP under visible (up) and UV (down) light in toluene.

μΜ	0	250	500	1000	5000	10000	15000	20000	25000	30000	35000
Visible light											
UV light (366 nm)											

Figure S48. Study of AR82s with TATP in DCM. Kinetic study (up left) and profile as function as time at 500 nm (up right) in presence of TATP excess. Titration (down left) and fluorescence profile at 500 nm (down right) under increasing concentrations of TATP.



There was a decrease in the emission intensity band immediately after adding TATP which was stabilized after 30 minutes. According to the fluorescence spectra, the LOD of TATP was somewhere between 0 and 250 μ M, it was the solvent that gave the lowest detection limit.

Figure S49. Kinetic study at different times under visible (up) and UV (down) light in DCM. In each photo, left vial was contained the probe and right vial was contained the probe and an excess of TATP.

	t=0	t=60 min	t=24 h
Visible light			
UV light (366 nm)			

Figure S50. Titration of AR82s with an excess of TATP under visible (up) and UV (down) light in DCM.



Titration with TATP in DCM and LOD calculations:

The working concentration of AR82s was 2.5 μ M, the TATP was directly as a solid and its concentration was between 0 and 25 mM. The measurements were carried out immediately after the addition of TATP. The excitation wavelength was 370 nm and the fluorescence changes were registered at 25°C.

Figure S51. Titration (up left), fluorescence profile at 500 nm (up right) and calibration for the LOD calculation using emission values at 500 nm (down) of a 2.5 μ M of AR82s solution in DCM under increasing concentrations of TATP.



An initial increase in the emission intensity was observed, followed by a significant decrease in emission. In addition, the blank (sample only containing probe) was measured three times obtaining an intensity value of 596722.725 a.u. and a standard deviation of 5682.909 a.u. The LOD of AR82s with TATP was calculated.

Calculation of the LOD by linear regression + false positive and negative at 500 nm.

For this method, the R software was used and the value of the LOD was 0.87 μ M.



Figure S52. Calculation of LOD by the R software.

Titration with m-CPBA in DCM and LOD calculations:

To compare the results and the possibility of having different processes, not only the response to TATP was evaluated. The changes with m-CPBA, other strong oxidant, were also measured, determining their optical changes and the LOD.

The working concentration of AR82s was 2.5 μ M, the excitation wavelength was 370 nm and the fluorescence changes were registered at 25°C. In the kinetic study, the m-CPBA was in excess (7 mM) and the fluorescence emission measurements were made during the subsequent 2 hours, at the increasing time intervals. In the titration, the concentration of the m-CPBA was between 0 and 20 mM, measuring the variation of the fluorescence after each addition.

Figure S53. Kinetic study (left) and fluorescence profile at 430 nm and 500 nm simultaneously (right) of a 2.5 μ M of AR82s solution in DCM under increasing concentrations of m-CPBA.



The fluorescence profile showed that the emission intensity in the band with a maximum of 500 nm fell drastically just after the addition of m-CPBA, at which point it begun to increase again, reaching a certain stability after 30 minutes. Initially, there was no band in the area of 430 nm but it begun to appear after the addition of m-CPBA and it increased gradually. It reached some stability after 30 minutes.

Figure S54. Titration (up left), fluorescence profile at 430 nm and 500 nm simultaneously (up right) and calibration for the LOD calculation using emission values at 430 nm (down right) and 500 nm (down left) of a 2.5 μ M of AR82s solution in DCM under increasing concentrations of m-CPBA.



In addition, the blank (sample only containing probe) was measured three times obtaining an intensity value of 553148.256 a.u. and a standard deviation of 5055.503 a.u. The LOD of AR82s with TATP was calculated.

Calculation of the LOD by linear regression + false positive and negative at 500 nm.

For this method, the R software was used and the value of the LOD was 2.1 nM.



Figure S55. Calculation of LOD by the R software.

Calculation of the LOD by linear regression + false positive and negative at 430 nm.

For this method, the R software was used and the value of the LOD was 2.1 nM.



Figure S56. Calculation of LOD by the R software.

TATP gas detection by modified nanoparticles:

Several nanoparticles samples were modified by adsorption of the probe and tested as potential TATP sensors. In all three cases, 100 mg of the SiO₂ nanoparticles $(5 - 15 \mu m particle size, 99.5\%$ trace metal basis, Aldrich Chemistry) and 1 mg of the probe were added to a 5 ml of CHCl₃ and thin foil was used to cover the vial to avoid degradation of the nanoparticles by the action of light. The resulting suspension was stirred at 60 °C under a stream of nitrogen until complete evaporation of solvent (approximately 30 minutes). After that, nanoparticles with the adsorbed compound (AR82s·SiO₂) were washed with hexane (3×3 ml) and the solvent residues were dried with a stream of nitrogen, working in the dark throughout the process.

The supported probe was evaluated by several methods that was given complementary information about the influence of different analytes. In a first instance, the color variation of the nanoparticles before and after exposure to TATP was tested. Afterwards, the detection capacity was evaluated by a more quantitative approach by registering changes in fluorescence intensity (by comparing the fluorescence quantum yield for higher precision and in the position of the maximum of emission. Finally, a quantitative study for the specific detection of TATP vapors was also performed.



Figure S57. Color comparation between the nanoparticles with the probe over glass plates before and after the exposure to TATP.

The increase in fluorescence of the solid was recorded by measuring the fluorescence of the solids and determining the increase in fluorescence quantum yield (\emptyset F). The most straightforward way to detect the presence of the analyte was the increase in fluorescence of the modified material. However, the measurements were sometimes reliant on the position of the solid in the fluorometer (especially in powder samples) and these factors were difficult to control without designing a specific device for doing so. As a consequence, fluorescence quantum yields were the most trustworthy way to measure it quantitatively.

The initial evaluation of the AR82s adsorbed on SiO_2 nanoparticles against TATP in vapor was carried out by placing, at the bottom of an Eppendorf, 30 mg of the corresponding nanoparticles and 4 mg of TATP in the upper part of the Eppendorff, which was placed horizontally. The Eppendorf was covered with tin foil to prevent the degradation of the nanoparticles by action of light and was heated, at a temperature not higher than 60°C, until the complete evaporation of TATP. Once evaporated, the probe and the TATP were kept in contact for ten minutes more before proceeding to measure the emission and the excitation spectrums and the quantum yield of the nanoparticles, both exposed to the TATP and before the exposure, with an Edinburg Instrument FLS – 980 fluorometer at an excitation wavelength of 370 nm an at 25°C.



*Figure S58. Response of fluorescence (down) and excitation (up) of AR82s adsorbed in SiO*₂ *nanoparticles.*

The variation of quantum yields between the probe and the probe after the exposure to TATP was a parameter that provides lower errors, less than 2%.

$$\frac{\phi_{probe+TATP}}{\phi_{probe}} = 0.081$$

Fluorescence intensity with the probe was around 12 times lower after the exposure to TATP vapor.

Titration with TATP in vapor phase:

The titration of AR82s adsorbed onto SiO₂ nanoparticles was carried out by placing 15 mg of nanoparticles in an Eppendorf, covered with tin foil to avoid degradation of the nanoparticles by the action of light, and increasing amounts of TATP (from 0.025 to 2 mg) in another Eppendorf. Both were connected to each other and to an air stream, air flow, 100 cm³/min, provided by a compressor. Working in the absence of light, TATP was heated, to a temperature below 50°C, until its complete evaporation, keeping probe and TATP under recirculation in the system for 10 additional minutes after the complete TATP evaporation. After this time, the emission measurements of the nanoparticles exposed to TATP were carried out in an Edinburg Instrument FLS-980 fluorometer at an excitation wavelength of 370 nm at 25°C.



Figure S59. Microfluidic device used for titrations in vapor phase (left) and the actual aspect of the system (right).

The microfluidic device dimensions were: Eppendorf: outer top diameter 11mm, inner top diameter 9 mm, outer bottom diameter 5 mm, inner bottom diameter 3 mm, volume 1.5 ml, height (with lid) 40 mm, height (without lid) 38 mm. Tubing: outer diameter 3 mm, inner diameter 1.5 mm, length (flow tube) 25 mm, length (return tube) 35 mm.

TATP (mg)	Time required for complete evaporation
0.032	1'41"
0.048	1'45"
0.083	2'14"
0.114	2'42"
0.158	3'10"
0.203	3'21"
0.247	3'57"
0.333	4'26"
0.483	5'03"
0.800	7'45"
1.000	11'23''
1.400	13'13"
2.100	19'37"

Figure S60. Time required for total evaporation of the different amounts of TATP used in the experiment.

The calibration was performed in steady state, fixed quantities of TATP were vaporized in the presence of a sample containing 15 mg of $AR82s \cdot SiO_2$ nanoparticles and the fluorescence increase was measured 10 minutes after the complete vaporization of each TATP sample.


Figure S61. Response of AR82s·SiO₂ exposed to different quantities of TATP.

Figure S62. Titration (up left), fluorescence profile at 535 nm (up right) and calibration for the LOD calculation using emission values at 535 nm (down) of AR82s·SiO₂ under increasing concentrations of TATP in vapor phase.



The blank (sample only containing probe) was measured three times obtaining an intensity value of 5454206.667 a.u. and a standard deviation of 154231.172 a.u. The LOD was calculated by linear regression + false positive and negative at 535 nm.

For this method, the R software was used and the value of the LOD was $1.303 \cdot 10^{-5}$ mg or 0.01303 µg.



Figure S63. Calculation of LOD by the R software.

Mobile phone application:

In this case an OPPO Find X3 Pro 5G (dedicated to the application) was used to detect the presence of TATP in a vapor flow. The application was developed in collaboration with J. Rafael Santana Tejada (Movilmatica, www.movilmatica.com).

The purpose of the application was to detect the amount of TATP from an imagen of AR82s \cdot SiO₂ sample exposed to TATP vapors. The user had to configure the system correctly in order to obtain a suitable result. The application has two systems for determining the amount of TATP by RGB:

- Interval search: each sample exposed to a known amount of TATP was configured with several colors, in which the target could be included. To detect the amount of TATP to which the nanoparticles were exposed, a color was selected and an interval search was carried out. Example: suppose we have a target color 50, 240, 70 and we have the following intervals [40, 130, 60], [10,250, 60] and [55, 244, 71]. The comparison algorithm will detect that the target 50, 240, 70 is in the interval [40, 230, 60] 50, 240, 70 [55, 244, 71].
- Radio search: suppose we have a target color 50, 240, 70 and we work with a radius of 10. The intervals we have are [40, 230, 60], [10, 250, 60] and [85, 244, 41]. When the compare algorithm will detect that the target 50, 240, 70 is in the radius of the interval [40, 230, 60] since if we add the radius ±10 we would have this radius [40, 230, 60] or [60, 250, 80].

The system of the application was composed of the following components:

- Mobile application: it was developed with IONIC technology with which we have the possibility of generating an APK for Android or IPA for IOS.
- *Laravel api*: communication with the backend where queries were made from the mobile application to the database.
- Database: system of tables designed to be able to store colors dynamically and to meet the needs of the application.



Figure S64. System structure of the mobile application

System configuration: The application is made up of different configurable sections:

- Login: to be able to enter the system, access credentials are required.
- Menu: allows us to view and access to the different options of the application.
- Users: the list of users authorized to access the application is displayed, as well as the option to delete them.
- User settings: personal data (name, email, password...) can be changed from this screen.
- New user: allows you to create new users by filling in the required information. The data will be verified before granting access to the application.
- Color list: list of the different nanoparticle colors for each amount of TATP in vapor phase, accessed from "menu" and then "my colors". In this screen, colors can be deleted and the data updated at any time as well as watch the rest of the stored colors.
- New color: to add a new color to a TATP measurement, we click on "add color" and select the TATP quantity and add an RGB color, manually or by selecting it on an image. The latter is the fastest method as it allows you to save and continue adding colors.
- Color detection: once the system is configured with the different TATP measurements, in the "analysis" screen you can take a picture or select an image from the gallery and choose the point where you want to detect how much TATP it has been exposed to. The images are very important because the higher the quality, the higher the accuracy in detecting RGB color.

a) Sign in	b) orte ubu	Cupers C
	Configuration user	► ionic
	I My colours	ionic@nomail.com
	20 My users	> soporte ubu
UNIVERSIDAD DE BURGOS	(¹) Logout	
🖾 Email		
Ô Password		
Forgot pass?		
	Analysis New colour New user Menu	Analysis New colour New user Menu



The figures showed an example for a 2 mg image obtained by the device from previous section and the system with the color configuration should detect it. As we can see, the system is correct since the range of colors is in the correct range. In the lower part we have the option to change the radio search algorithm that is named in the introduction section.



Figure S66. Images of the data base

The app was based on:

Ionic <u>https://ionicframework.com</u>, Laravel, <u>https://laravel.com/docs/8.x/eloquent-resources</u>, Data base, <u>https://www.mysql.com</u>

The application was part of an autonomous system for capturing and quantitative evaluation of fluorescent probe images based on mobile telephony composed of: (1) customized display box (2) 360° metal bracket (3) customized APP and (4) OPPO Find X3 Pro5G. The system allows photographs to be taken with a mobile camera from very short distances of a sensor material previously exposed to an explosive atmosphere. For this, the sensing material was placed in a dark box with a metal holder and a UV lamp (366 nm) to illuminate the material. The application to compare the photo with a database of images of the material exposed to different amounts of TATP in vapor phase is loaded on the mobile phone so, that from the comparison and the calculated calibration curve, the amount of the explosive that has come into contact with the AR82s·SiO₂ can be extracted.







Figure S67. Images of the data base used for search and compare selected samples.
a) Selected image (0.428 mg of TATP in system Figure 2a).
b) A positive result.
c) Selected image (1.720 mg of TATP in system Figure 2a).
d) A positive result.
e) Selected image (0.061 mg of TATP in system Figure 2a).
f) A positive result.
g) Selected image (no TATP in system Figure 2a).
h) A negative result.
The yellow arrows indicate the point where the selected color is measured



Figure S68. The dark box (left) with a metal support (right).

Validation of fluorescent device in real life scenarios:

The experiments were carried out in an office with dimensions $3.07 \times 3.54 \times 2.15$ m (height \times length \times width). An air compressor was placed inside it, which was generated a continuous stream of clean air, that was passed through the TATP in the direction of the point where the particles were arranged, and three heaters that help regulate the temperature inside the room.



Figure S69. Office experiments with AR82s·SiO₂ with TATP in vapor phase. a) shows the dimensions and the arrangement of the elements inside the office. b) shows the compressor used in the experiments.

Varying amounts of TATP (specified in each experiment) were placed 30 cm from the source of the airflow. Approximately 0.75 mg of AR82s \cdot SiO₂ were placed on the surface of a cover glass, adhered to it with the help of a small amount of adhesive tape. This process was repeated 6 times and the cover glasses were placed at increasing distances from the point where the TATP was located (10 - 25 - 50 - 100 - 150 - 200 cm).



Figure S70. Office experiments with AR82s·SiO₂ with TATP in the vapour phase.
a) arrangement of the TATP in front of yhe airflow outlet.
b) distribution of the sensing nanoparticles in the room.
c) and d) assembly for measurement of the particles.



To carry out the measurements in the fluorometer, they were covered with a quartz sheet that ensured the nanoparticles remained in a fixed position. This sheet had then to be removed to leave the particles exposed to TATP vapors during the experiment. A measurement of the particles without being subjected to TATP was also made before and after remaining the same time as the particles that were subjected to TATP vapors to use as a reference.

Fluorescence measurements were made for each group of particles before and after the exposure to TATP vapors using a excitation wavelength of 370 nm. Photos of the particles were taken under identical conditions. Two representative experiments are detailed.

Experiment 4. 250 mg of TATP at 26°C during 30 minutes.

Of the 252.3 mg of TATP initially weighed, only 38.6 mg were evaporated under the temperature and time conditions used in this experiment, remaining 216.7 mg of TATP as solid. The experimental procedure was similar to the previously explained.



Figure S71. Comparative photograph taken under 366 nm light before and after exposure to TATP vapors at 26°C during 30 minutes.

Experiment 5. 250 mg of TATP at 28°C during 30 minutes.

Of the 252.3 mg of TATP initially weighed, only 97.3 mg were evaporated under the temperature and time conditions used in this experiment, remaining 155 mg of TATP as solid. The experimental procedure for these measurements was similar to the previous one.



Figure S72. Comparative photograph taken under 366 nm light before and after exposure to TATP vapors at 28°C during 30 minutes.

From the experiments, the following Figures have been made, representing the normalized emission intensity variation before and after exposure to TATP vapors versus distance to the TATP source.







The following graph shows all the data together to facilitate comparison.



Figure S74. Summary of experiments at different temperatures. Normalized emission intensity variation before and after exposure to TATP versus distance to the TATP source.



Figure S75. Summary 3D graph of experiments at different temperatures. Increase in normalized emission intensity variation before and after exposure to TATP versus distance to the TATP source.

Following the results of the previous series of experiments, it was observed that the best temperature conditions to perform the experiment was around 26°C. Therefore, a series of 6 additional experiments were performed in which room temperature was raised up to 30°C by the use of an air heater. Once this temperature was reached, the heater was turned off and the room temperature was left to decrease until an average temperature of 26°C was obtained. The experiments had a duration of 30 minutes.

The experiments were carried out in an office with dimensions $3.07 \times 3.54 \times 2.15$ m (height \times length \times width). The following figures illustrate both the dimensions and the arrangement of the different elements in the room when proceeding with the experiments.



Figure S76. Images of the dimensions and arrangement of the elements inside the office. **S47**

Inside it, an air compressor was placed, which generated a continuous stream of clean air that passed through the TATP the direction of the points where the particles were arranged, and a heater that helped regulate the temperature inside the room. Once a temperature around 30°C was reached, the heater was turned off and the temperature was left to reach an average temperature of 26°C to perform the experiments.

For all experiments, 250 mg of TATP were placed 30 cm from the source of the airflow. Approximately, 0.75 mg of AR82s·SiO₂ nanoparticles were placed on the surface of a cover glass, adhered to it with the help of a small amount of adhesive tape, as the previous experiments. This process was repeated six times and the cover glasses were placed at increasing distances from the point where the TATP was located (10, 25, 50, 100, 150 and 200 cm).



Figure S77. Arrangement the AR82·SiO₂ nanoparticles in the office.
a) Arrangement of the TATP in front of the airflow outlet.
b) distribution of sensing nanoparticles in the room.
c) assembly for measurement of the nanoparticles.

Once TATP and nanoparticles were placed, these were left in the presence of TATP vapors for a period of 30 minutes. To carry out the measurements in the fluorometer, they were covered with a quartz sheet that ensured the particles remained in a fixed position. This sheet was removed to leave the particles exposed to TATP vapors during the experiment. A measurement of the particles without being subjected to TATP was also made before and after remaining the same time as the particles that were subjected to TATP vapors to use as a reference.

Fluorescence measurements were made for each group of particles before and after the exposure to TATP vapors. Photos of the particles were taken under identical conditions and the excitation wavelength was 370 nm.

Experiment 7: at 26 °C during 30 minutes.

Of the 256 mg of TATP initially weighed, only 23 mg were evaporated under the temperature and time conditions used in this experiment, remaining 233 mg of TATP as solid.



Figure S78. Emission spectra of the nanoparticles before and after exposure to TATP vapors at 26 - 30°C during 30 minutes.

10 cm	25 cm	50 cm	100 cm	150 cm	200 cm	
						Before exposure
P.V.	1	Sal				After exposure

Figure S79. Comparative photograph taken under 366 nm light before and after exposure to TATP vapors at 26°C during 30 minutes.



Figure S80. Histogram plotting the peak emission before and after exposure to TATP vapors at 26°C during 30 minutes.

The average increase in the emission intensity for particles were located 50 cm or less from the TATP source was 206%, while the increase for the emission intensity for nanoparticles were located one meter or more was only 112%.

Figure S81.	Increment in flu	orescence	emission	intensity of	of sensing	nanoparticles	as a
	function	of their dis	tance from	n the TAT	<i>P source</i> .		

Sensing AR82s·SiO ₂ particles position	Increase in emission intensity
10 cm	170%
25 cm	214%
50 cm	234%
100 cm	119%
150 cm	113%
200 cm	105%

Experiment 8: at 26 °C during 30 minutes.

Of the 256 mg of TATP initially weighed, only 23 mg were evaporated under the temperature and time conditions used in this experiment, remaining 233 mg of TATP as solid in this case too.







Figure S83. Comparative photograph taken under 366 nm light before and after exposure to TATP vapors at 26°C during 30 minutes.



Figure S84. Histogram plotting the peak emission before and after exposure to TATP vapors at 26°C during 30 minutes.

The average increase in the emission intensity for particles were located 50 cm or less from the TATP source was 205%, while the increase for the emission intensity for nanoparticles were located one meter or more was only 118%.

Sensing AR82s·SiO2 particles position	Increase in emission intensity
10 cm	166%
25 cm	268%
50 cm	180%
100 cm	117%
150 cm	115%
200 cm	122%

Figure S85. Increment in fluorescence emission intensity of sensing nanoparticles as a function of their distance from the TATP source.

Experiment 9: at 26 °C during 30 minutes.

Of the 251 mg of TATP initially weighed, only 36 mg were evaporated under the temperature and time conditions used in this experiment, remaining 215 mg of TATP as solid.



Figure S86. Emission spectra of the nanoparticles before and after exposure to TATP vapors at 26°C during 30 minutes.



Figure S87. Comparative photograph taken under 366 nm light before and after exposure to TATP vapors at 26°C during 30 minutes.



Figure S88. Histogram plotting the peak emission before and after exposure to TATP vapors at 26°C during 30 minutes.

The average increase in the emission intensity for particles were located 50 cm or less from the TATP source was 214%, while the increase for the emission intensity for nanoparticles were located one meter or more was only 126%.

Sensing AR82s·SiO2 particles position	Increase in emission intensity
10 cm	302%
25 cm	192%
50 cm	148%
100 cm	138%
150 cm	146%
200 cm	93%

Figure S89. Increment in fluorescence emission intensity of sensing nanoparticles as a function of their distance from the TATP source.

Experiment 10: at 26 °C during 30 minutes.

Of the 250 mg of TATP initially weighed, only 36 mg were evaporated under the temperature and time conditions used in this experiment, remaining 214 mg of TATP as solid.



Figure S90. Emission spectra of the nanoparticles before and after exposure to TATP vapors at 26°C during 30 minutes.



Figure S91. Comparative photograph taken under 366 nm light before and after exposure to TATP vapors at 26°C during 30 minutes.



Figure S92. Histogram plotting the peak emission before and after exposure to TATP vapors at 26°C during 30 minutes.

The average increase in the emission intensity for particles were located 50 cm or less from the TATP source was 257%, while the increase for the emission intensity for nanoparticles were located one meter or more was only 117%.

Sensing AR82s·SiO ₂ particles position	Increase in emission intensity
10 cm	249%
25 cm	229%
50 cm	293%
100 cm	118%
150 cm	115%
200 cm	117%

Figure S93. Increment in fluorescence emission intensity of sensing nanoparticles as a function of their distance from the TATP source.

Experiment 11: at 26 °C during 30 minutes.

Of the 254 mg of TATP initially weighed, only 32 mg were evaporated under the temperature and time conditions used in this experiment, remaining 222 mg of TATP as solid.



Figure S94. Emission spectra of the nanoparticles before and after exposure to TATP vapors at 26°C during 30 minutes.



Figure S95. Comparative photograph taken under 366 nm light before and after exposure to TATP vapors at 26°C during 30 minutes.



Figure S96. Histogram plotting the peak emission before and after exposure to TATP vapors at 26°C during 30 minutes.

The average increase in the emission intensity for particles were located 50 cm or less from the TATP source was 235%, while the increase for the emission intensity for nanoparticles were located one meter or more was only 110%.

Figure S97.	Increment in fluoresc	cence emission	intensity of	sensing	nanoparticles	as a
	function of the	eir distance fro	m the TATP	source.		

Sensing AR82s·SiO2 particles position	Increase in emission intensity
10 cm	147%
25 cm	199%
50 cm	359%
100 cm	122%
150 cm	106%
200 cm	102%

Experiment 12: at 26 °C during 30 minutes.

Of the 254 mg of TATP initially weighed, only 32 mg were evaporated under the temperature and time conditions used in this experiment, remaining 222 mg of TATP as solid.



Figure S98. Emission spectra of the nanoparticles before and after exposure to TATP vapors at 26°C during 30 minutes.



Figure S99. Comparative photograph taken under 366 nm light before and after exposure to TATP vapors at 26°C during 30 minutes.



Figure S100. Histogram plotting the peak emission before and after exposure to TATP vapors at 26°C during 30 minutes.

The average increase in the emission intensity for particles were located 50 cm or less from the TATP source was 299%, while the increase for the emission intensity for nanoparticles were located one meter or more was only 133%.

Figure S101. Increment in fluorescence emission intensity of sensing nanoparticles as a function of their distance from the TATP source.

Sensing AR82s·SiO2 particles position	Increase in emission intensity
10 cm	338%
25 cm	357%
50 cm	201%
100 cm	161%
150 cm	117%
200 cm	122%

For an easier visualization of the whole data, the following graphs have been made, representing the normalized emission intensity variation before and after exposure to TATP vapors versus distance to the TATP source.

Figure S102. Summary of experiments 7 - 12 with TATP vapor at 26°C. Emission intensity variation before and after exposure to TATP (left) and emission intensity proportionality factor (divided between the target, right) versus distance to the TATP source in both cases.



Maximum emission intensity data from previous experiments were organized according to the distance to the TATP source, representing, on the one hand, emission before contact with TATP vapors and, on the other hand, emission after contact with said vapors. These data were also normalized with respect to the maximum of emission from each experiment.



Figure S103. Summary of experiments 7 – 12. Normalized emission intensity variation before and after exposure to TATP versus distance to the TATP source.

After this, a common initial emission level was established using the lowest value measured for the nanoparticles prior to the exposure to TATP vapors. And finally, the subtraction was made between the final and initial emission.

Figure S104. Summary of experiments 7 - 12. Normalized emission intensity setting a common initial particulate emission level before and after exposure to TATP vapors (left) and variation of the normalized emission intensity (right) as a function of the distance to the TATP source in both cases.





Figure S105. Summary of experiments 7 - 12. 3D representation of the variation of the normalized emission intensity as a function of the distance to the TATP source.

A selection of the measurements was made, discarding outliers and choosing only 3 measurements for each of the distances to the TATP source. Using these measurements, the average and standard deviation of the normalized variation of the emission intensity for each point were calculated and represented.

Distance (am)	Experiment					
Distance (cm)	7	8	9	10	11	12
10	0.312	0.043	0.669	0.252	0.321	0.055
25	0.533	0.234	0.182	0.564	0.166	0.349
50	0.168	0.035	0.076	0.366	0.116	0.271
100	0.069	0.014	0.059	0.025	0.035	0.050
150	0.089	0.071	0.074	0.029	0.022	0.145
200	0.018	0.181	-0.054	0.014	0.004	0.011

Figure S106. Values of variation of normalized the emission intensity as a function of the distance to the TATP source.

Figure S107. Values of the average and standard deviation of normalized emission variation as a function of the distance to the TATP source.

Distance (cm)	Average normalized emission variation	Standard deviation
10	0.434	0.203
25	0.304	0.225
50	0.185	0.079
100	0.059	0.011
150	0.042	0.028
200	0.014	0.004



Figure S108. Average and standard deviation of normalized emission variation as a function of the distance to the TATP source.

¹H-NMR titration of AR82s with TATP in the presence of Amberlite:

An NMR titration was performed. Initially, ¹H-NMR of TATP (5 mg in 0.5 ml in CDCl₃), AR82s (3 mg in 0.5 ml in CDCl₃) and Amberlite (1.5 mg) were performed as a reference. After this, the following amounts of TATP were added to the NMR tube corresponding to AR82s to carry out the titration.

Addition number	TATP mass added (µg)	Total mass of TATP added (µg)	Addition number	TATP mass added (µg)	Total mass of TATP added (µg)
1	0	0	10	60	150
2	5	5	11	150	300
3	5	10	12	200	500
4	5	15	13	300	800
5	5	20	14	600	1400
6	10	30	15	1800	3200
7	10	40	16	3600	6800
8	20	60	17	3600	10400
9	30	90	18	5400	15800

Figure S109. Additions of TATP to the ¹H-NMR titration of AR82s.



TATP in the presence of amberlite.

pH of AR82s solution was 7 and the pH of AR82s solution with Amberlite was approximately 7, measured using litmus paper.

¹H-NMR titration of AR82s with m-CPBA

The mechanism of oxidation of AR82s in the presence of increasing amounts of peroxide was studied by NMR titration by using m-CPBA.

Addition number	m-CPBA mass added (µg)	Total mass of m-CPBA added (µg)	Addition number	m-CPBA mass added (µg)	Total mass of m-CPBA added (µg)
1	0	0	9	30	90
2	5	5	10	60	150
3	5	10	11	150	300
4	5	15	12	200	500
5	5	20	13	300	800
6	10	30	14	600	1400
7	10	40	15	1800	3200
8	20	60			

Figure S111. Amounts of m-CPBA used in NMR titration.



The titration showed oxidation of the unprotected secondary amino group with progressive disappearance of the proton signal of the amine group and chemical shift of the methylene groups of the oxidized piperidine group (the signals between 1.8 and 3.7 ppm).

Synthesis of *N*-(piperidin-4-yl)-4-[2-(piperazin-1-yl)pyrimidin-5-yl]naphthalene-1,8-dicarboxylmonoimide (AR82d)



In a 50 ml flask, 150 mg of N-(Boc-piperidin-4-yl)-4-[2-(4-Boc-piperazin-1-yl)pyrimidin-5yl]naphthalene-1,8-dicarboxylmonoimide (AR82p, 0.23 mmol) were dissolved in 15 ml of chloroform and, then, 2 ml of trifluoroacetic acid (26 mmol) were added. After 4 hours under stirring at room temperature, 10 ml of water were poured into the flask and the resulting mixture was neutralized employing a 40% sodium hydroxide solution until it reached a neutral pH. Then, 20 ml of water were added into the flask and the mixture was extracted three times with chloroform (3×30 ml). The organic extracts were combined, dried over anhydrous Na₂SO₄ and the solvent was evaporated under reduce pressure, to obtain 100 mg of AR82d, yellow solid, 98% yield. m.p.: 230 - 233°C. IR (ATR, cm⁻¹): 3448 (N-H), 3007 - 2501, 1655 (C=O), 1584, 1506, 1449, 1349, 1271, 1236, 1197, 1173 (C-N), 1116, 1041, 948, 828, 781, 720, 535, 505, 432. ¹H-NMR (300 MHz, TFA) δ (ppm): 8.72 (m, 2H, 2×H_{Ar}), 8.63 (m, 2H, 2×H_{Ar}), 8.04 (m, 1H, H_{Ar}), 7.74 (m, 2H, 2×H_{Ar}), 5.57 – 5.39 (m, 1H, CH), 4.37 (s, 4H, 2×CH₂), 3.64 (s, 6H, 3×CH₂), 3.36 – 3.09 (m, 4H, 2×CH₂), 2.05 (m, 2H, CH₂). ¹³C-NMR (75 MHz, TFA) δ (ppm): 169.9 (C=O), 169.5 (C=O), 169.2 (C=O), 168.9 (C=O), 160.0 (C_{Ar}H), 159.3 (C_{Ar}H), 155.6 (CAr), 154.9 (CAr), 139.8 (CAr), 139.2 (CAr), 137.2 (CArH), 136.5 (CArH), 135.7 (CArH), 135.1 (CArH), 134.5 (CArH), 133.1 (CAr), 132.4 (CArH), 132.3 (CArH), 132.0 (CAr), 131.9 (CArH), 131.6 (C_{Ar}H), 131.4 (C_{Ar}), 126.3 (C_{Ar}), 126.2 (C_{Ar}), 125.6 (C_{Ar}), 125.5 (C_{Ar}), 125.3 (C_{Ar}), 124.7 (CAr), 52.6 (CH), 51.9 (CH), 49.2 (CH₂), 48.6 (CH₂), 46.9 (CH₂), 46.3 (CH₂), 45.4 (CH₂), 44.7 (CH₂), 44.2 (Cq), 28.0 (CH₂), 27.3 (CH₂). HRMS (MALDI, DCTB+) m/z: calculated for $C_{25}H_{26}N_6O_2$: 443.2190 (M⁺ + H); found 443.2193. UV-Vis (CHCl₃), λ_{max} nm (log ε): 380 (3.3). Ø (EtOAc, %): 67.62. τ (375 nm, EtOAc, ns): 4.270 (χ^2 : 1.015).







Figure S115. DEPT NMR (TFA, 75 MHz) of AR82d.



Figure S116. HRMS (MALDI, DCTB+) of AR82d.

Solvatochromism:

The concentration of the compound was 20 μ M and the excitation wavelength was 360 nm.





Figure S118. Solvatochromism of AR82d under visible and 366 nm light.



Fluorescence decay lifetime (τ):

The solvent used was ethyl acetate. The concentration of the compound was 20 μ M.



Figure S119. Fluorescence decay lifetime of AR82d.

Water aceptance test:

The solvent used was MeOH and the concentration of the compound was 20 μ M.

Figure S120. Water acceptance test of AR82d under visible (up) and UV (down) light.



There was a quenching in the fluorescent emission as the amount of water was increased (ACQ).

pH test:

The amount of water that the compound could accept was 30%. The concentration of the compound was 20 μ M and the solvent used was MeOH.



Figure S121. pH test of AR82d under visible (up) and UV (down) light.



There was a quenching in the emission at very basic pH. At intermediate pH, the colour of the fluorescence was changed from green to yellow (3.4 - 7.9).

Cations and anions test:

The concentration of the compound was 20 μM and the solvent used was MeOH.

Figure S122. Cations test of AR82d under visible (up) and UV (down) light.



The most significant changes under UV light were the turn of the fluorescence to blue against Ag^+ and Au^{3+} .

The concentration of the compound was 20 μM and the solvent used was MeOH.

Figure S123. Anions test of AR82d under visible (up) and UV (down) light.



No significant changes were observed.
Oxidants and reductants test:

The solvent used was MeOH and concentration of the compound was 20 μ M.

Figure S124. Oxidants and reductants test of AR82d under visible (up) and UV (down) light.



There was a slight change against the TATP, it was changed from yellow to green.

Preliminary solvents test:

In order to determine the fluorescence variation of the compound under study in the presence of TATP, some tests were carried out for the probe in 9 different solvents. The AR82d concentration was 20 μ M and the TATP was added in excess (7 mg in each vial). All the tests were performed at room temperature and the photographs were taken immediately after the addition of TATP and again after 24 hours.

Figure S125. Pairs of AR82d (left) and AR82d with excess of TATP (right) in different solvents at time 0 h (up) and 24 h (down).



The biggest fluorescence changes were seen in DMF, MeCN and DCM:MeOH (9:1) with a emission intensity increase and in toluene a fluorescence color change was observed. The studies were also performed in CHCl₃, THF, EtOAc and DCM with a color change and in CHCl₃:MeOH (9:1) with a increase in the emission intensity, but less than the previous ones.

Work concentration test:

In order to choose an optimum concentration for the next quantitative studio, the absorbance and fluorescence of the probe was checked to be linear while concentration changes was small. The solvents used for this test were toluene, EtOAc and DCM. The excitation wavelength for all measurements was 360 nm.

Figure S126. Absorbance spectrum (up left), absorbance profile at 380 nm (up right), fluorescence spectrum (down left) and fluorescence profile at 470 nm (down right) of increasing concentrations of AR82d solution in toluene.



The ideal working concentration should stay at 0.1 of absorbance or less, to avoid inner filter effects, possible dynamic quenching or stacking processes. It was implied a concentration below 13 μ M. Taking the results into account, the chosen concentration was 1 μ M, value around which the Lambert – Beer Law (or pseudo – Lambert – Beer linear behavior for fluorescence) was fulfilled.

With m-CPBA the obtained result was the same: the concentration of the probe had to be less than 7 μ M, so a concentration of 1 μ M was adequate.

Figure S127. Absorbance spectrum (up left), absorbance profile at 360 nm (up right), fluorescence spectrum (down left) and fluorescence profile at 450 nm (down right) of increasing concentrations of AR82d solution with a m-CPBA excess (7000 μ M) in toluene.



Figure S128. Absorbance spectrum (up left), absorbance profile at 375 nm (up right), fluorescence spectrum (down left) and fluorescence profile at 475 nm (down right) of increasing concentrations of AR82d solution in EtOAc.





The obtained result was similar to the previous one. To work at an absorbance of less than 0.1, the probe concentration had to be below 12 μ M, so a good working concentration was 2.5 μ M, while with m-CPBA, the concentration should be below 6 μ M so, 1 μ M was also suitable. It was a concentration around which the Lambert – Beer Law (or pseudo – Lambert – Beer) was fulfilled.

Figure S129. Absorbance spectrum (up left), absorbance profile at 370 nm (up right), fluorescence spectrum (down left) and fluorescence profile at 485 nm (down right) of increasing concentrations of AR82d solution with a m-CPBA excess (7000 μ M) in EtOAc.



Figure S130. Absorbance spectrum (up left), absorbance profile at 380 nm (up right), fluorescence spectrum (down left) and fluorescence profile at 475 nm (down right) of increasing concentrations of AR82d solution in DCM.



The result obtained with this solvent was similar to that obtained with the two previous ones.

Figure S131. Absorbance spectrum (up left), absorbance profile at 360 nm (up right), fluorescence spectrum (down left) and fluorescence profile at 465 nm (down right) of increasing concentrations of AR82d solution with a m-CPBA excess (7000 μ M) in DCM.



Solvents test with TATP:

For all solvents, the working concentration of AR82d was 2.5 μ M, the excitation wavelenght was 360 nm and the temperature for all test was 25°C.

In the kinetic studies, TATP was in excess (20 mM) and the fluorescence emission measurements were made during 30 minutes in the case of DMF, 150 minutes in MeCN, 120 minutes in DCM, 45 minutes in the mixture DCM:MeOH (9:1) and 60 minutes for all other solvents, at 5- or 10-minutes time intervals. In the titration, the TATP concentration was between 0 and 35000 μ M and it was added directly as solid. The measurements were carried out immediately after the addition of the probe.

Figure S132. Study of AR82d with TATP in DMF. Kinetic study (up left) and profile as function as time at 390 nm and 475 nm simultaneously (up right) in presence of TATP excess. Titration (down left) and fluorescence profile at 470 nm and 595 nm simultaneously (down right) under increasing concentrations of TATP.



There was a change in the emission intensity band immediately after adding TATP. According to the fluorescence spectra, the LOD of TATP was somewhere between 15000 and 20000 μ M with an increase of the emission intensity.

Figure S133. Kinetic study at different times under visible (up) and UV (down) light in DMF. In each photo, left vial contained probe and right vial contained probe and excess TATP.



Figure S134. Titration of AR82d with excess TATP under visible (up) and UV (down) light in DMF.



Figure S135. Study of AR82d with TATP in MeCN. Kinetic study (up left) and profile as function as time at 450 nm and 570 nm simultaneously (up right) in presence of TATP excess. Titration (down left) and fluorescence profile at 450 nm and 580 nm simultaneously (down right) under increasing concentrations of TATP.



There was an increase in the emission intensity band immediately after adding TATP and it was stabilized after 140 - 160 minutes. According to the fluorescence spectra, the LOD was somewhere between 5000 and 10000 μ M with a increase of the emission intensity.

Figure S136. Kinetic study at different times under visible (up) and UV (down) light in MeCN. In each photo, left vial was contained the probe and right vial was contained the probe and an excess of TATP.

	t=0	t=60 min	t=24 h
Visible light			
UV light (366 nm)			

Figure S137. Titration of AR82d with an excess of TATP under visible (up) and UV (down) light in MeCN.

μΜ	0	250	500	1000	5000	10000	15000	20000	25000	30000	35000
Visible light								1000 M			
UV light (366 nm)											

Figure S138. Study of AR82d with TATP in toluene. Kinetic study (up left) and profile as function as time at 450 nm and 570 nm simultaneously (up right) in presence of TATP excess. Titration (down left) and fluorescence profile at 505 nm and 570 nm simultaneously (down right) under increasing concentrations of TATP.



There was a drop in the emission intensity band immediately after adding TATP. According to the fluorescence spectra, the LOD was somewhere between 0 and 250 μ M.

Figure S139. Kinetic study at different times under visible (up) and UV (down) light in toluene. In each photo, left vial was contained the probe and right vial was contained the probe and an excess of TATP.



Figure S140. Titration of AR82d with an excess of TATP under visible (up) and UV (down) light in toluene.

μΜ	0	250	500	1000	5000	10000	15000	20000	25000	30000	35000
Visible light											
UV light (366 nm)								-			

Figure S141. Study of AR82d with TATP in DCM:MeOH (9:1). Kinetic study (up left) and profile as function as time at 450 nm and 555 nm simultaneously (up right) in presence of TATP excess. Titration (down left) and fluorescence profile at 450 nm and 555 nm simultaneously (down right) under increasing concentrations of TATP.



There was an increase in the emission intensity band immediately after adding TATP. According to the fluorescence spectra, the LOD was somewhere between 5000 and 10000 μ M.

Figure S142. Kinetic study at different times under visible (up) and UV (down) light in DCM:MeOH (9:1). In each photo, left vial was contained the probe and right vial was contained the probe and an excess of TATP.

	t=0	t=60 min	t=24 h
Visible light			
UV light (366 nm)			

Figure S143. Titration of AR82d with an excess of TATP under visible (up) and UV (down) light in DCM:MeOH (9:1).

μΜ	0	250	500	1000	5000	10000	15000	20000	25000	30000	35000
Visible light		in the set									
UV light (366 nm)											

Figure S144. Study of AR82d with TATP in CHCl₃. Kinetic study (up left) and profile as function as time at 500 nm and 510 nm simultaneously (up right) in presence of TATP excess. Titration (down left) and fluorescence profile at 505 nm and 530 nm simultaneously (down right) under increasing concentrations of TATP.



There was a decrease in the emission intensity band immediately after adding TATP. According to the fluorescence spectra, the LOD was somewhere between 1000 and 5000 μ M, a high limit.

Figure S145. Kinetic study at different times under visible (up) and UV (down) light in CHCl₃. In each photo, left vial was contained the probe and right vial was contained the probe and an excess of TATP.

	t=0	t=60 min	t=24 h
Visible light			
UV light (366 nm)			

Figure S146. Titration of AR82d with an excess of TATP under visible (up) and UV (down) light in CHCl3.



Figure S147. Study of AR82d with TATP in THF. Kinetic study (up left) and profile as function as time at 400 nm and 550 nm simultaneously (up right) in presence of TATP excess. Titration (down left) and fluorescence profile at 400 nm and 550 nm simultaneously (down right) under increasing concentrations of TATP.



Related to the 550 nm band, there was a low increase in the emission intensity and a shifted to higher wavelengths immediately after adding TATP. According to the fluorescence spectra, the LOD was somewhere between 1000 and 5000 μ M, a bit high limit.

Figure S148. Kinetic study at different times under visible (up) and UV (down) light in THF. In each photo, left vial was contained the probe and right vial was contained the probe and an excess of TATP.

	t=0	t=60 min	t=24 h
Visible light			
UV light (366 nm)			

Figure S149. Titration of AR82d with an excess of TATP under visible (up) and UV (down) light in THF.

	0	250	500	1000	5000	10000	15000	20000	25000	30000	35000
Visible			Ma S	and the second			the cost	ALCOND.	the state	Ne on	to the second
light		-	-			0		_			
UV light (366 nm)								-			

Figure S150. Study of AR82d with TATP in EtOAc. Kinetic study (up left) and profile as function as time at 380 nm, 400 nm and 520 nm simultaneously (up right) in presence of TATP excess. Titration (down left) and fluorescence profile at 500 nm and 560 nm simultaneously (down right) under increasing concentrations of TATP.



There was a drop in the emission intensity and a shifted to higher wavelengths immediately after adding TATP. According to the fluorescence spectra, the LOD was somewhere between 1000 and 5000 μ M, with a displacement of the maximum emission.





Figure S152. Titration of AR82d with an excess of TATP under visible (up) and UV (down) light in EtOAc.



Figure S153. Study of AR82d with TATP in DCM. Kinetic study (up left) and profile as function as time at 430 nm and 530 nm simultaneously (up right) in presence of TATP excess. Titration (down left) and fluorescence profile at 430 nm and 525 nm simultaneously (down right) under increasing concentrations of TATP.



There was a change in the emission intensity immediately after adding TATP. The LOD was somewhere between 0 and 250 μ M, with an increase of the intensity.

Figure S154. Kinetic study at different times under visible (up) and UV (down) light in DCM. In each photo, left vial was contained the probe and right vial was contained the probe and an excess of TATP.

	t=0	t=140 min	t=24 h
Visible light			
UV light (366 nm)			

Figure S155. Titration of AR82d with an excess of TATP under visible (up) and UV (down) light in DCM.

	0	250	500	1000	5000	10000	15000	20000	25000	30000	35000
Visible light											
UV light (366 nm)							*				

Figure S156. Kinetic study (up left) and profile as function as time at 445 nm and 545 nm simultaneously (up right) in presence of TATP excess in CHCl3:MeOH (9:1).



There was an increase in the emission intensity immediately after adding TATP. It was seemed that the probe itself increased its emission in this solvent.

Figure S157. Kinetic study at different times under visible (up) and UV (down) light in CHCl₃:MeOH (9:1). In each photo, left vial was contained the probe and right vial was contained the probe and an excess of TATP.

	t=0	t=45 min	t=24 h
Visible light			
UV light (366 nm)			

Titrations in toluene and LOD calculations:

Titration of AR82d with TATP in toluene.

The working concentration of AR82d was 1 μ M and the TATP was directly added as a solid because when it was added in solution a strange and inconsistent behavior was observed. Its concentration was between 0 and 25 mM.

The measurements were carried out immediately after the addition of TATP. The excitation wavelength was 370 nm and the fluorescence changes were registered at 25°C.

Figure S158. Titration (up left), fluorescence profile at 470 nm (up right) and calibration for the LOD calculation using emission values at 470 nm (down) of a 1 μ M of AR82d solution in toluene under increasing concentrations of TATP.



A decrease in the emission intensity was observed until it was stabilized around 6000 μ M TATP. The LOD of AR82d with TATP was calculated. Calculation of the LOD by linear regression + false positive and negative at 470 nm.

For this method, the R software was used and the value of the LOD was 9.3 nM.

Finally, titration of 1 μ M AR82d was performed with increasing amounts of m-CPBA in toluene. In this case, the oxidant was added in solution and the excitation wavelength was 360 nm. The measurements were also registered at 25°C.

Figure S159. Titration (up left), fluorescence profile at 475 nm (up right) and calibration for the LOD calculation using emission values at 470 nm (down) of a 1 µM of AR82d solution in toluene under increasing concentrations of m-CPBA.



An increase in the emission intensity was observed until it was stabilized around 10000 μ M m-CPBA. The LOD of AR82d with m-CPBA was calculated

Calculation of the LOD by linear regression + false positive and negative at 475 nm.

For this method, the R software was used and the value of the LOD was 2.16 nM.

TATP gas detection in modified nanoparticles:

Several nanoparticles samples were modified by adsorption of the probe and tested as potential TATP sensors. In all three cases, 100 mg of the corresponding nanoparticles and 1 mg of the probe were used to adsorption using the method described above:

- Modified silica nanoparticles (AR82d·SiO₂).
- Modified titanium (IV) oxide nanoparticles (AR82d·TiO₂).
- Modified titanium silicon oxide nanoparticles (AR82d·SiTiO₄).

The supported probe was evaluated by several methods. In a first instance, the color variation of the nanoparticles before and after exposure to TATP was tested. Afterwards, the detection capacity was evaluated by a more quantitative approach by registering changes in fluorescence intensity (by comparing the fluorescence quantum yield for higher precision and in the position of the maximum of emission. Finally, a quantitative study for the specific detection of TATP vapors was also performed.



Figure S160. Color comparation between the nanoparticles with the probe over glass plates before and after the exposure to TATP.

The increase in fluorescence of solid was recorded by measuring the solids and determining the increase in fluorescence quantum yield (ØF). The most straightforward way to detect the presence of the analyte was the increase in fluorescence of the modified material.

The excitation wavelength for all experiments was 360 nm and the temperature for all test was 25°C.



*Figure S161. Response of fluorescence (down) and excitation (up) of AR82d adsorbed in SiO*₂ *nanoparticles.*

*Figure S162. Response of fluorescence (down) and excitation (up) of AR82d adsorbed in TiO*₂ *nanoparticles.*





Figure S163. Response of fluorescence(down) and excitation (up) of AR82d adsorbed in SiTiO4 nanoparticles.



As it could be seen in the excitation spectra, the maximum for $AR82d \cdot TiO_2$ (342 nm) was differed respecting the maximum for $AR82d \cdot SiO_2$ and $AR82d \cdot SiTiO_4$ (380 nm) and the one previously obtained for the compound in solution (360 nm). Emission measurements were made at the aforementioned wavelengths with similar results.

In doing so, it was necessary to have a fluorometer provided with an integration sphere. The accuracy of this method was checked by measuring several replicas, obtaining errors of less than 2 %.

	ØProbe+TATP	ØProbe	ØProbe+TATP/ØProbe
AR82d·SiO ₂	<2%	18.6%	0.11
AR82d·TiO2	2.75%	2.72%	1.01
AR82d·SiTiO ₄	3.15%	11.1%	0.27

Figure S164. Variation in fluorescence between the initial and after being under excess of TATP

In general, we could conclude that quantum yields followed the pattern seen in the previously shown emission intensity spectra. In the case of AR82d·SiO2 and AR82d·SiTiO4, it decreased after exposure to TATP while in the case of AR82d TiO2, it increased very slightly. However, vapor phase detection tests were not continued due to problems of solubility and subsequent adsorption onto the nanoparticles of the compound AR82d.

Synthesis of *N*-(*N*'-Boc-piperidin-4-yl)-4-[2-(4-Boc-piperazin-1-yl)pyridin-5-yl]naphthalene-1,8-dicarboxylmonoimide (AR83p)



250 mg of N-(N'-Boc-piperidin-4-yl)-4-bromonaphthalene-1,8-dicarboxylmonoimide (AR43p, 0.54 mmol), 221 mg of 2-(4-Boc-piperazin-1-yl)pyridin-5-yl boronic acid pinacol ester (0.57 mmol) and 752 mg of potassium carbonate (5.44 mmol) were dissolved in a mixture of toluene:butanol:water (4:1:2 ml) under nitrogen atmosphere and the catalyst, Pd(PPh₃)₄ (5% mmol) was added. The resulting mixture was stirred overnight at 110 °C. Afterwards, the solvent was evaporated under reduce pressure and the solid was dissolved in DCM and washed with water (3×100 ml). The organic extracts were combined, dried over anhydrous Na₂SO₄ and the solvent was evaporated under reduce pressure. The organic solid was purified by column chromatography (SiO₂, DCM:MeOH 50:1) getting 256 mg of AR83p, yellow solid, 74% yield. m.p.: 244 – 247 °C. IR (ATR, cm⁻¹): 2973 – 2841, 1693 (C=O), 1677 (C=O), 1650 (C=O), 1585, 1495, 1453 (C-O), 1408, 1348, 1235, 1159 (C-N), 1103, 1034, 995, 932, 899, 861, 777, 755, 630, 454. ¹H-NMR (300 MHz, CDCl₃) δ (ppm): 8.49 (d, J = 7.6 Hz, 2H, 2×H_{Ar}), 8.31 – 8.19 $(m, 2H, 2 \times H_{Ar}), 7.67 - 7.52 (m, 3H, 3 \times H_{Ar}), 6.76 (d, J = 8.8 Hz, 1H, H_{Ar}), 5.11 (m, 1H, CH),$ 4.24 (s, 2H, CH₂), 3.58 (m, 8H, 4×CH₂), 2.87 – 2.61 (m, 4H, 2×CH₂), 1.61 (m, 2H, CH₂), 1.44 $(d, J = 2.4 \text{ Hz}, 18\text{H}, 6 \times \text{CH}_3)$. ¹³C-NMR (75MHz, CDCl₃) δ (ppm): 164.4 (C=O), 164.2 (C=O), 158.7 (CAr), 154.7 (CAr), 148.6 (CArH), 143.6 (CAr), 138.9 (CArH), 132.0 (CArH), 131.2 (CArH), 130.9 (C_{Ar}H), 129.8 (C_{Ar}), 128.8 (C_{Ar}), 127.5 (C_{Ar}H), 126.8 (C_{Ar}H), 123.7 (C_{Ar}), 123.2 (C_{Ar}), 121.6 (C_{Ar}), 106.5 (C_{Ar}H), 80.0 (Cq), 79.4 (Cq), 51.7 (CH), 44.8 (CH₂), 43.6 (CH₂), 28.5 (CH₃), 28.4 (CH₃), 28.2 (CH₂). HRMS (MALDI, DIT-) m/z: calculated for C₃₆H₄₃N₅O₆: 641.3208 (M⁻); found 641.3232. UV-Vis (CHCl₃), λ_{max} nm (log ϵ): 390 (3.9). Ø (Hex, %): 85.40. τ (375 nm, Hex, ns): 3.199 (χ^2 : 1.153).







50 145 140 135 130 125 120 115 110 105 100 95 90 85 80 75 70 65 60 55 50 45 40 35 30 ppm





Figure S168. HRMS (MALDI, DIT-) of AR83p

Solvatochromism:

The concentration of the compound was 20 μ M and the excitation wavelength was 380 nm.





Figure S170. Solvatochromism of AR83p under visible and 366 nm light.



Fluorescence decay lifetime (τ):

The solvent used was hexane. The concentration of the compound was 20 μ M.



Figure S171. Fluorescence decay lifetime of AR83p.

Water acceptance test:

The solvent used was MeOH and the concentration of the compound was 20 μ M.

Figure S172. Water acceptance test, AR83p in MeOH under visible (up) and UV (down) light.



There was a strong increase in emission intensity from 70% water (AIE), which also coincided with what was observed using THF as solvent. In addition, using THF, there was a shift of the emission maximum from the red – purple region (580 - 650 nm) to the yellow-greenish region (550 - 570 nm).

Figure S173. Water acceptance test of AR83p in THF under visible (up) and UV (down) light.





Figure S174. Emission intensity graph (left) and graph of relationship between the emission intensity and the wavelenght of the maximum of emission (right) as a function of the amount of water added for AR83p using THF as a solvent.



pH test:

The amount of water that the compound could accept was 90%. The concentration of the compound was 20 μ M and the solvent used was MeOH.

Figure S175. pH test of AR83p under visible (up) and UV (down) light.



*HCl is included in all experiments in a close position to HNO₃ to distinguish between the redox action of HNO₃ and a possible effect due only to the concomitant acidity of nitric acid.

There was a quenching in the emission at very acidic pH, while at physiological pH, the compound precipitated.

Cations and anions test:

The concentration of the compound was 20 μ M and the solvent used was a mixture of MeOH and water (90%).

Figure S176. Cations test of AR83p under visible (up) and UV (down) light.



There was the quenching of the fluorescence in the presence of Au^{3+} under UV light. The concentration was 20 µM and the solvent used was a mixture of MeOH and water (90%). *Figure S177. Anions test of AR83p under visible (up) and UV (down) light.*



Oxidants and reductants test:

Solvent was a mixture of MeOH-water (90%) and concentration of the compound was 20 μ M. *Figure S178. Oxidants and reductants test of AR83p under visible (up) and UV (down) light.*



*HCl is included in all experiments in a close position to HNO₃ to distinguish between the redox action of HNO₃ and a possible effect due only to the concomitant acidity of nitric acid. No significant changes were observed. S104

Synthesis of *N*-(piperidin-4-yl)-4-[2-(4-Boc-piperazin-1-yl)pyridin-5-yl]naphthalene-1,8-dicarboxylmonoimide (AR83s)



150 mg of N-(piperidin-4-yl)-4-bromonaphthalene-1,8-dicarboxylmonoimide (AR43d, 0.42 mmol), 171 mg of 2-(4-Boc-piperazin-1-yl)pyridin-5-yl boronic acid pinacol ester (0.44 mmol) and 451 mg of potassium carbonate (3.26 mmol) were dissolved in a mixture of toluene:butanol:water (4:1:2 ml) under nitrogen atmosphere and the catalyst, Pd(PPh₃)₄ (5% mmol)was added. After that it was stirred overnight at 110 °C, the solvent was evaporated under reduce pressure, the solid was dissolved in CH_2Cl_2 and washed with water (3×100 ml). The organic extracts were combined, dried over anhydrous Na₂SO₄ and the solvent was evaporated under reduce pressure. The organic solid was purified by column chromatography (SiO₂, CH₂Cl₂:MeOH 50:1) getting 113 mg of AR83s, brown solid, 50% yield. m.p.: 216 - 218 °C (decomposition). IR (ATR, cm⁻¹): 3425 (N-H), 2970 – 2851, 1689 (C=O), 1585, 1405 (C-O), 1351, 1238, 1118 (C-N), 992, 929, 753, 717, 693, 535. ¹H-NMR (300 MHz, CDCl₃) δ (ppm): 8.60 (d, J = 7.5 Hz, 2H, 2×H_{Ar}), 8.30 (m, 2H, 2×H_{Ar}), 7.72 – 7.61 (m, 3H, 3×H_{Ar}), 6.80 (d, J =9.0 Hz, 1H, H_{Ar}), 5.40 – 5.25 (m, 1H, CH), 3.83 (s, 2H, CH₂), 3.69 – 3.57 (m, 8H, 4×CH₂), 3.41 -3.09 (m, 5H, 2×CH₂ y NH), 1.97 (d, J = 11.1 Hz, 2H, CH₂), 1.50 (s, 9H, 3×CH₃). ¹³C-NMR (75 MHz, CDCl₃) δ (ppm): 164.4 (C=O), 164.2 (C=O), 158.9 (C_{Ar}), 154.9 (C_{Ar}), 148.7 (C_{Ar}H), 144.1 (C_{Ar}), 139.1 (C_{Ar}H), 132.4 (C_{Ar}H), 131.7 (C_{Ar}H), 131.4 (C_{Ar}H), 130.1 (C_{Ar}), 128.9 (C_{Ar}), 127.8 (C_{Ar}H), 127.0 (C_{Ar}H), 123.8 (C_{Ar}), 123.0 (C_{Ar}), 121.5 (C_{Ar}), 106.6 (C_{Ar}H), 80.3 (Cq), 48.9 (CH), 44.9 (CH₂), 43.3 (CH₂), 28.5 (CH₃), 25.8 (CH₂). HRMS (MALDI, DIT+) m/z: calculated for C₃₁H₃₅N₅O₄: 543.2714 (M⁺ + 2); found 543.2748. UV-Vis (CHCl₃), λ_{max} nm (log ϵ): 415 (3.9). Ø (CHCl₃, %): 99.41. τ (375 nm, CHCl₃, ns): 5.291 (χ^2 : 1.040).







150 145 140 135 130 125 120 115 110 105 100 95 90 85 80 75 70 65 60 55 50 45 40 35 30 25 ppm

Figure S181. DEPT NMR (CDCl₃, 75 MHz) of AR83s.



Figure S182. HRMS (MALDI, DIT+) of AR83s.

Solvatochromism:

The concentration of the compound was 20 μ M and the excitation wavelength was 390 nm.





Figure S184. Solvatochromism of AR83s under visible and 366 nm light.


Fluorescence decay lifetime (τ):

The solvent used was chloroform. The concentration of the compound was 20 μ M.



Figure S185. Fluorescence decay lifetime of AR83s.

Water acceptance test:

The solvent used was acetone and the concentration of the compound was 20 μ M.

Figure S186. Water acceptance test, AR83s, acetone under visible (up) and UV (down) light.



There was a slight increase in emission intensity from 70% water (AIE) using both acetone and THF as solvent. At low amounts of water, the emission maximum was in the red region (about 580 nm) while, between 30% and 60% water, it was shifted to the blue region (480 - 490 nm). At high percentages of water, from 70%, the emission maximum was shifted to the green region (530 - 560 nm).

Figure S187. Water acceptance test of AR83s in THF under visible (up) and UV (down) light.





Figure S188. Emission intensity graph (left) and graph of relationship between the emission intensity and the wavelength of the maximum of emission (right) as a function of the amount of water added for AR83s using THF as a solvent.



pH test:

The amount of water that the compound could accept was 20%. The concentration of the compound was 20 μ M and the solvent used was acetone.

Figure S189. pH test of AR83s under visible (up) and UV (down) light.



No significant changes were observed.

Cations and anions test:

The concentration of the compound was 20 μM and the solvent used was acetone.

Figure S190. Cations test of AR83s under visible (up) and UV (down) light.



The fluorescence increased with Ag^+ and Au^{3+} . Concentration, 20 μ M, solvent, acetone. *Figure S191. Anions test of AR83s under visible (up) and UV (down) light.*



Oxidants and reductants test:

The solvent used was a mixture of MeOH-water and concentration of compound was 20 µM. *Figure S192. Oxidants and reductants test of AR83p under visible (up) and UV (down) light.*



*HCl is included in all experiments in a close position to HNO₃ to distinguish between the redox action of HNO₃ and a possible effect due only to the concomitant acidity of nitric acid. The most important change was the change in fluorescence from orange to blue with m-CPBA.

Synthesis of *N*-(piperidin-4-yl)-4-[2-(piperazin-1-yl)pyridin-5-yl]naphthalene-1,8dicarboxylmonoimide (AR83d)



150 mg of N-(N'-Boc-piperidin-4-yl)-4-[2-(4-Boc-piperazin-1-yl)pyridin-5-yl]naphthalene-1,8-dicarboxylmonoimide (AR83p, 0.23 mmol) were dissolved in 15 ml of chloroform and, then, 2 ml of trifluoroacetic acid (26 mmol) were added. The mixture was stirred 4 hours at room temperature. Afterwards, 10 ml of water were poured into the flask and the mixture was neutralized employing a 40% sodium hydroxide solution until it reached a neutral pH. Then, 20 ml of water were added into the flask and the mixture was extracted with chloroform (3×30 ml). The organic extracts were combined, dried over anhydrous Na₂SO₄ and the solvent was evaporated under reduce pressure, to obtain 100 mg of AR83d, yellow solid, 99% yield. m.p.: 274 – 277 °C. IR (ATR, cm⁻¹): 3236 (N-H), 2925 – 2737, 1693 (C=O), 1650 (C=O), 1579, 1495, 1348, 1235, 1151 (C-N), 1105, 782, 755, 459. ¹H-NMR (300 MHz, CDCl₃) δ (ppm): 8.60 (d, J = 7.6 Hz, 2H, $2 \times H_{Ar}$), 8.34 – 8.29 (m, 2H, $2 \times H_{Ar}$), 7.73 – 7.63 (m, 3H, $3 \times H_{Ar}$), 6.81 (d, J = 8.8Hz, 1H, H_{Ar}), 5.25 – 5.16 (m, 1H, CH), 3.65 (m, 4H, 2×CH₂), 3.27 (m, 2H, CH₂), 3.04 (m, 4H, 2×CH₂), 2.86 – 2.65 (m, 4H, 2×CH₂), 2.01 (s, 2H, 2×NH), 1.72 (m, 2H, CH₂). ¹³C-NMR (75 MHz, CDCl₃) δ (ppm): 164.7 (C=O), 164.6 (C=O), 159.4 (C_{Ar}), 148.8 (C_{Ar}H), 143.9 (C_{Ar}), 138.9 (CArH), 132.2 (CArH), 131.3 (CArH), 131.1 (CArH), 130.1 (CAr), 129.0 (CAr), 127.7 (CArH), 126.9 (CArH), 123.6 (CAr), 123.5 (CAr), 121.9 (CAr), 106.5 (CArH), 52.1 (CH), 47.0 (CH₂), 46.3 (CH₂), 46.1 (CH₂), 30.2 (CH₂). HRMS (MALDI, DCTB+) m/z: calculated for $C_{26}H_{27}N_5O_2$: 442.2238 (M⁺ + H); found 442.2233. UV-Vis (CHCl₃), λ_{max} nm (log ε): 350 (3.3). Ø (MeOH, %): <3. τ (350 nm, MeOH, ns): 1.991/4.406 (χ^2 : 1.116).



Figure S194. ¹³C-NMR (CDCl₃, 75 MHz) of AR83d.



Figure S196. HRMS (MALDI, DCTB+) of AR83d.

Solvatochromism:

The concentration of the compound was 20 μ M and the excitation wavelength was 350 nm.





Figure S198. Solvatochromism of AR83d under visible and 366 nm light



Fluorescence decay lifetime (τ):

The solvent used was MeOH and the concentration of the compound was 20 μ M.



Figure S199. Fluorescence decay lifetime of AR83d.

Water acceptance test:

The solvent used was MeOH and the concentration of the compound was 20 μ M.

Figure S200. Water acceptance test of AR83d under visible (up) and UV (down) light.



The fluorescent emission was quenched as the amount of water increased (ACQ).

pH test:

The amount of water for the performance of this test was 90%. The concentration of the compound was 20 μ M and the solvent used was MeOH.

Figure S201. pH	test of AR83d	under visible (u	(p) and UV	(down) light.
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	R	HCl	3.4	4.8	6.4	7.4	7.9	10.5	КОН
Visible light				*					



There was a slight increase in the fluorescence intensity at intermediate pHs.

Cations and anions test:

The concentration of the compound was 20 μM and the solvent used was a mixture of MeOH and water (90%).



Figure S202. Cations test of AR83d under visible (up) and UV (down) light.

The most significant changes under UV light were the increase in the fluorescence emission when the probe was exposed to Ag^+ and Au^{3+} .

The concentration of the compound was 20 μM and the solvent used was a mixture of MeOH and water (90%).



Figure S203. Anions test of AR83d under visible (up) and UV (down) light.

No significant changes were observed.

Oxidants and reductants test:

The solvent used was a mixture of MeOH and water (90%) and concentration of the compound was 20 μ M.



Figure S204. Oxidants and reductants test of AR83d under visible (up) and UV (down) light.

*HCl is included in all experiments in a close position to HNO₃ to distinguish between the redox action of HNO₃ and a possible effect due only to the concomitant acidity of nitric acid. No significant changes were observed.

Preliminary solvents test:

In order to determine the fluorescence variation of the compound under study in the presence of TATP, some test were carried out for the probe in 9 different solvents. The AR83d concentration was 25 μ M and the TATP was added in excess (7 mg in each vial). All the tests were performed at room temperature and the photographs were taken immediately after the addition of TATP and again after 24 hours.

Figure S205. Pairs of AR83d (left) and AR83d with excess of TATP (right) in different solvents at time 0 h (up) and 24 h (down).



The most noticeable fluorescence changes were seen in DMF, MeCN, CHCl₃:MeOH (9:1) and DCM:MeOH (9:1).

Solvents test with TATP:

For all solvents, the working concentration of AR83d was 2.5 μ M, the excitation wavelength was 350 nm and the temperature for all tests was 25°C.

In the kinetic studies, TATP was in excess (20 mM) and the fluorescence emission measurements were made during 40 minutes, at 5 minutes time intervals. In the titration, the TATP concentration was between 0 and 35000 μ M and it was added directly as solid. The measurements were carried out immediately after the addition of the probe.

Figure S206. Study of AR83d with TATP in DMF. Kinetic study (up left) and profile as function as time at 390 nm, 460 nm and 550 nm simultaneously (up right) in presence of TATP excess. Titration (down left) and fluorescence profile at 390 nm, 465 nm and 585 nm simultaneously (down right) under increasing concentrations of TATP.



There was an increase in the emission intensity band immediately after adding TATP. The LOD of TATP was somewhere between 25000 and 30000 μ M with the appearance of a new maximum peak.

Figure S207. Kinetic study at different times under visible (up) and UV (down) light in DMF. In each photo, left vial was contained the probe and right vial was contained the probe and an excess of TATP.

	t=0	t=60 min	t=48 h
Visible light			
UV light (366 nm)			

Figure S208. Titration of AR83d with an excess of TATP under visible (up) and UV (down) light in DCM.

μΜ	0	250	500	1000	5000	10000	15000	20000	25000	30000 35000
Visible light										
UV light (366 nm)										

Figure S209. Study of AR83d with TATP in MeCN. Kinetic study (up left) and profile as function as time at 390 nm and 450 nm simultaneously (up right) in presence of TATP excess. Titration (down left) and fluorescence profile at 390 nm, 450 nm and 570 nm simultaneusly (down right) under increasing concentrations of TATP.



There was a increase change in the emission intensity band immediately after adding TATP. According to the spectra, the LOD of TATP was somewhere between 10000 and 15000 μ M, a fairly high limit, with the appearance of a new emission peak.

Figure S210. Kinetic study at different times under visible (up) and UV (down) light in MeCN. In each photo, left vial contained probe and right vial contained probe and excess TATP.



Figure S211. Titration of AR83d with an excess of TATP under visible (up) and UV (down) light in MeCN.



Figure S212. Study of AR83d with TATP in CHCl₃. Kinetic study (up left) and profile as function as time at 455 nm and 545 nm simultaneously (up right) in presence of TATP excess. Titration (down left) and fluorescence profile at 465 nm and 540 nm simultaneusly (down right) under increasing concentrations of TATP.



There was a decrease change in the emission intensity band immediately after adding TATP. The LOD of TATP was somewhere between 1000 and 5000 μ M with the dissappearance of a maximum emission.

Figure S213. Kinetic study at different times under visible (up) and UV (down) light in CHCl3. In each photo, left vial was contained the probe and right vial was contained the probe and an excess of TATP.

	t=0	t=60 min	t=24 h
Visible light			
UV light (366 nm)			

*Figure S214. Titration of AR83d with an excess of TATP under visible (up) and UV (down) light in CHCl*₃.

μΜ	0	250	500	1000	5000	10000	15000	20000	25000	30000	35000
Visible light											and the state
UV light (366 nm)											

Figure S215. Study of AR83d with TATP in CHCl₃:MeOH (9:1). Kinetic study (up left) and profile as function as time at 450 nm and 550 nm simultaneously (up right) in presence of TATP excess. Titration (down left) and fluorescence profile at 450 nm and 550 nm simultaneously (down right) under increasing concentrations of TATP.



There was an increase in the emission intensity band immediately after adding TATP. According to the spectra, the LOD of TATP was somewhere between 5000 and 10000 μ M.





Figure S217. Titration of AR83s with an excess of TATP under visible (up) and UV (down) light in CHCl₃:MeOH (9:1).



Figure S218. Study of AR83d with TATP in DCM:MeOH (9:1). Kinetic study (up left) and profile as function as time at 450 nm and 550 nm simultaneously (up right) in presence of TATP excess. Titration (down left) and fluorescence profile at 415 nm and 560 nm simultaneously (down right) under increasing concentrations of TATP.



There was a increase change in the emission intensity band immediately after adding TATP. According to the fluorescence spectra, the LOD of TATP was somewhere between 1000 and 5000 μ M.

Figure S219. Kinetic study at different times under visible (up) and UV (down) light in DCM:MeOH (9:1). In each photo, left vial was contained the probe and right vial was contained the probe and an excess of TATP.

	t=0	t=30 min	t=24 h
Visible light			
UV light (366 nm)			

Figure S220. Titration of AR83d with an excess of TATP under visible (up) and UV (down) light in DCM:MeOH (9:1).

μΜ	0	250	500	1000	5000	10000	15000	20000	25000	30000	35000
Visible light											
UV light (366 nm)											

Because of the low sensitivity the study was discontinued.

Synthesis of *N*-(*N*'-Boc-piperidin-4-yl)-4-[(4-Boc-piperazin-1-yl)phenyl]naphthalene-1,8dicarboxylmonoimide (AR90p)



250 mg of N-(N'-Boc-piperidin-4-yl)-4-bromonaphthalene-1,8-dicarboxylmonoimide (AR43p, 0.54 mmol), 220 mg of 4-(4-Boc-piperazin-1-yl)phenylboronic acid pinacol ester (0.57 mmol) and 752 mg of potassium carbonate (5.44 mmol) were dissolved in a mixture of toluene:butanol:water (4:1:2 ml) under nitrogen atmosphere and, then, the catalyst, Pd(PPh₃)₄ (5% mmol) was added. The resulting mixture was stirred overnight at 110 °C. After that, the solvent was evaporated under reduce pressure, the solid was dissolved in CH₂Cl₂ and washed with water (3×100 ml). The organic extracts were combined, dried over anhydrous Na₂SO₄ and the solvent was evaporated under reduce pressure. The organic solid was purified by column chromatography (SiO₂, CH₂Cl₂:MeOH) getting 242 mg of AR90p as a yellow solid, 70% yield. m.p.: 250 – 252°C. IR (ATR, cm⁻¹): 3006 – 2832, 1686 (C=O), 1650 (C=O), 1582, 1507, 1420 (C-O), 1351, 1232, 1159 (C-N), 1124, 1097, 1037, 977, 915, 902, 861, 812, 780, 755, 663, 639, 531, 457. ¹H-NMR (300 MHz, CDCl₃) δ (ppm): 8.58 (d, J = 7.7 Hz, 2H, 2×H_{Ar}), 8.32 (d, J =8.5 Hz, 1H, H_{Ar}), 7.67 (t, J = 8.0 Hz, 2H, 2×H_{Ar}), 7.42 (d, J = 8.5 Hz, 2H, 2×H_{Ar}), 7.07 (d, J =8.5 Hz, 2H, 2×H_{Ar}), 5.20 (m, 1H, CH), 4.30 (s, 2H, CH₂), 3.64 (m, 4H, 2×CH₂), 3.27 (m, 4H, 2×CH₂), 2.94 – 2.69 (m, 4H, 2×CH₂), 1.68 (m, 2H, CH₂), 1.49 (s, 18H, 6×CH₃). ¹³C-NMR (75MHz, CDCl₃) δ (ppm): 164.8 (C=O), 164.6 (C=O), 154.8 (C=O), 154.7 (C=O), 151.2 (C_{Ar}), 146.8 (CAr), 132.8 (CArH), 131.2 (CAr), 131.1 (CAr), 131.0 (CArH), 130.1 (CAr), 129.0 (CAr), 127.7 (C_{Ar}H), 126.8 (C_{Ar}H), 123.3 (C_{Ar}), 121.5 (C_{Ar}), 116.1 (C_{Ar}H), 80.2 (Cq), 79.6 (Cq), 51.9 (CH), 48.9 (CH₂), 43.8 (CH₂), 28.6 (CH₃), 28.5 (CH₃), 28.4 (CH₂). HRMS (MALDI, DIT-) m/z: calculated for C₃₇H₄₄N₄O₆: 640.3255 (M⁻); found 640.3261. Ø (MCH, %): 93.19. τ (375 nm, MCH, ns): 3.542 (χ^2 : 0.975).



165 160 155 150 145 140 135 130 125 120 115 110 105 100 95 90 85 80 75 70 65 60 55 50 45 40 35 30 ppm









Figure S224. HRMS (MALDI, DIT+) of AR90p

Solvatochromism:

The concentration of the compound was 20 μ M and the excitation wavelength was 380 nm.





Figure S226. Solvatochromism of AR90p under visible and 366 nm light.



Fluorescence decay lifetime (τ):

The solvent used was MCH. The concentration of the compound was 20 μ M.



Figure S227. Fluorescence decay lifetime of AR90p.

Water acceptance test:

The solvent used was THF and the concentration of the compound was 20 μ M.

Figure S228. Water acceptance test of AR90p under visible (up) and UV (down) light.



There was an increase in emission intensity from 60% water (AIE). At low amount of water, the maximum emission was shifted from red region to yellow greenish region.

pH test:

The amount of water for the performance of this test were 10% and 60%. The concentration of the compound was 20 μM and the solvent used was THF.

Figure S229. pH test of AR90p at 10% of water under visible (up) and UV (down) light.



Figure S230. pH test of AR90p at 60% of water under visible (up) and UV (down) light.

	R	HCl	3.4	4.8	6.4	7.4	7.9	10.5	КОН
Visible light									
UV light (366 nm)									

The most important change was the response to highly acidic media in both amounts of water. **Cations and anions test:**

The concentration of the compound was 20 μ M and the solvent used was THF.

Figure S231. Cations test of AR90p under visible (up) and UV (down) light.

	$R \qquad Ag^{+} Ni^{2+} Sn^{2+} Cd^{2+} Zn^{2+} Pb^{2+} Cu^{2+} Fe^{3+} Sc^{3+} Al^{3+} Hg^{2+} Au^{3+} Co^{2+} Pd^{2+} Ir^{3+} Cu^{+} Ru^{3+} Pt^{2+} Ru^{3+} Pt$
Visible light	



The only change observed was the change in fluorescence from red to green in the presence of Au^{3+} .

The concentration of the compound was 20 μ M and the solvent used was THF.

Figure S232. Anions test of AR90p under visible (up) and UV (down) light.



No significant changes were observed.

Oxidants and reductants test:

The solvent used was THF and concentration of the compound was 20 μ M.

Figure S233. Oxidants and reductants test of AR90p under visible (up) and UV (down) light.



*HCl is included in all experiments in a close position to HNO₃ to distinguish between the redox action of HNO₃ and a possible effect due only to the concomitant acidity of nitric acid. No significant changes were observed.

Synthesis of *N*-(piperidin-4-yl)-4-[(4-Boc-piperazin-1-yl)phenyl]naphthalene-1,8dicarboxylmonoimide (AR90s)



150 mg of N-(piperidin-4-yl)-4-bromonaphthalene-1,8-dicarboxylmonoimide (AR43d, 0.42 mmol), 179 mg of 4-(4-Boc-piperazin-1-yl)phenylboronic acid pinacol ester (0.44 mmol) and 451 mg of potassium carbonate (3.26 mmol) were dissolved in a mixture of toluene:nbutanol:water (4:1:2 ml) under nitrogen atmosphere and, then, the catalyst, Pd(PPh₃)₄ (5% mmol) was added and the resulting mixture was stirred overnight at 110°C. After that, the solvent was evaporated under reduce pressure, the solid was dissolved in CH₂Cl₂ and washed with water (3×100 ml). The organic extracts were combined, dried over anhydrous Na₂SO₄ and the solvent was evaporated under reduce pressure. The organic solid was purified by column chromatography (SiO₂, CH₂Cl₂:MeOH) obtaining 190 mg of AR90s as a brown solid, 84% yield. m.p.: 188 – 190°C. IR (ATR, cm⁻¹): 3407 (N-H), 3054 – 2851, 1689 (C=O), 1590, 1489, 1399, 1363, 1235, 1159 (C-N), 1118, 992, 929, 809, 750, 717, 693, 540. ¹H-NMR (300 MHz, CDCl₃) δ (ppm): 8.59 (d, J = 7.5 Hz, 2H, 2×H_{Ar}), 8.33 (dd, J = 8.5, 0.9 Hz, 1H, H_{Ar}), 7.68 (t, J= 8.0 Hz, 2H, 2×H_{Ar}), 7.43 (d, J = 8.6 Hz, 2H, 2×H_{Ar}), 7.07 (d, J = 8.7 Hz, 2H, 2×H_{Ar}), 5.21 (m, 1H, CH), 3.64 (m, 4H, 2×CH₂), 3.27 (m, 6H, 3×CH₂), 2.86 – 2.68 (m, 4H, 2×CH₂), 2.14 (s, 1H, NH), 1.73 (m, 2H, CH₂), 1.50 (s, 9H, 3×CH₃). ¹³C-NMR (75MHz, CDCl₃) δ (ppm): 164.6 (C=O), 164.4 (C=O), 154.8 (C_{Ar}), 151.3 (C_{Ar}), 147.1 (C_{Ar}), 133.1 (C_{Ar}H), 131.5 (C_{Ar}H), 131.4 (C_{Ar}H), 131.0 (C_{Ar}H), 130.1 (C_{Ar}), 129.9 (C_{Ar}), 129.0 (C_{Ar}), 127.8 (C_{Ar}H), 126.8 (C_{Ar}H), 122.9 (C_{Ar}), 121.1 (C_{Ar}), 116.1 (C_{Ar}), 80.2 (Cq), 48.8 (CH₂), 44.7 (CH₂), 43.6 (CH₂), 28.6 (CH₃), 25.8 (CH₂). UV-Vis (CHCl₃), λ_{max} nm (log ε): 310 (4.0). HRMS (MALDI, DIT+) m/z: calculated for C₃₂H₃₆N₄O₄+H: 541.2815 (M⁺+H); found 541.2807. Ø (THF, %): 3.07. τ (280 nm, THF, ns): 1.786/6.278 (χ²: 1.114).



Figure S235. ¹³C-NMR (CDCl₃, 75 MHz) of AR90s.



Figure S236. DEPT NMR (CDCl₃, 75 MHz) of AR90s.



Figure S237. HRMS (MALDI, DIT+) of AR90s

Fluorescence decay lifetime (τ):

The solvent used was THF. The concentration of the compound was 20 μ M.



Figure S238. Fluorescence decay lifetime of AR90s.

Solvatochromism:

The concentration of the compound was 20 μ M and the excitation wavelength was 310 nm.

Figure S239. Solvatochromism of AR90s.

Figure S240. Solvatochromism of AR90s under visible and 366 nm light

Water acceptance test:

The solvent used was MeOH and the concentration of the compound was 20 μ M.

Figure S241. Water acceptance test of AR90s under visible (up) and UV (down) light.

The maximum amount of water that was admitted by the product was 40%. From 60% water, the fluorescence was quenched (ACQ).

pH test:

The amount of water for the performance of this test was 40%. The concentration of the compound was 20 μ M and the solvent used was MeOH.

Figure S242. pH test of AR90s under visible (up) and UV (down) light.

The most important change in fluorescence was the response to highly acidic media.

Cations and anions test:

The concentration of the compound was 20 μM and the solvent used was MeOH.

Figure S243. Cations test of AR90s under visible (up) and UV (down) light.

There was fluorescence quenching in the presence of Fe³⁺, Au³⁺, Pd²⁺ and Ru³⁺. The concentration of the compound was 20 µM and the solvent used was MeOH. *Figure S244. Anions test of AR90s under visible (up) and UV (down) light.*

	R	F	Cŀ	Br ⁻	I-	BzO-	NO3 ⁻	H ₂ PO ₄ - H	ISO4 ⁻ A	AcO-	CN-	SCN-
Visible light												

No significant changes were observed.

Oxidants and reductants test:

The solvent used was THF and concentration of the compound was 20 µM.

Figure S245. Oxidants and reductants test of AR90s under visible (up) and UV (down) light.

	R	HCl* HNO ₃ Oxn Hz H ₂ O ₂	R m-CPBA TNB TNT	R m-CPBA TATP HMTD
Visible light				
	K	and the second second		Charles and
UV light (366 nm)				

*HCl is included in all experiments in a close position to HNO₃ to distinguish between the redox action of HNO₃ and a possible effect due only to the concomitant acidity of nitric acid.

The most significant change was the fluorescence quenching when exposed to TATP. In addition, in front of HMTD the color of the fluorescence became turquoise blue.

Preliminary solvents test:

In order to determine the fluorescence variation of the compound under study in the presence of TATP, some test were carried out for the probe in 9 different solvents. The AR90s concentration was 20 μ M and the TATP was added in excess (7 mg in each vial). All the test were performed at room temperature and the photographs were taken immediately after the addition of TATP and again after 72 hours.

Figure S246. Pairs of AR90s (left) and AR90s with excess of TATP (right) in different solvents at time 0 h (up) and 72 h (down).

The most noticeable fluorescence changes were seen in EtOAc, MeCN, CHCl₃, DCM, CHCl₃:MeOH (9:1) and DCM:MeOH (9:1) so kinetic studies and TATP titrations were carried out in these solvents.

Solvents test with TATP:

For all solvents, the working concentration of AR90s was 2.5 μ M, the excitation wavelenght was 310 nm and the temperature for all test was 25°C.

In the kinetic studies, TATP was in excess (20 mM) and the fluorescence emission measurements were made during the subsequent minutes at 5 min, 10 min and 30 min time intervals. In the titration, the TATP concentration was between 0 and 35000 μ M and it was added directly as solid. The measurements were carried out inmediately after the adittion of the probe.

Figure S247. Study of AR90s with TATP in MeCN. Kinetic study (up left) and profile as function as time at 420 nm (up right) in presence of TATP excess. Titration (down left) and fluorescence profile at 415 nm and 560 nm simultaneously (down right) under increasing concentrations of TATP.

There was a decrease change in the emission intensity band immediately after adding TATP. The LOD of TATP was somewhere between 5000 and 10000 μ M, a fairly high limit, with a decrease in emission intensity.

Figure S248. Kinetic study at different times under visible (up) and UV (down) light in MeCN. In each photo, left vial was contained the probe and right vial was contained the probe and an excess of TATP.

	t=0	t=40 min	t=24 h		
Visible light					
UV light (366 nm)					

Figure S249. Titration of AR90s with an excess of TATP under visible (up) and UV (down) light in MeCN.

μΜ	0	250	500	1000	5000	10000	15000	20000	25000	30000	35000
Visible light											
UV light (366 nm)											

Figure S250. Study of AR90s with TATP in DCM. Kinetic study (up left) and profile as function as time at 415 nm and 550 nm simultaneously (up right) in presence of TATP excess.

Titration (down left) and fluorescence profile at 415 nm and 560 nm simultaneously (down right) under increasing concentrations of TATP.

There was a decrease in the emission intensity band immediately after adding TATP. The LOD of TATP was somewhere between 1000 and 5000 μ M, slightly less than in the previous solvent, but still too high a range, with a decrease in emission intensity.

Figure S252. Titration of AR90s with an excess of TATP under visible (up) and UV (down) light in DCM.

Figure S253. Study of AR90s with TATP in CHCl₃. Kinetic study (up left) and profile as function as time at 420 nm and 545 nm simultaneously (up right) in presence of TATP excess. Titration (down left) and fluorescence profile at 420 nm and 545 nm simultaneously (down right) under increasing concentrations of TATP.

There was a decrease in the emission intensity band immediately after adding TATP. According to the fluorescence spectra, the LOD of TATP was somewhere between 1000 and 5000 μ M, as in the previous solvent, with a strong decrease in emission intensity.
Figure S254. Kinetic study at different times under visible (up) and UV (down) light, CHCl₃. In each photo, left vial contained probe and right vial contained probe and excess TATP.



Figure S255. Titration of AR90s with an excess of TATP under visible (up) and UV (down) light in CHCl₃.



Figure S256. Study of AR90s with TATP in EtOAc. Kinetic study (up left) and profile as function as time at 355 nm and 405 nm simultaneously (up right) in presence of TATP excess. Titration (down left) and fluorescence profile at 355 nm and 405 nm simultaneously (down right) under increasing concentrations of TATP.





There was a decrease in emission intensity as time elapsed until stabilization was reached at around 60 minutes. According to the fluorescence spectra, the LOD was somewhere between 1000 and 5000 μ M with a small decrease in emission intensity.

Figure S257. Kinetic study at different times under visible (up) and UV (down) light in EtOAc. In each photo, left vial was contained the probe and right vial was contained the probe and an excess of TATP.



Figure S258. Titration of AR90s with an excess of TATP under visible (up) and UV (down) light in EtOAc.



Figure S259. Study of AR90s with TATP in CHCl₃:MeOH (9:1). Kinetic study (up left) and profile as function as time at 445 nm and 555 nm simultaneously (up right) in presence of TATP excess. Titration (down left) and fluorescence profile at 425 nm and 560 nm simultaneously (down right) under increasing concentrations of TATP.



There was a decrease in the emission intensity band immediately after adding TATP but, in the 445 nm band, an increase in the intensity was observed. According to the fluorescence spectra, the LOD of TATP was somewhere between 1000 and 5000 μ M with an increase in emission intensity in the 560 nm band stronger than the decrease in the 425 nm band.

*Figure S260. Kinetic study at different times under visible (up) and UV (down) light in CHCl*₃:*MeOH (9:1). In each photo, left vial contained probe and right vial contained probe and an excess of TATP.*

	t=0	t=60 min	t=72 h
Visible light			
UV light (366 nm)			

*Figure S261. Titration of AR90s with an excess of TATP under visible (up) and UV (down) light in CHCl*₃:*MeOH (9:1).*

μΜ	0	250	500	1000	5000	10000	15000	20000	25000	30000	35000
Visible light					-						
UV light (366 nm)											

Because of the low sensitivity the study was discontinued.

Synthesis of *N*-(piperidin-4-yl)-4-[(4-piperazin-1-yl)phenyl]naphthalene-1,8dicarboxylmonoimide (AR90d)



150 of N-(4-Boc-piperidin-1-yl)-4-[(4-Boc-piperazin-1-yl)phenyl]naphthalene-1,8mg dicarboxylmonoimide (AR90p, 0.34 mmol) were dissolved in 15 ml of chloroform and, then, 2 ml of trifluoroacetic acid (26 mmol) was added. The mixture was stirred for 3 hours at room temperature. After that, 10 ml of water were poured into the flask and the mixture was neutralized employing a 40 % sodium hydroxide solution until it reached a neutral pH. Then, 20 ml of water were added into the flask and the mixture was extracted three times with chloroform (3×30 ml). The organic extracts were combined, dried over anhydrous Na₂SO₄ and the solvent was evaporated under reduce pressure, to obtain 148 mg of AR90d as a yellow solid, 99% yield. m.p.: 285 - 288°C. IR (ATR, cm⁻¹): 3422 (N-H), 2940 - 2827, 1696 (C=O), 1650 (C=O), 1585, 1507, 1396, 1351, 1232, 1154 (C-N), 1115, 912, 777, 755, 583. ¹H-NMR (300 MHz, CDCl₃) δ (ppm): 8.59 (d, J = 7.5 Hz, 2H, 2×H_{Ar}), 8.34 (dd, J = 8.5, 1.1 Hz, 1H, H_{Ar}), 7.66 (m, 2H, $2 \times H_{Ar}$), 7.42 (d, J = 8.8 Hz, 2H, $2 \times H_{Ar}$), 7.07 (d, J = 8.8 Hz, 2H, $2 \times H_{Ar}$), 5.20 (m, 1H, CH), 3.32 – 3.22 (m, 6H, 2×CH₂ + 2×NH), 3.07 – 3.10 (m, 4H, 2×CH₂), 2.81 – 2.68 (m, 4H, 2×CH₂), 1.72 (m, 4H, 2×CH₂). ¹³C-NMR (75MHz, CDCl₃) δ (ppm): 164.9 (C=O), 164.7 (C=O), 151.9 (C_{Ar}), 146.9 (C_{Ar}), 132.8 (C_{Ar}H), 131.2 (C_{Ar}H), 131.1 (C_{Ar}H), 131.0 (C_{Ar}H), 130.1 (CAr), 129.5 (CAr), 129.0 (CAr), 127.7 (CArH), 126.7 (CArH), 123.4 (CAr), 121.5 (CAr), 115.6 (C_{Ar}H), 52.1 (CH), 49.9 (CH₂), 47.1 (CH₂), 46.2 (CH₂), 30.3 (CH₂). UV-Vis (CHCl₃), λ_{max} nm (log ε): 390 (3.6). HRMS (MALDI, DIT-) m/z: calculated for C₂₇H₂₈N₄O₂+H: 441.2285 (M^++H) ; found 441.2306. Ø (Toluene, %): 96.12. τ (375 nm, Toluene, ns): 5.278 (χ^2 : 1.025).



Figure S263. ¹³C-NMR (CDCl₃, 75 MHz) of AR90d.



450

439.2157

440

0.0 -

453.2316

465 m/z

460

Figure S265. HRMS (MALDI, DIT+) of AR90d.

Fluorescence decay lifetime (τ):

The solvent used was toluene. The concentration of the compound was 20 μ M.



Figure S266. Fluorescence decay lifetime of AR90d.

Solvatochromism:

The concentration of the compound was 20 μ M and the excitation wavelength was 385 nm.







Figure S268. Solvatochromism of AR90d under visible and 366 nm light



Water acceptance test:

The solvent used was MeOH and the concentration of the compound was 20 μ M.

Figure S269. Water acceptance test of AR90d under visible (up) and UV (down) light.



There was no increase in emission intensity.

pH test:

The amount of water for the performance of this test was 20%. The concentration of the compound was 20 μ M and the solvent used was MeOH.



Figure S270. pH test of AR90d under visible (up) and UV (down) light.

The most important change was the fluorescence quenching at very basic pH.

Cations and anions test:

The concentration of the compound was 20 μ M and the solvent used was MeOH.

Figure S271. Cations test of AR90d under visible (up) and UV (down) light.



The most significant change was the green color acquired by the fluorescence when was confronted with Au^{3+} .

The concentration of the compound was 20 μ M and the solvent used was MeOH.

Figure S272. Anions test of AR90d under visible (up) and UV (down) light.





No significant changes were observed.

Oxidants and reductants test:

The solvent used was THF and concentration of the compound was 20 µM.

Figure S273. Oxidants and reductants test of AR90d under visible (up) and UV (down) light.

	R	HCl* HNO ₃ Oxn Hz H ₂ O ₂	R m-CPBA TNB TNT	R m-CPBA TATP HMTD
Visible light				
0		the the the state of the state	Et to the	
UV light (366 nm)				

*HCl is included in all experiments in a close position to HNO₃ to distinguish between the redox action of HNO₃ and a possible effect due only to the concomitant acidity of nitric acid. No significant changes were observed except for an increase in fluorescence with m-CPBA.

Preliminary solvents test:

In order to determine the fluorescence variation of the compound under study in the presence of TATP, some tests were carried out for the probe in 9 different solvents. The AR90d concentration was 20 μ M and the TATP was added in excess (7 mg in each vial). All the tests were performed at room temperature and the photographs were taken immediately after the addition of TATP and again after 72 hours.





The biggest fluorescence changes were seen in CHCl₃, toluene and CHCl₃:MeOH (9:1) so these were the chosen solvents to perform the kinetic studies and TATP titrations experiments.

Solvents test with TATP:

For all solvents, the working concentration of AR90d was 2.5 μ M, the excitation wavelength was 385 nm and the temperature for all test was 25°C.

In the kinetic studies, TATP was in excess (20 mM) and the fluorescence emission measurements were made during the subsequent minutes at 5 min, 10 min and 30 min time intervals. In the titration, the TATP concentration was between 0 and 35000 μ M and it was added directly as solid. The measurements were carried out immediately after the addition of the probe.

Figure S275. Study of AR90d with TATP in CHCl₃. Kinetic study (up left) and profile as function as time at 555 nm (up right) in presence of TATP excess. Titration (down left) and fluorescence profile at 550 nm and 600 nm simultaneusly (down right) under increasing concentrations of TATP.



There was a change in the emission intensity band immediately after adding TATP. According to the fluorescence spectra, the LOD was somewhere between 1000 and 5000 μ M.

Figure S276. Kinetic study at different times under visible (up) and UV (down) light in CHCl3. In each photo, left vial was contained the probe and right vial was contained the probe and an excess of TATP.

	t=0	t=60 min	t=24 h
Visible light			
UV light (366 nm)			

*Figure S277. Titration of AR90d with an excess of TATP under visible (up) and UV (down) light in CHCl*₃.

μΜ	0	250	500	1000	5000	10000	15000	20000	25000	30000	35000
Visible light										li li	li .
UV light (366 nm)											

Figure S278. Study of AR90d with TATP in toluene. Kinetic study (up left) and profile as function as time at 555 nm (up right) in presence of TATP excess. Titration (down left) and fluorescence profile at 535 nm (down right) under increasing concentrations of TATP.



There was a marked decrease change in the emission intensity band immediately after adding TATP. According to the fluorescence spectra, the LOD was somewhere between 1000 and 5000 μ M with a decrease of the intensity.





Figure S280. Titration of AR90d with an excess of TATP under visible (up) and UV (down) light in toluene.



Figure S281. Study of AR90d with TATP in CHCl₃:MeOH (9:1). Kinetic study (up left) and profile as function as time at 500 nm and 585 nm simultaneusly (up right) in presence of TATP excess. Titration (down left) and fluorescence profile at 500 nm and 575 nm simultaneously (down right) under increasing concentrations of TATP.



There was an increase in the emission intensity band immediately after adding TATP but emission intensity was remained stable. According to the fluorescence spectra, the LOD was somewhere between 1000 and 5000 μ M with an increase of the intensity.

*Figure S282. Kinetic study at different times under visible (up) and UV (down) light in CHCl*₃:*MeOH (9:1). In each photo, left vial was contained the probe and right vial was contained the probe and an excess of TATP.*

	t=0	t=60 min	t=24 h
Visible light			
UV light (366 nm)			

*Figure S283. Titration of AR90d with an excess of TATP under visible (up) and UV (down) light in CHCl*₃:*MeOH (9:1).*

	0	250	500	1000	5000	10000	15000 2000	25000 30000 35000
Visible light								
UV light (366 nm)								5

Because of the low sensitivity the study was discontinued.